

Молекулярная организация нервной системы  
**Лекция 6: Молекулярная организация  
пресинаптического окончания**

**Казанский медицинский  
университет**

**Казань**

**Лекция**

**ноябрь 2015**

**П.Д. Брежестовский**

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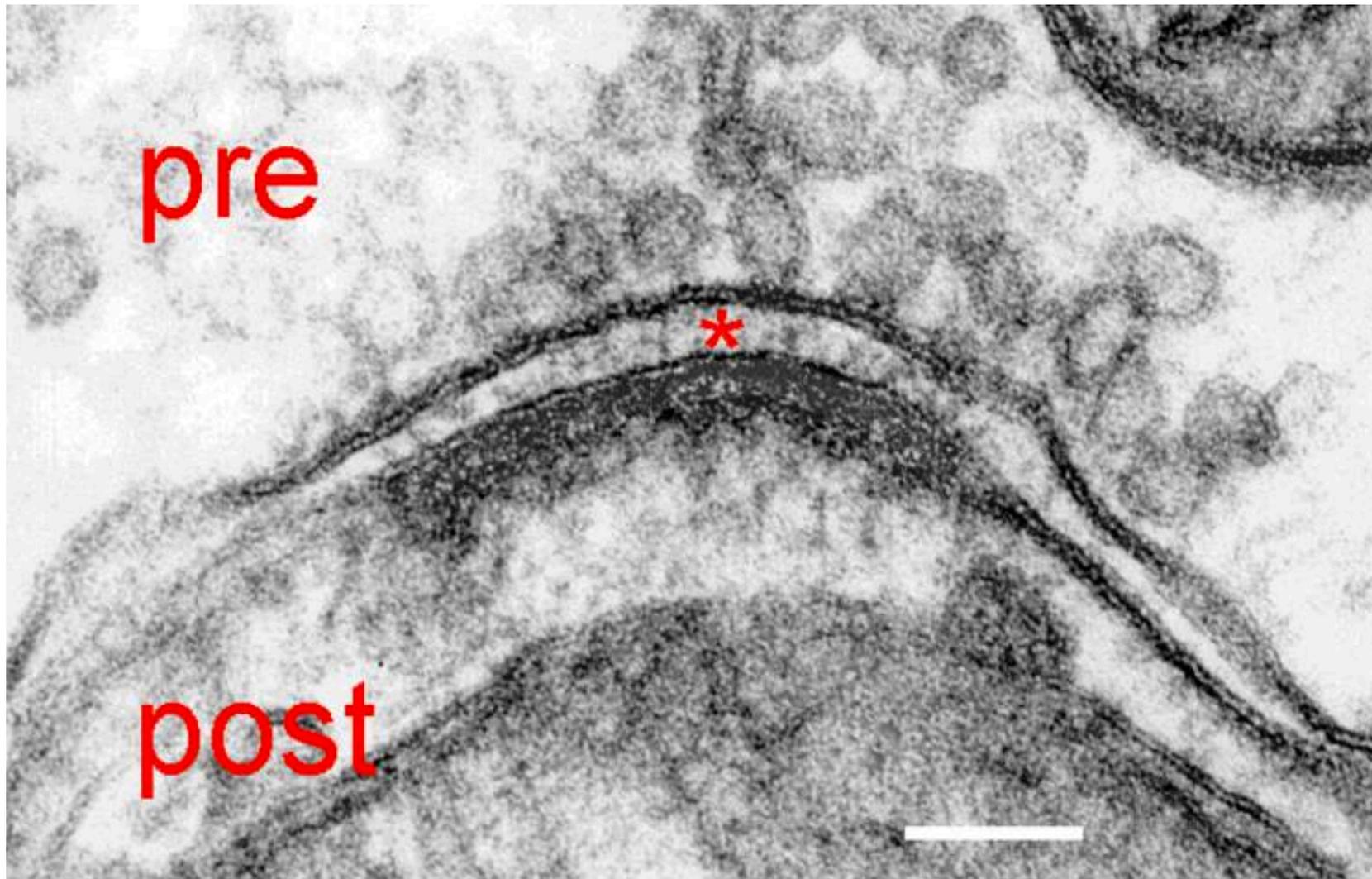
# План

- Ключевые этапы выброса нейромедиатора
- Процесс синтеза нейромедиатора
- Транспорт нейромедиатора по микротрубочкам
- Что представляет собой везикула?
- Загрузка нейромедиатора в везикулы
- Молекулярная организация основных белков, обеспечивающих слияние везикул с синаптической мембраной:
  - SNARE комплекс
  - Синаптотагмин
- Нейропатологии, связанные с нарушением выброса нейромедиаторов

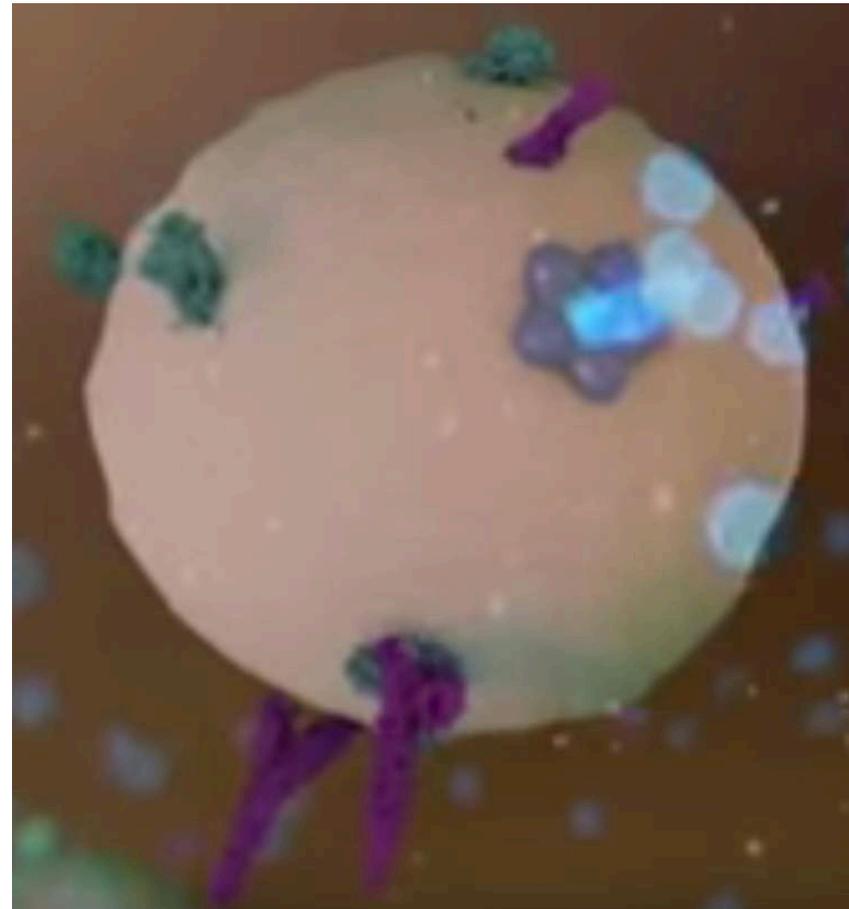
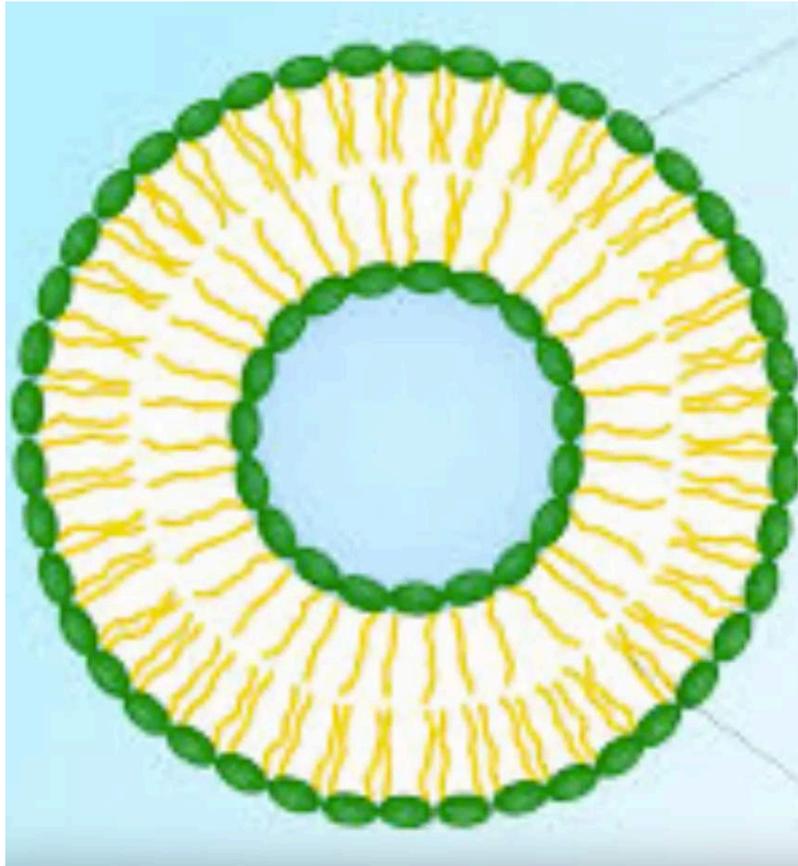
# Синаптическая щель



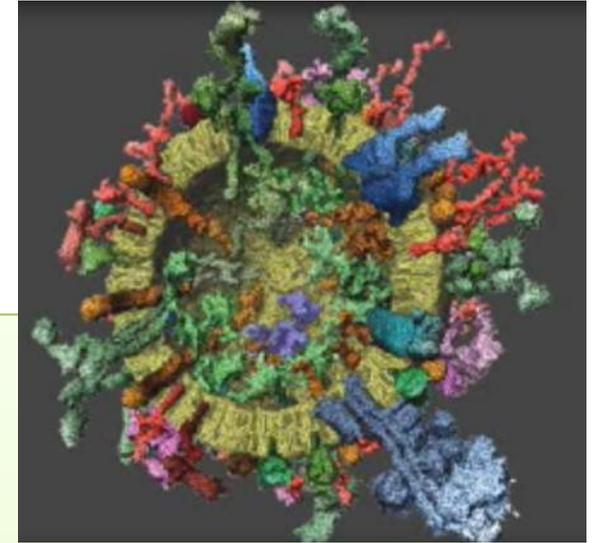
# Синаптическая щель



Что собой представляет везикула?



# Везикула содержит около 50 разных белков



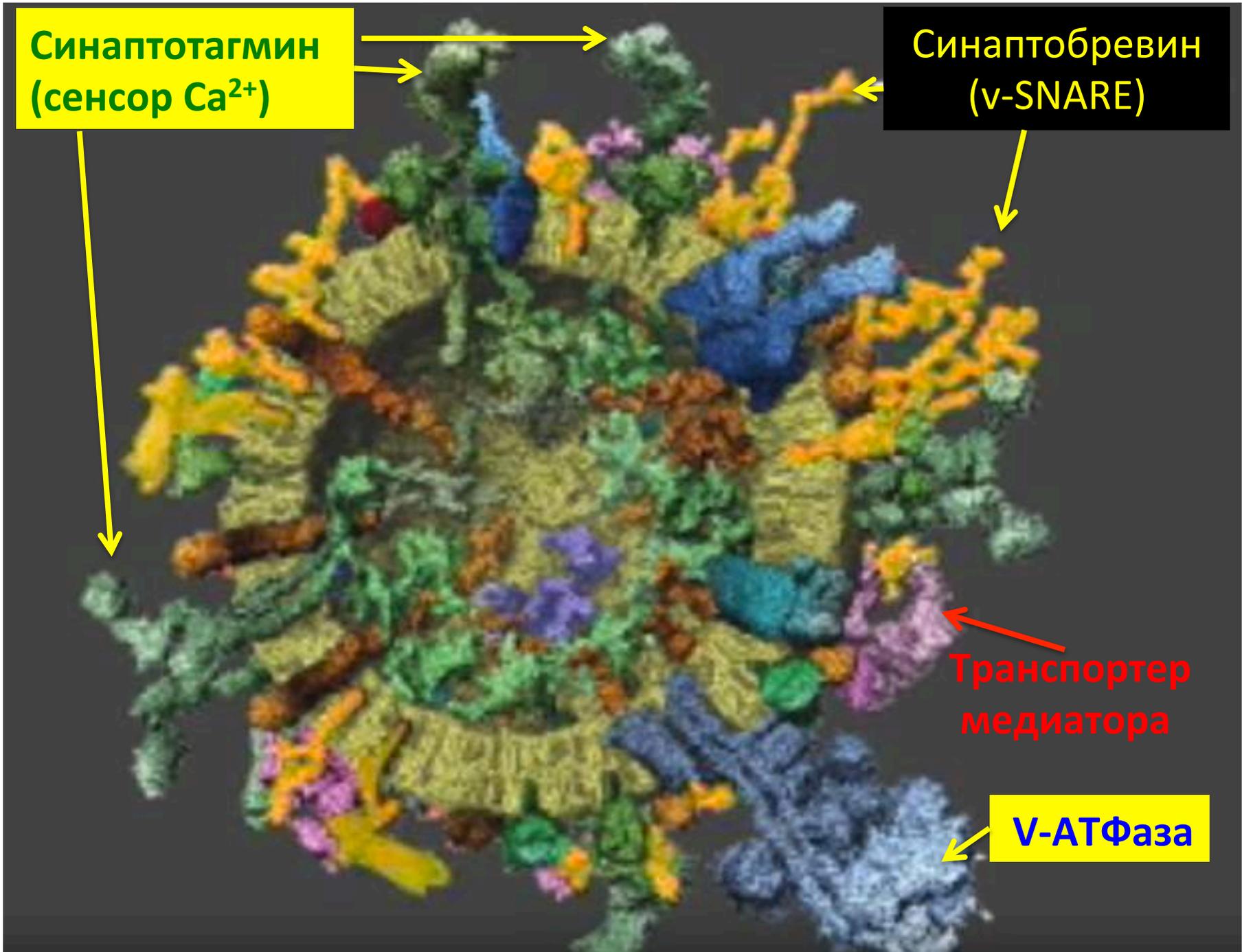
- 7 000 молекул фосфолипидов
- 5 700 молекул холестерина
- 70 копий **синаптобrevина** (v-SNARE)
- 1-2 **V-АТФазы**, которая использует энергию гидролиза АТФ для закачки водорода внутрь
- **Транспортеры** нейромедиаторов
- **Синаптотагмин** - рецептор  $\text{Ca}^{2+}$

**Синаптотагмин  
(сенсор  $\text{Ca}^{2+}$ )**

**Синаптобревин  
(v-SNARE)**

**Транспортер  
медиатора**

**V-АТФаза**



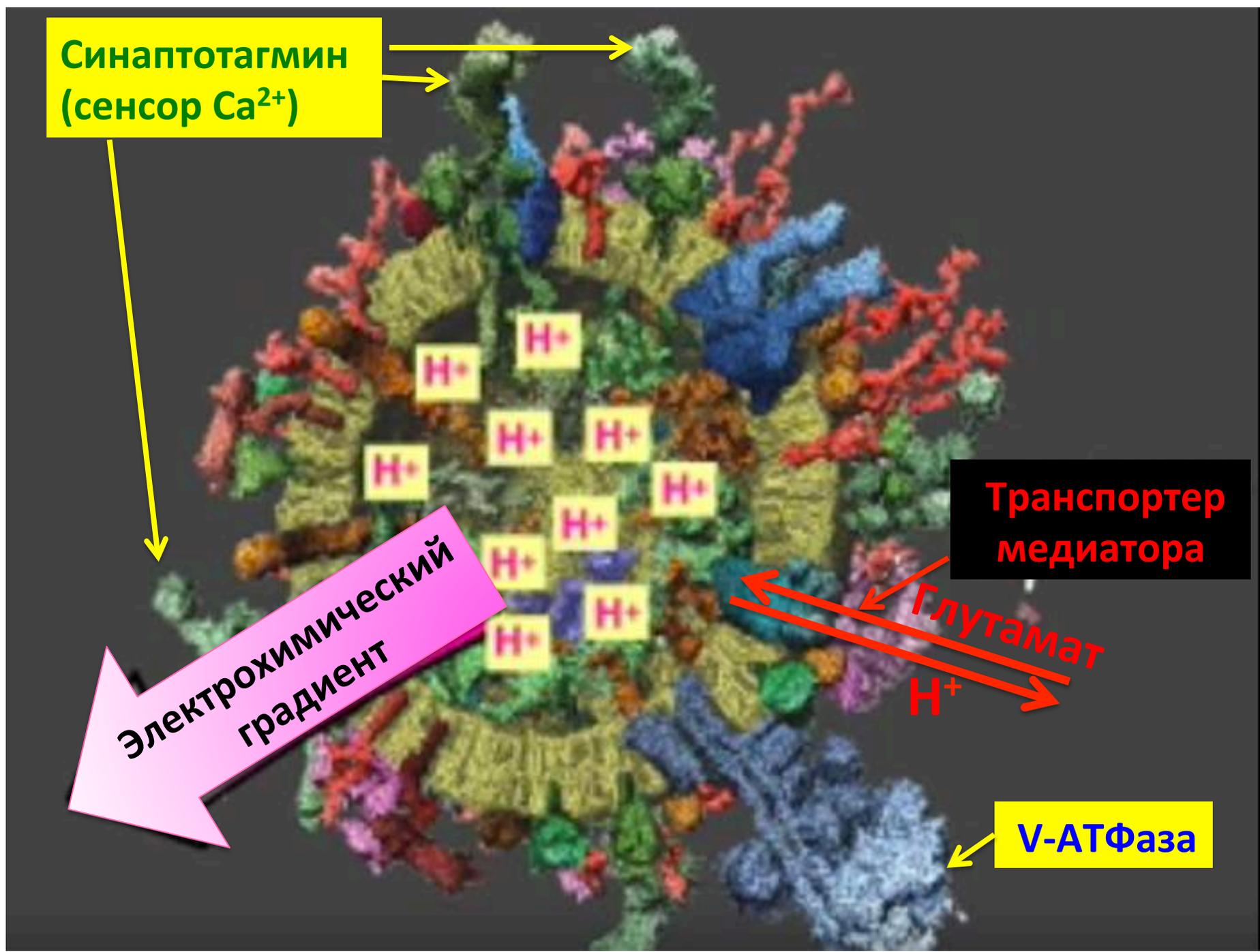
Синаптотагмин  
(сенсор  $\text{Ca}^{2+}$ )

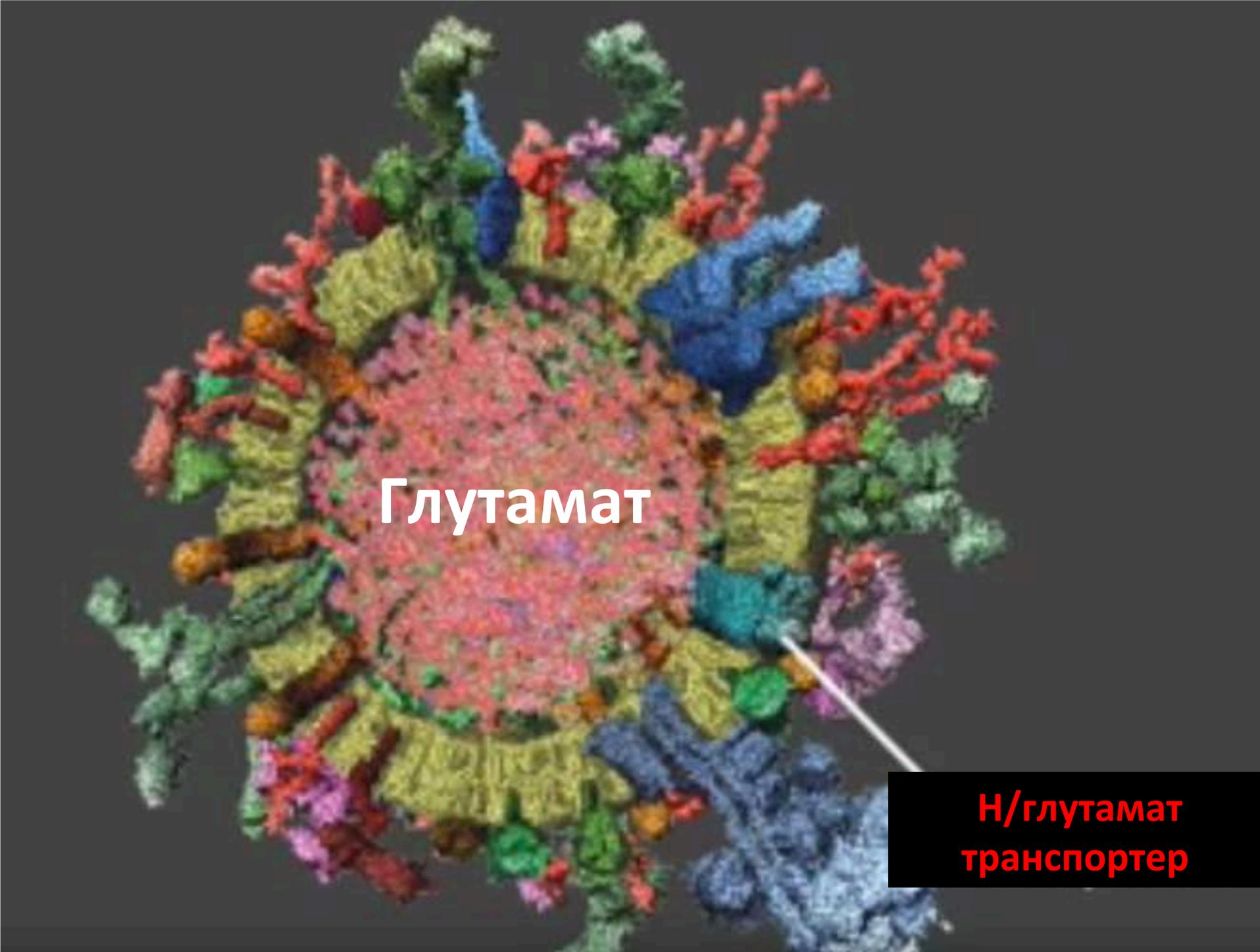
Электрохимический  
градиент

Транспортер  
медиатора

Глутамат  
 $\text{H}^+$

V-АТФаза



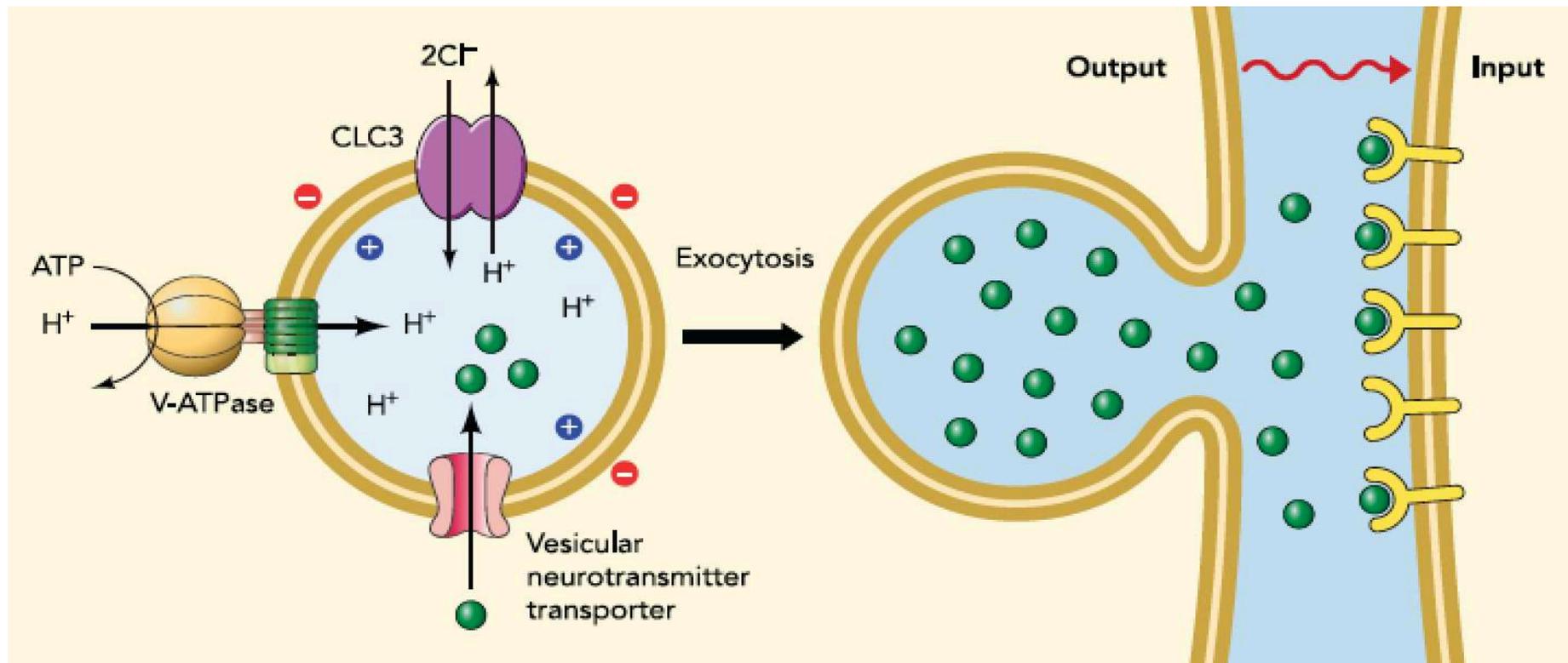


Глутамат

A 3D molecular model of a cell membrane, showing various proteins and lipids. The membrane is represented by a complex, multi-layered structure with different colors (red, green, blue, yellow, purple) representing different components. A white line points from a text box in the bottom right to a specific protein embedded in the membrane.

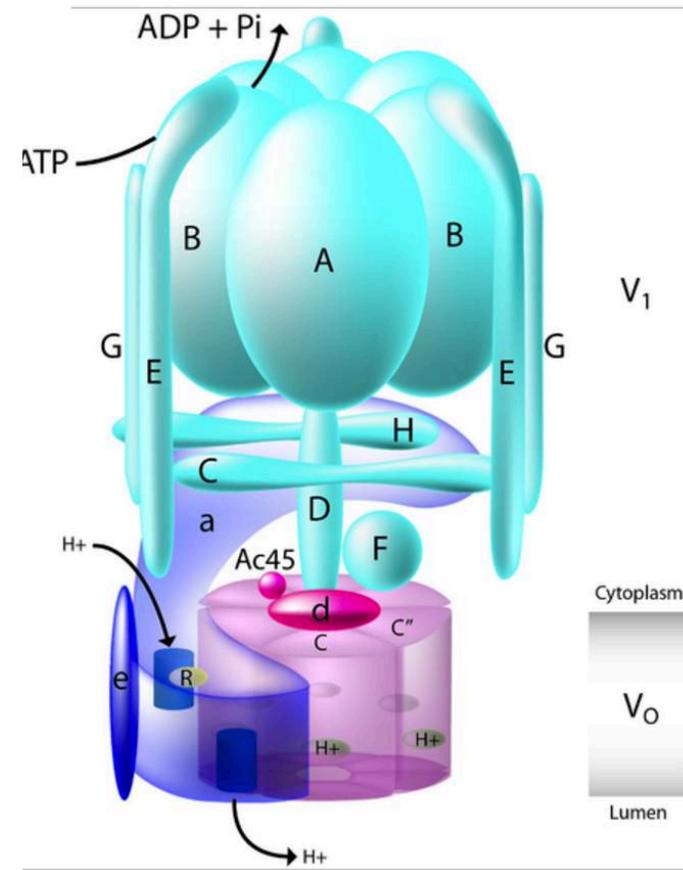
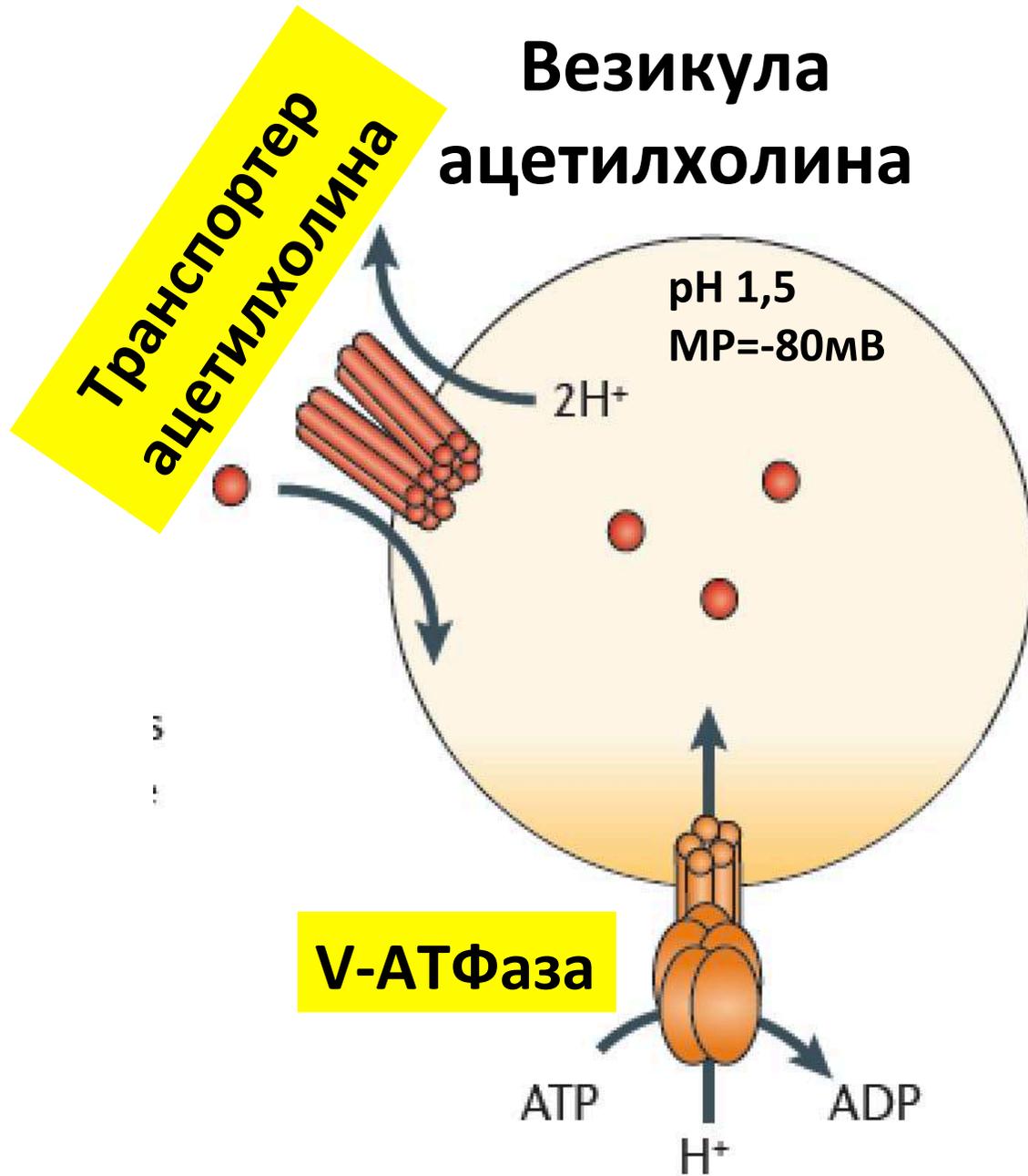
**Н/глутамат  
транспортер**

# Транспортеры нейромедиаторов



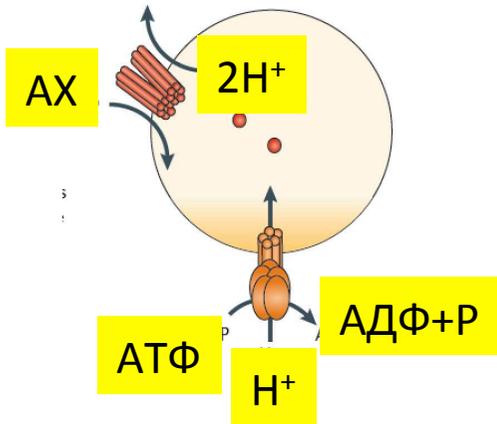
# Закачка нейромедиатора

## Везикула ацетилхолина

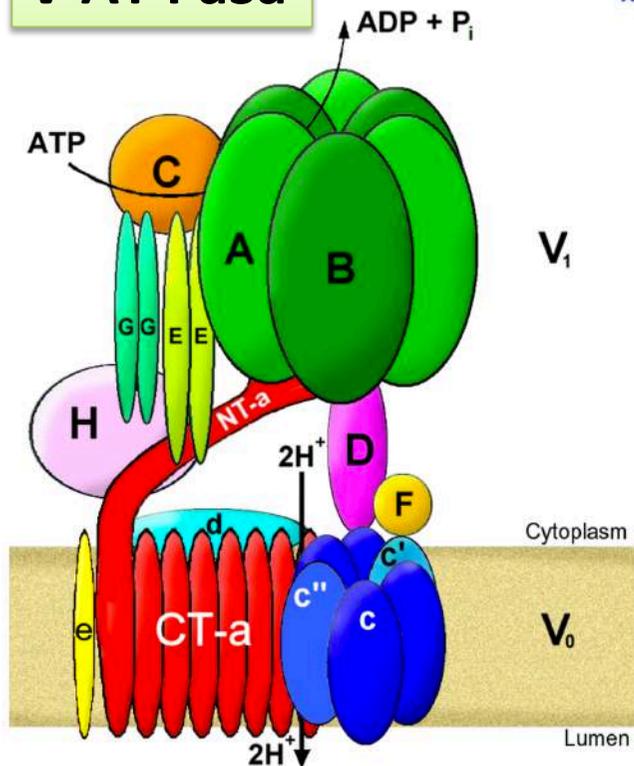


V-АТФаза

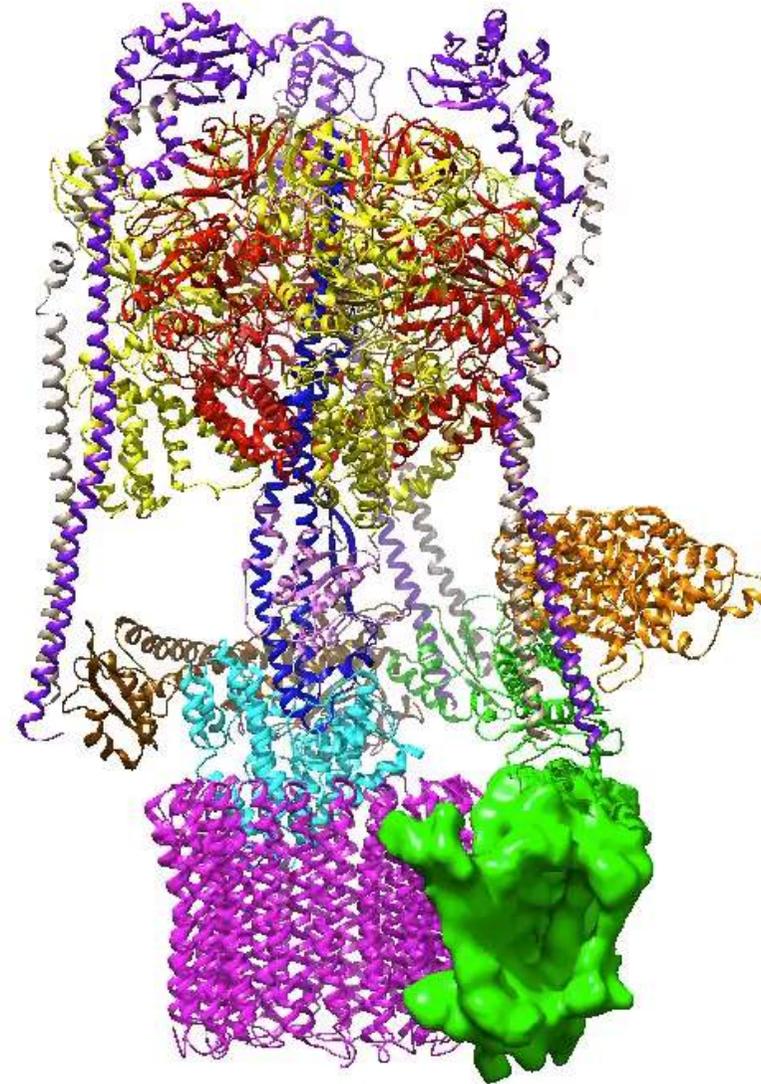
## Везикула ацетилхолина



## v-ATФаза

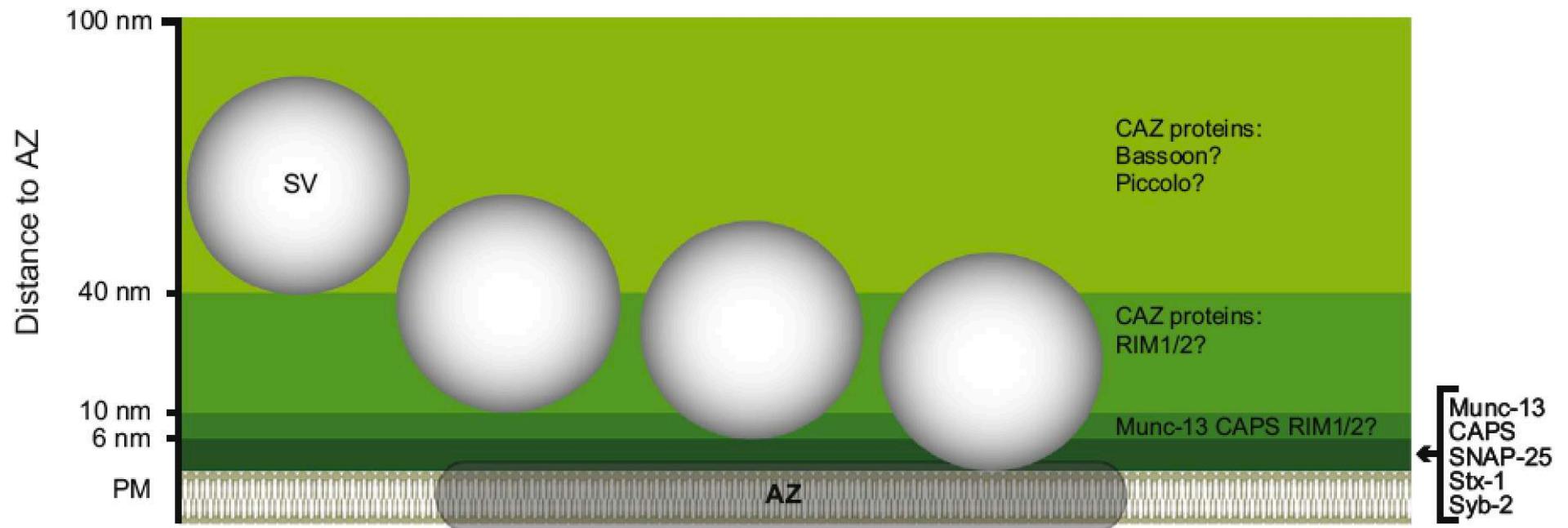


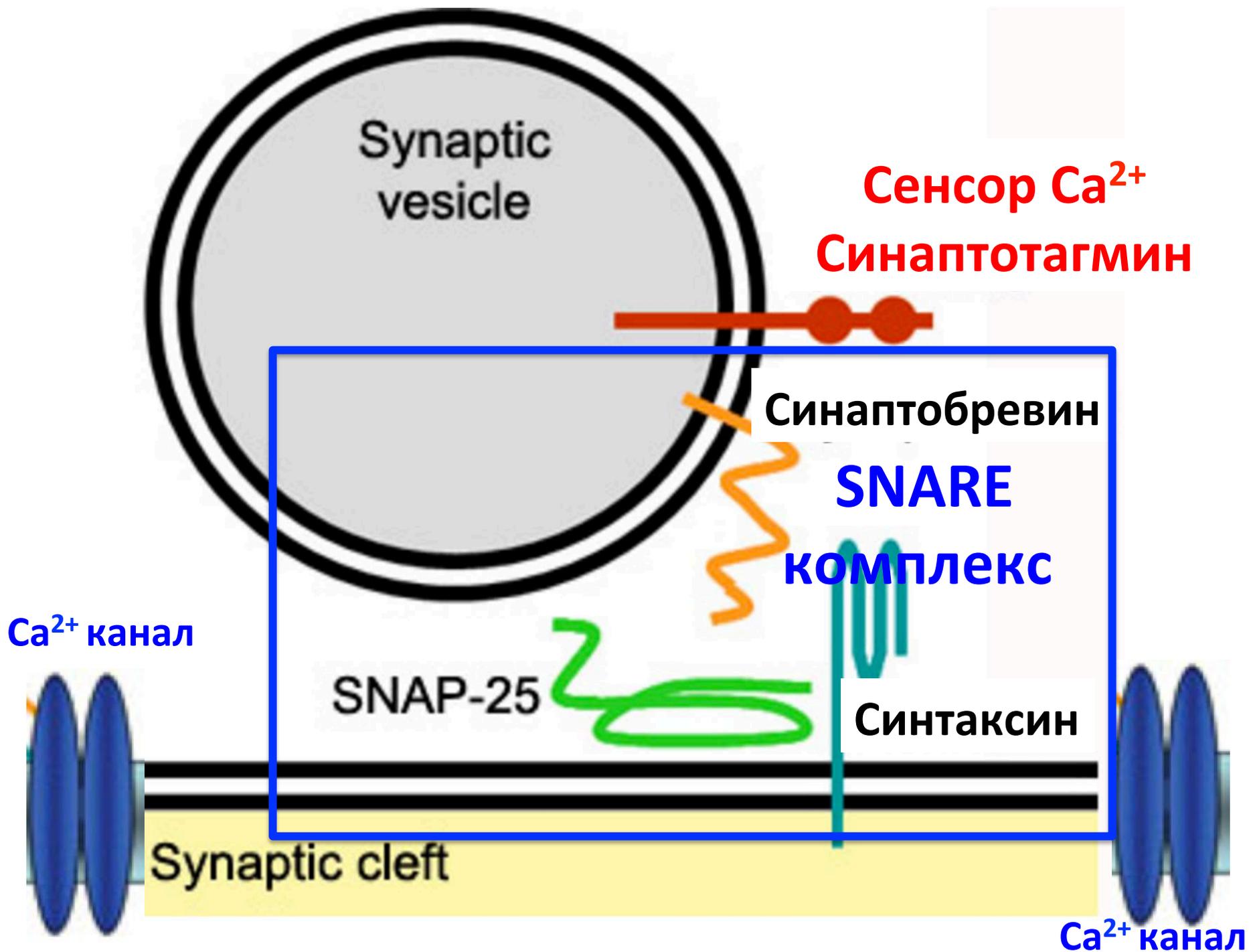
## Модель v-ATФазы



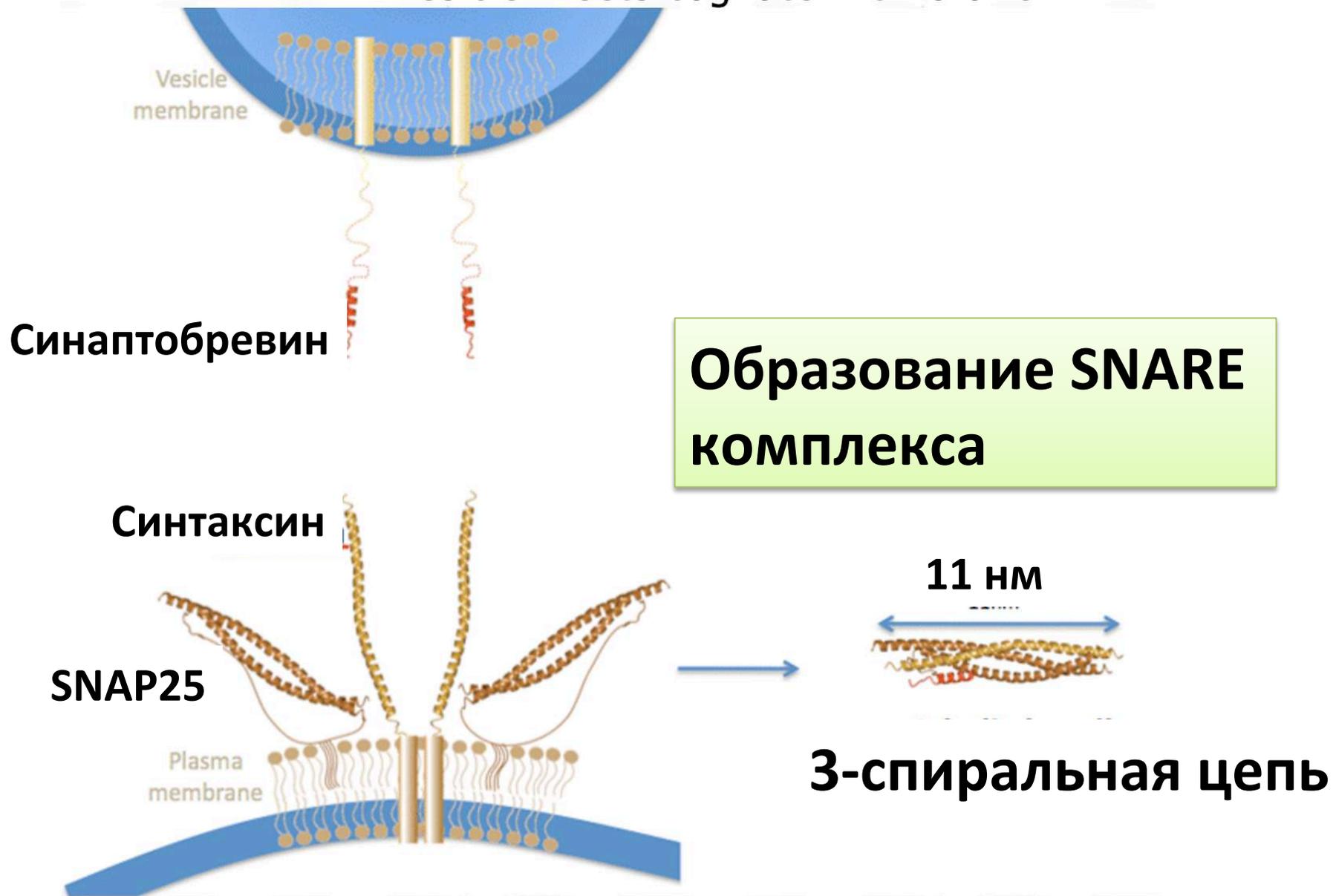
Zhao, Rubinstein et al., *Nature* 521, 241–245, 2015

# Как происходит выброс нейромедиатора?

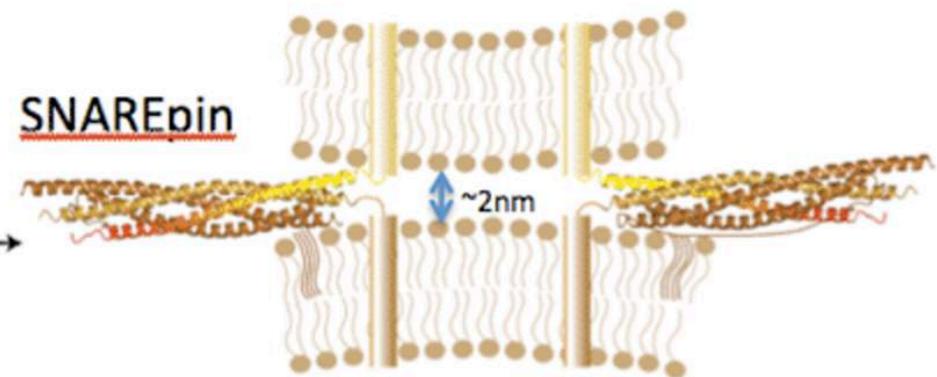
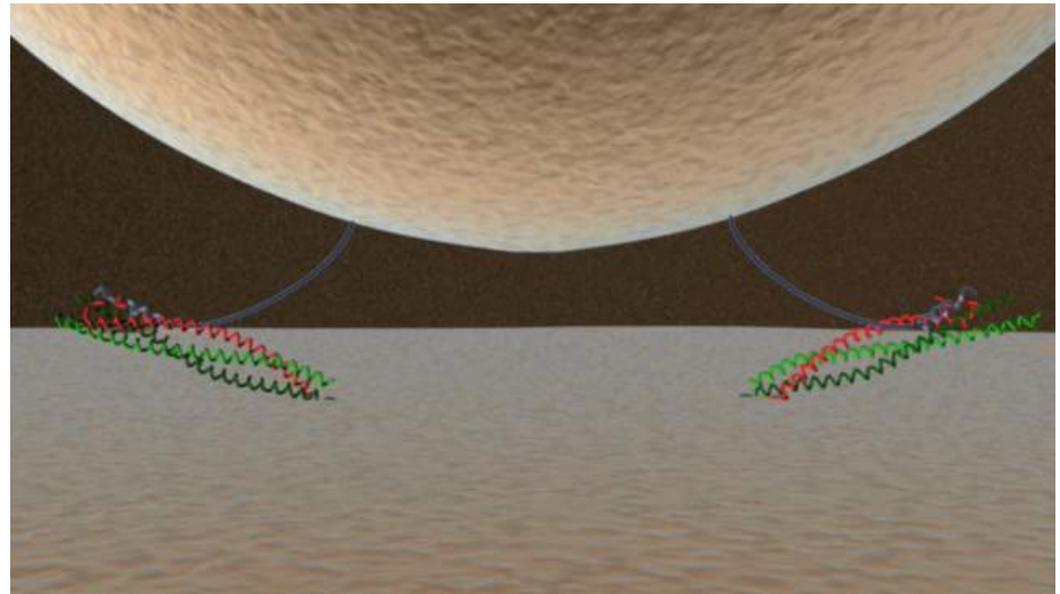
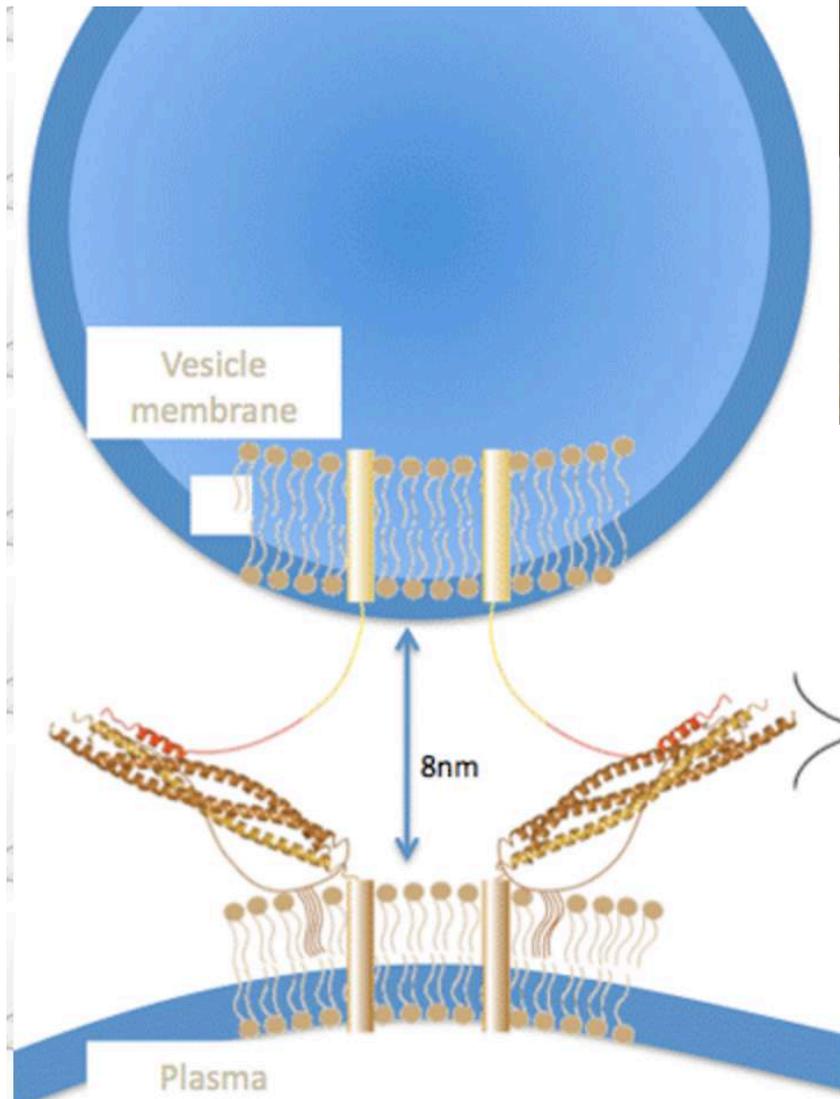




# Везикула встречается родственный участок мембраны

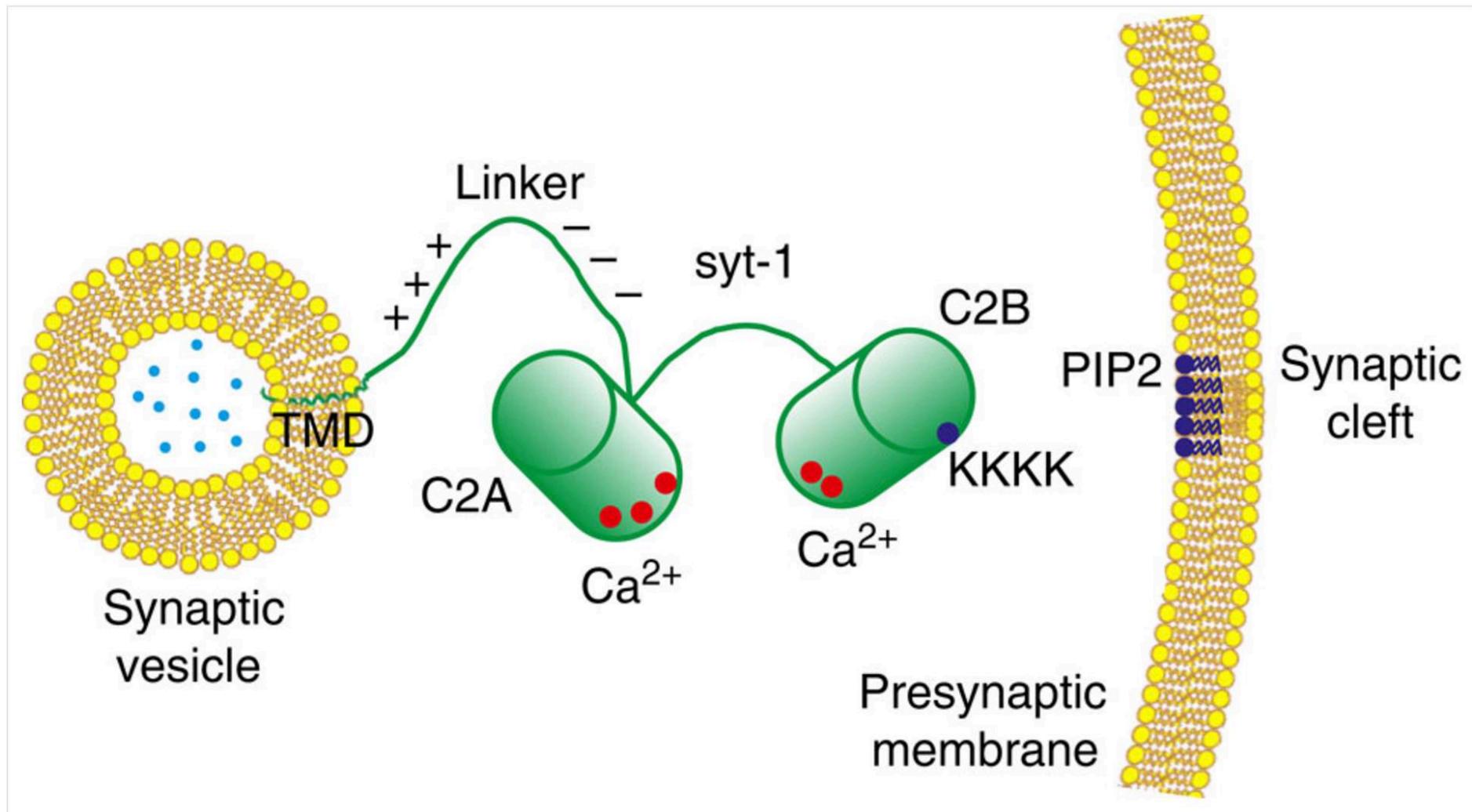


# Притягивание везикулы к мембране

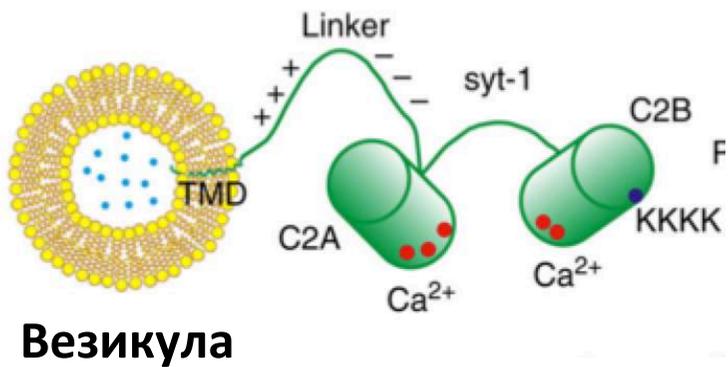


**Образование SNARE  
комплекса**

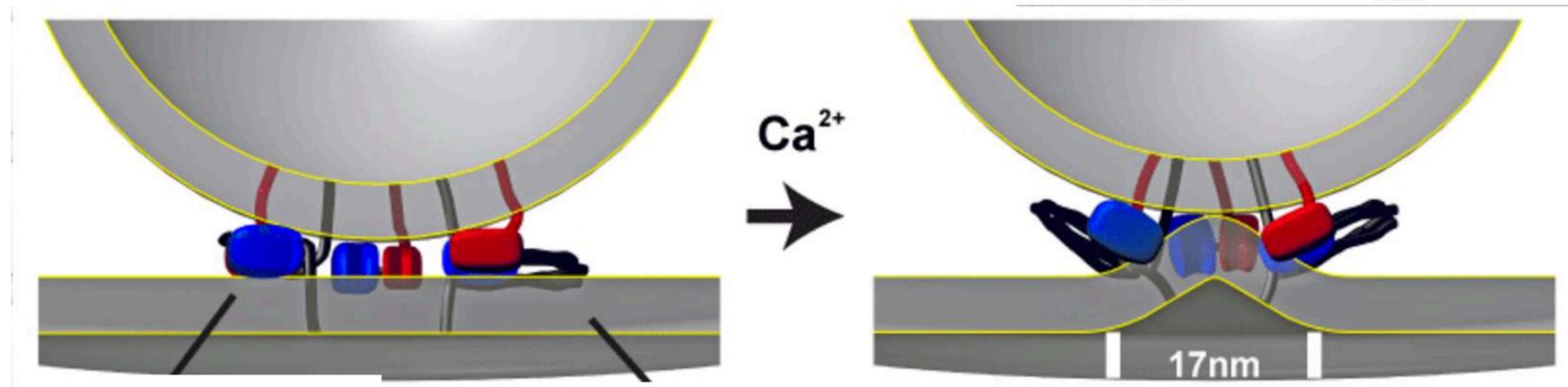
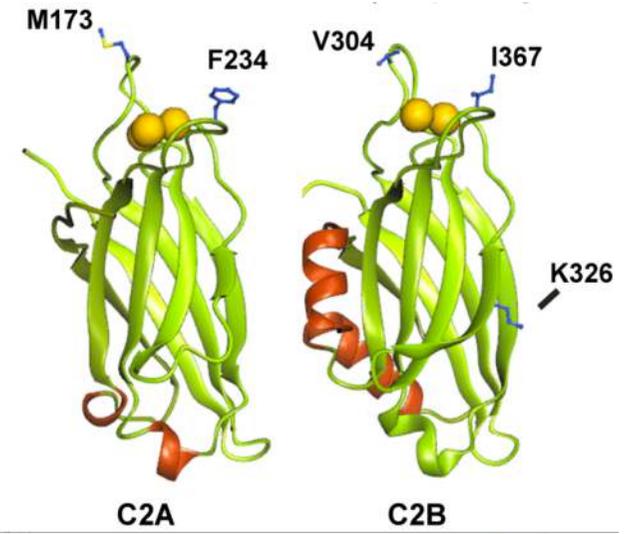
# Синаптоагмин – сенсор кальция



# Синаптоагмин

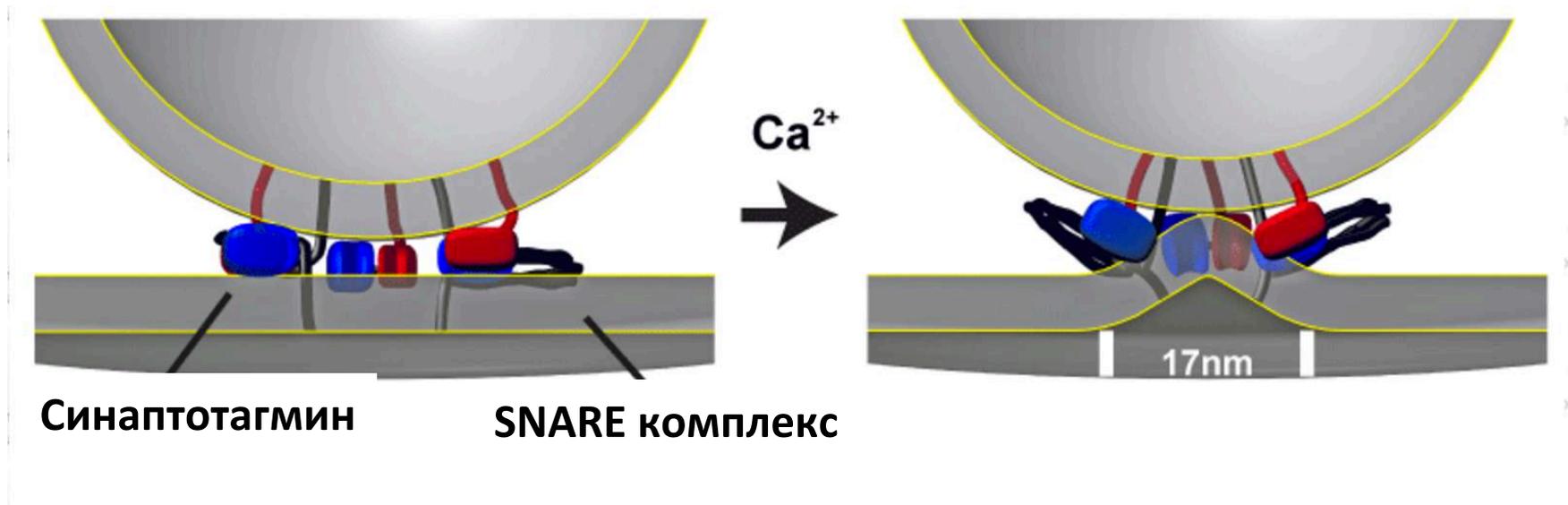


Везикула

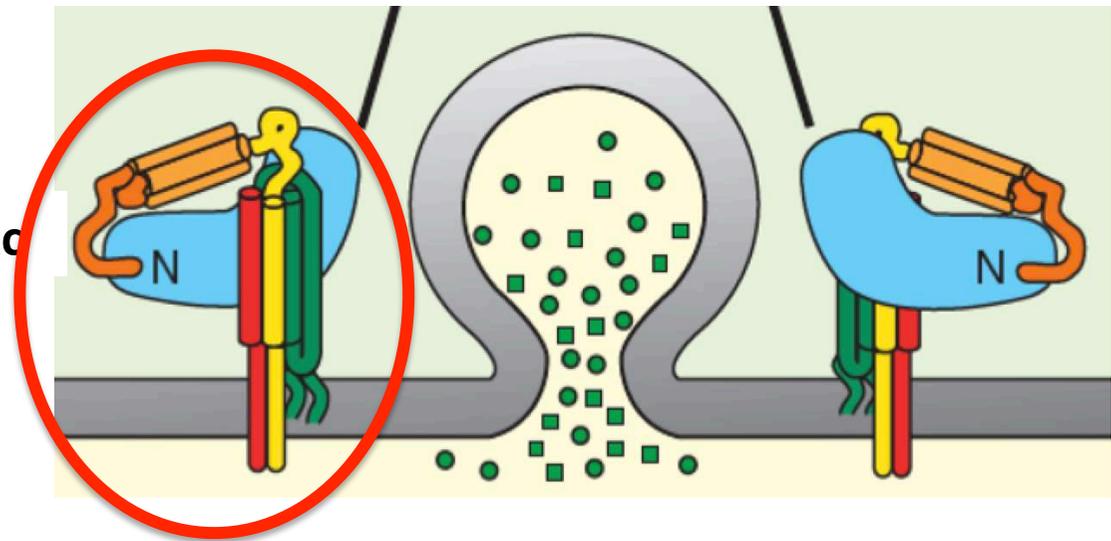


Синаптоагмин  
C2A – красный  
C2B - синий  
SNARE комплекс

# Выброс нейромедиатора

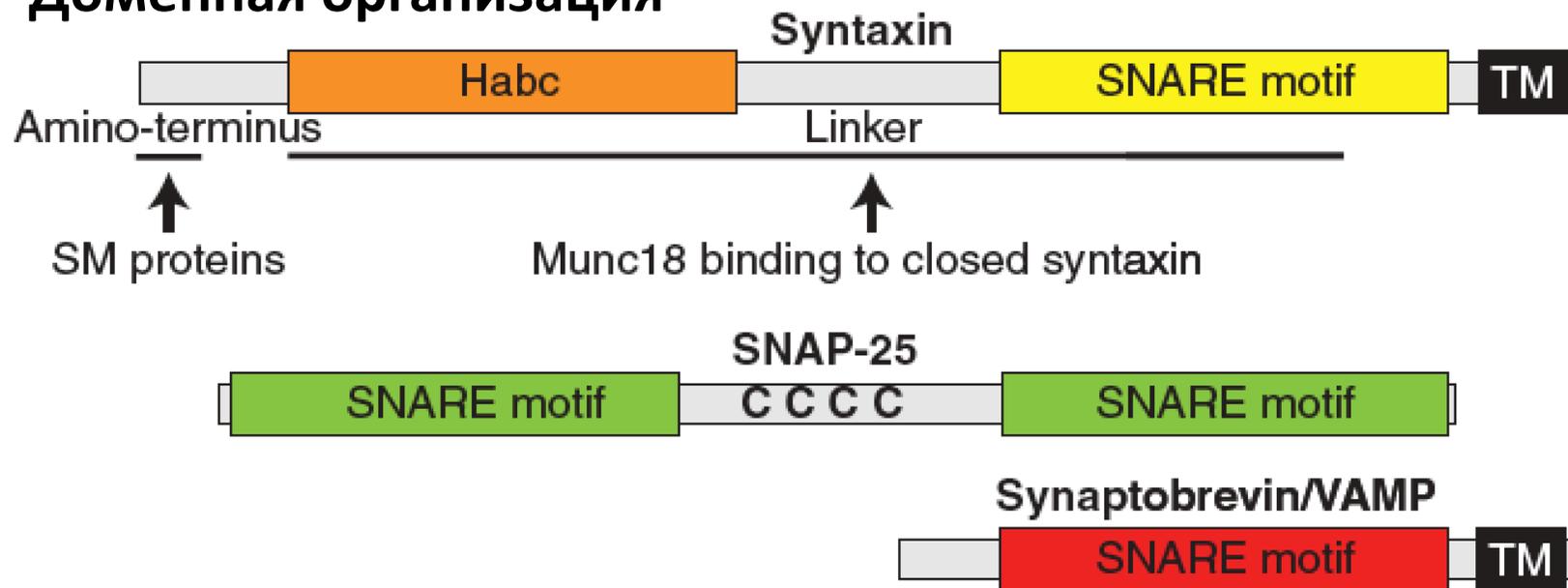


Синаптоагмин/SNARE комплекс

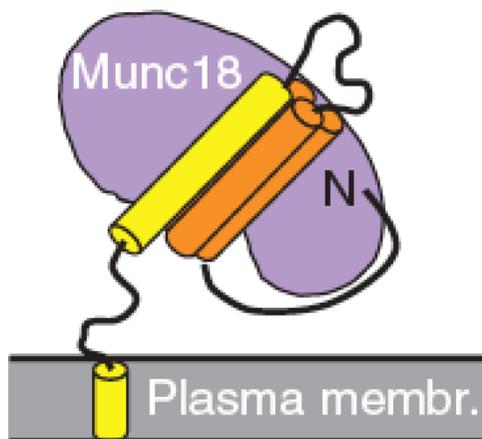


# Организация SNARE белков

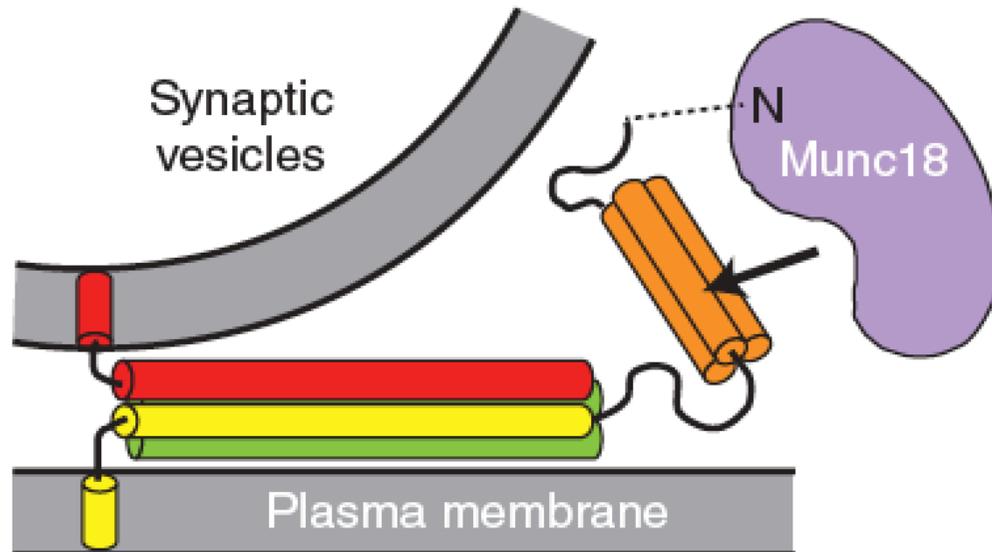
## Доменная организация



**B** *Syntaxin/Munc18 heterodimer*

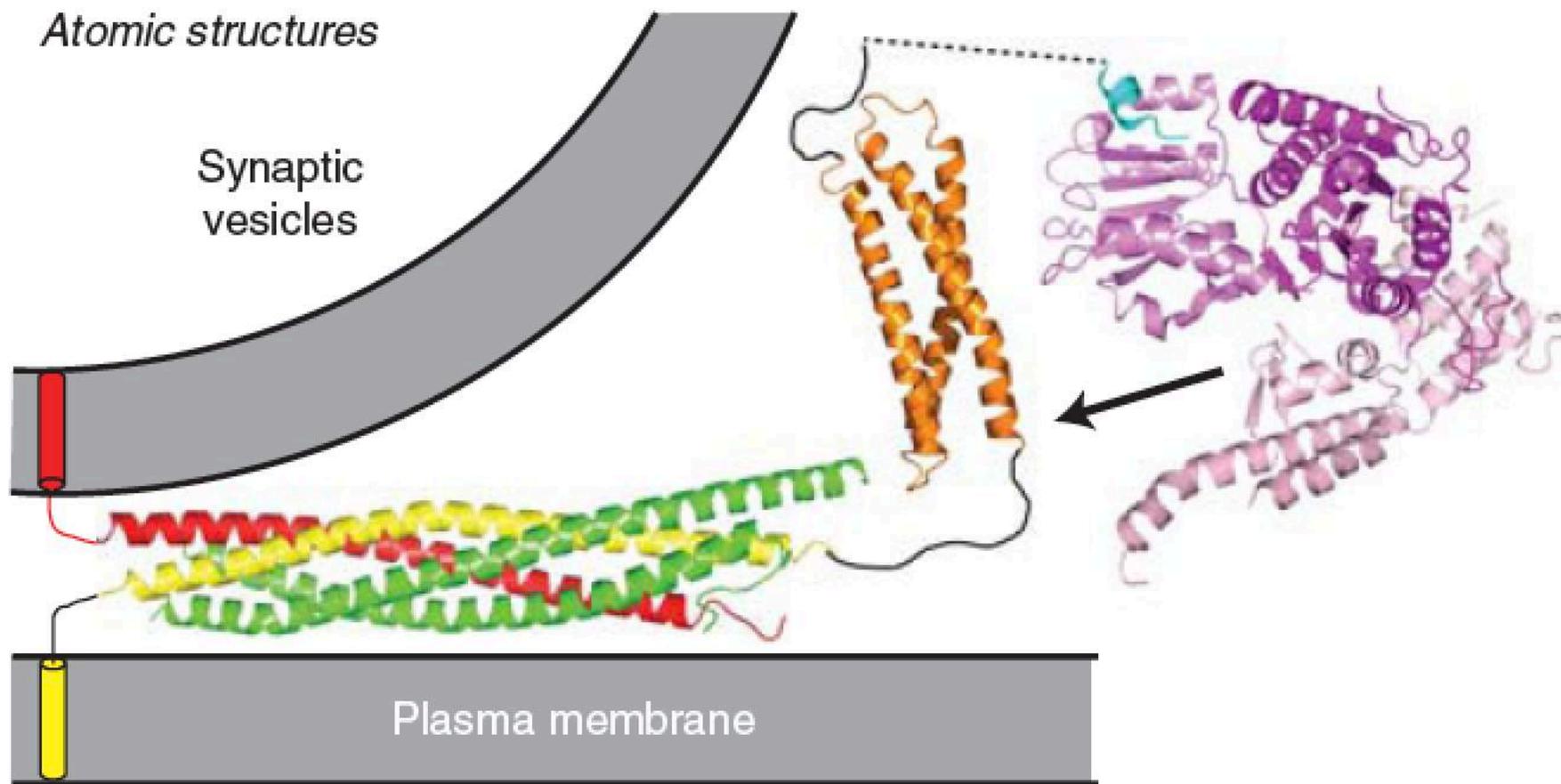


**C** *SNARE complex/Munc18 assembly*

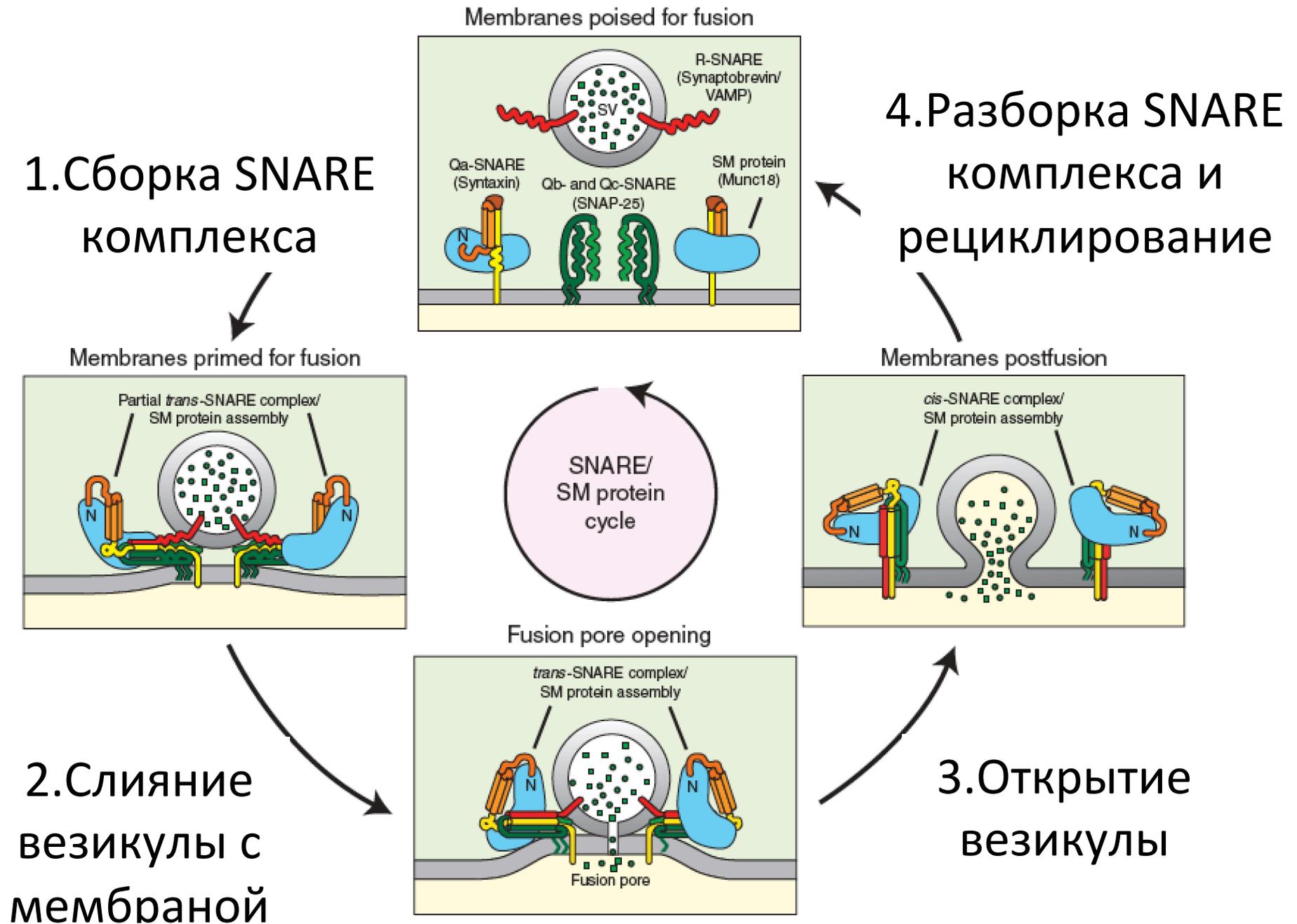


# Атомная структура SNARE комплекса

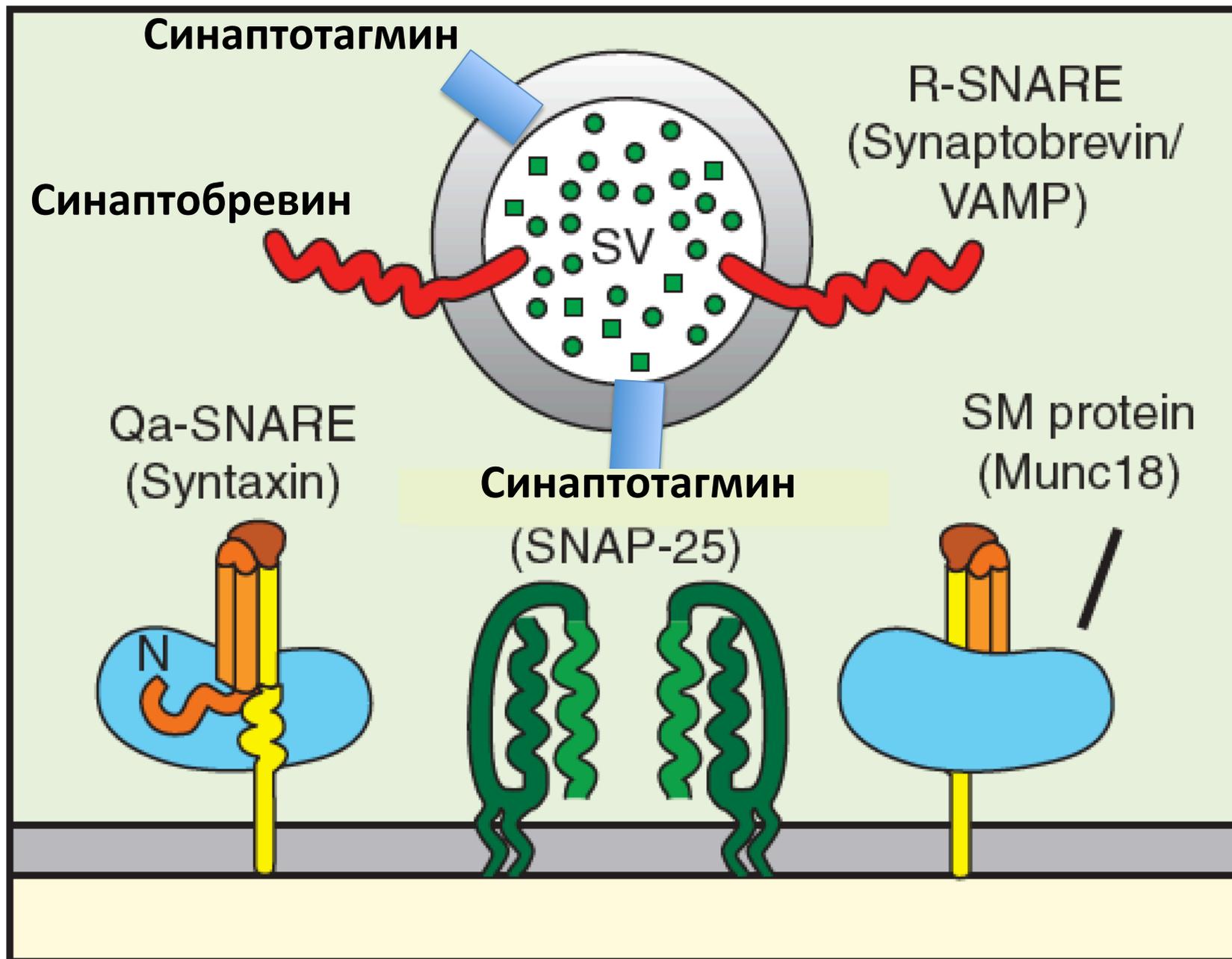
**D** *Atomic structures*



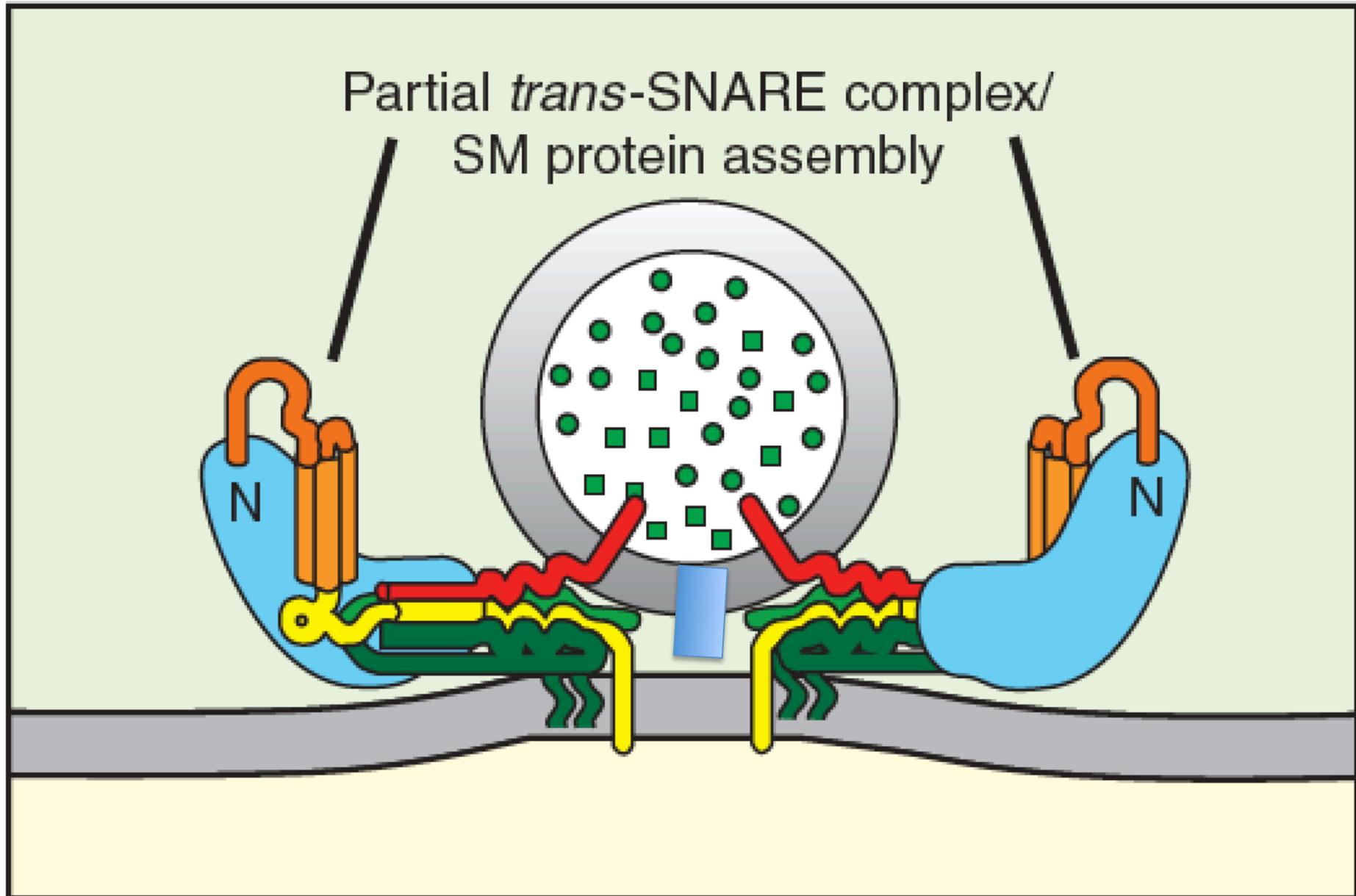
# Цикл везикул нейромедиатора



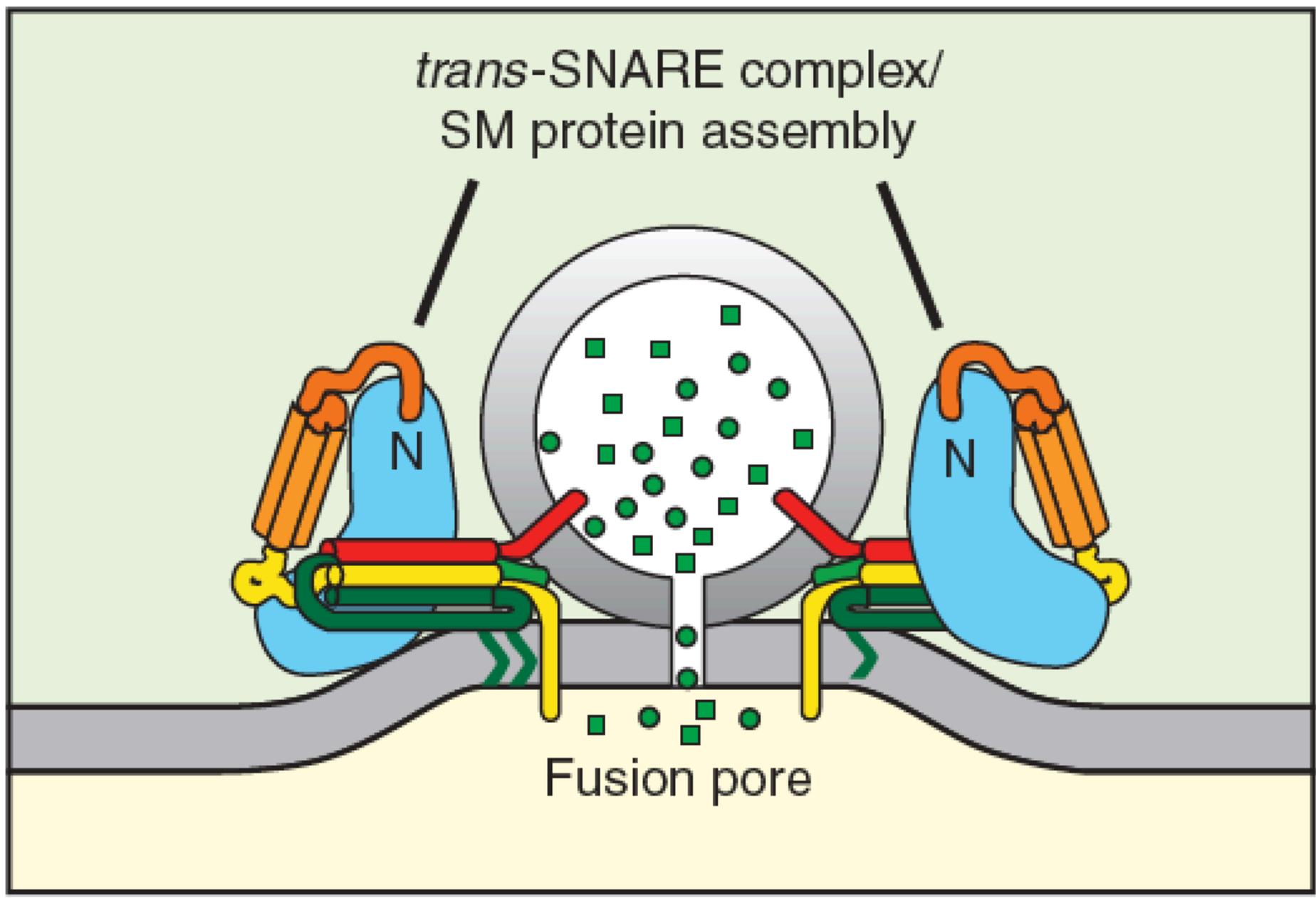
# Везикула не может слиться с мембраной



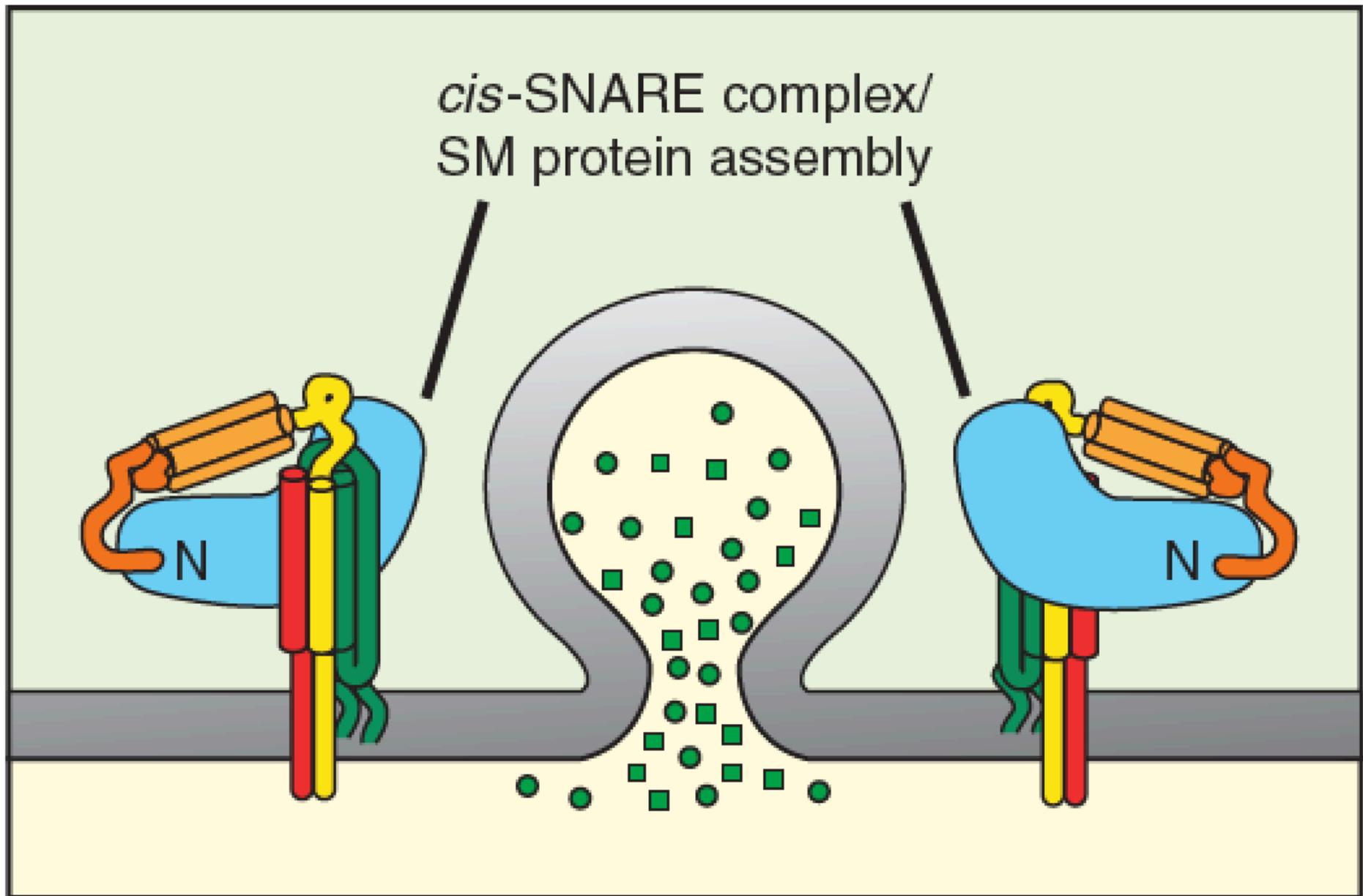
# 1. Сборка SNARE комплекса 1



# 2.Слияние везикулы с мембраной



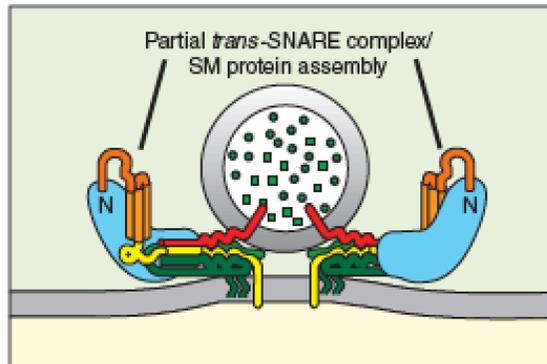
### 3.Выброс медиатора из везикулы



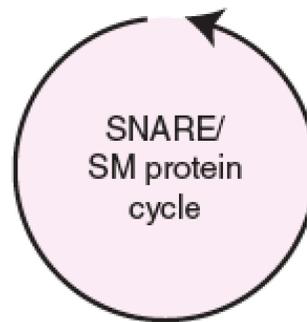
# Везикула не может слиться с мембраной

1. Сборка SNARE комплекса

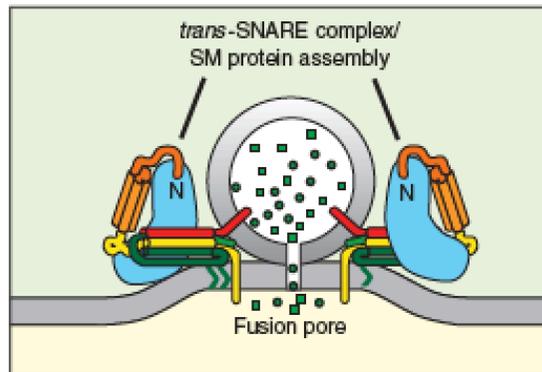
Membranes primed for fusion



2. Слияние везикулы с мембраной

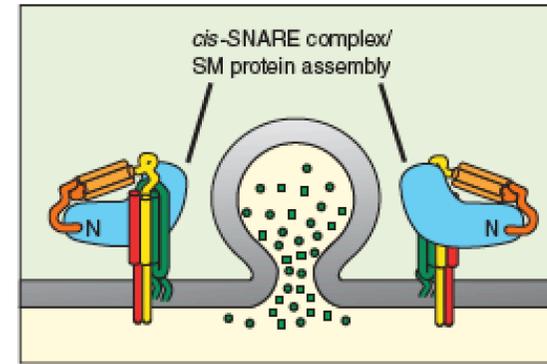


Fusion pore opening



4. Разборка SNARE комплекса и рециклирование

Membranes postfusion

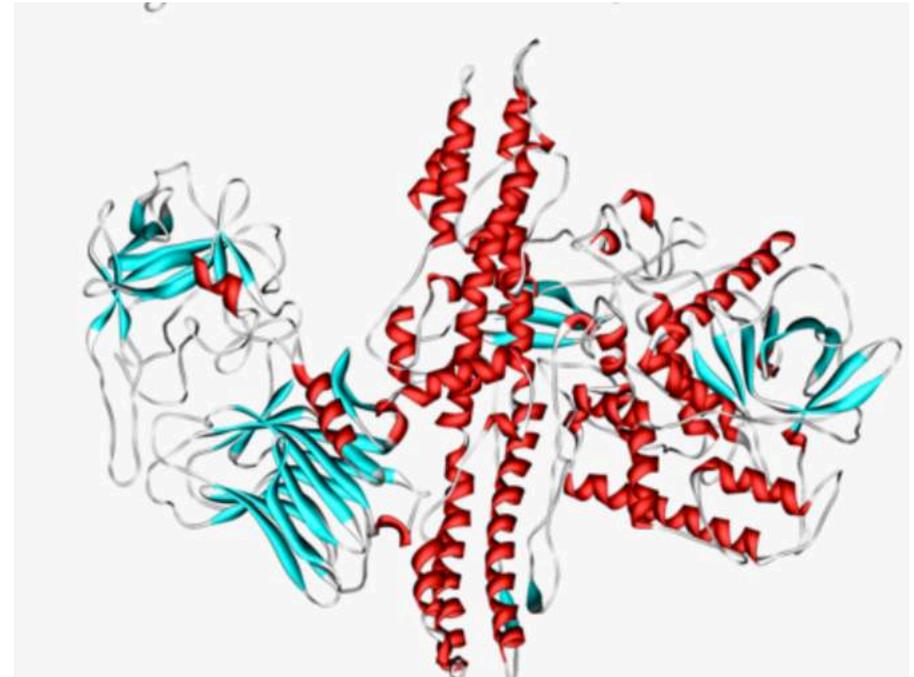


3. Открытие везикулы

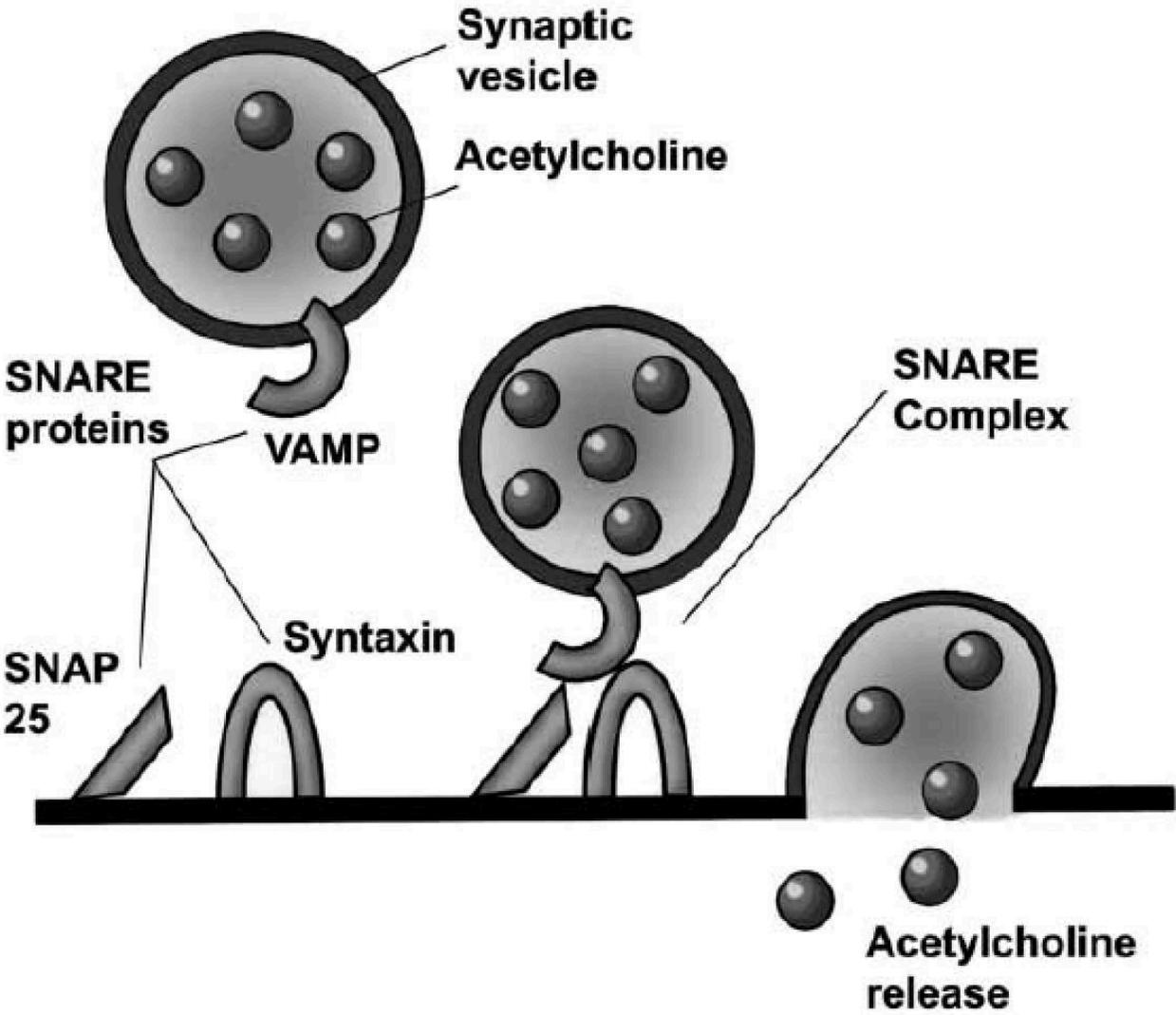
# Выброс нейромедиатора



# Ботулиновый токсин - блокатор выброса нейромедиаторов

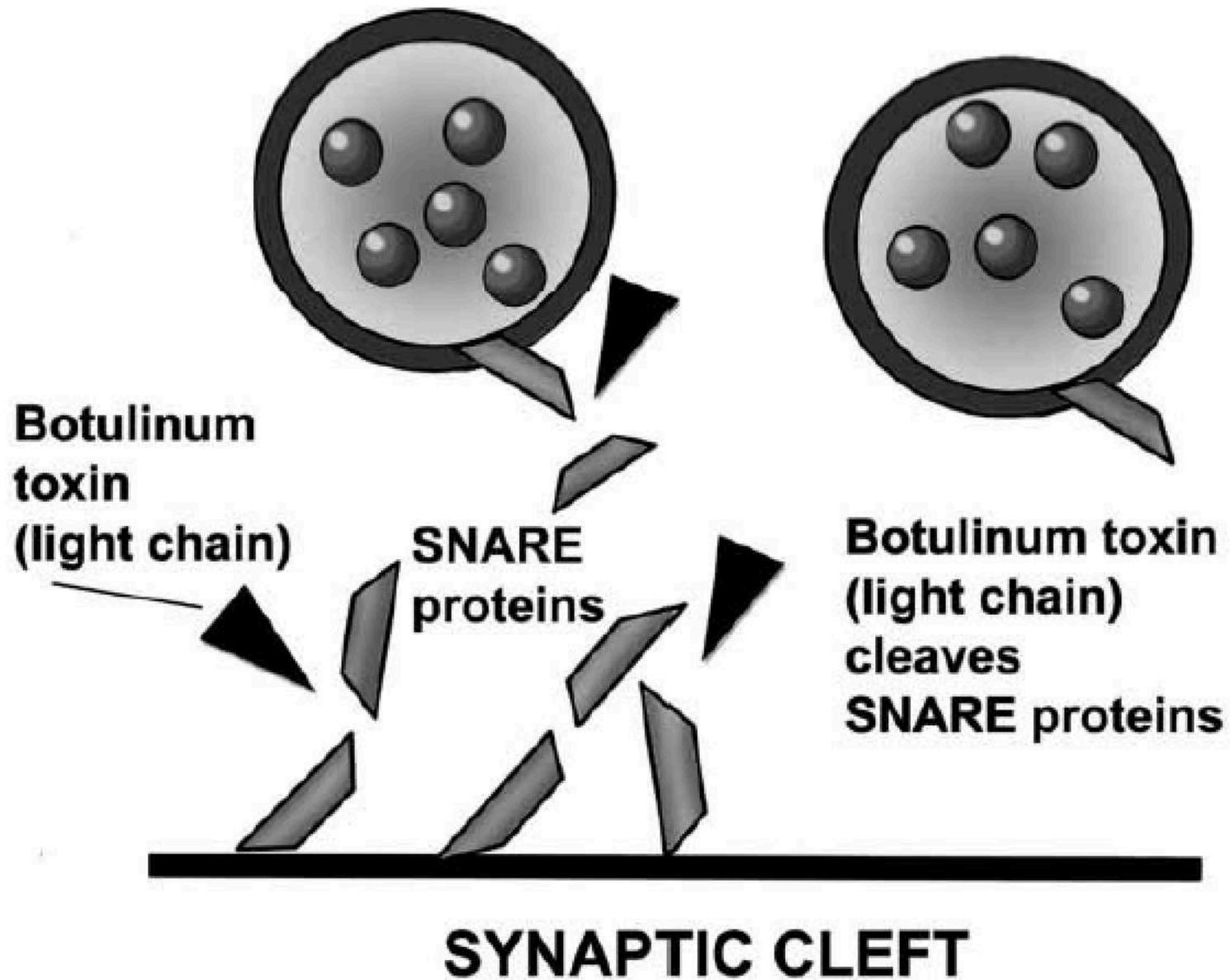


**NEURON**



**SYNAPTIC CLEFT**

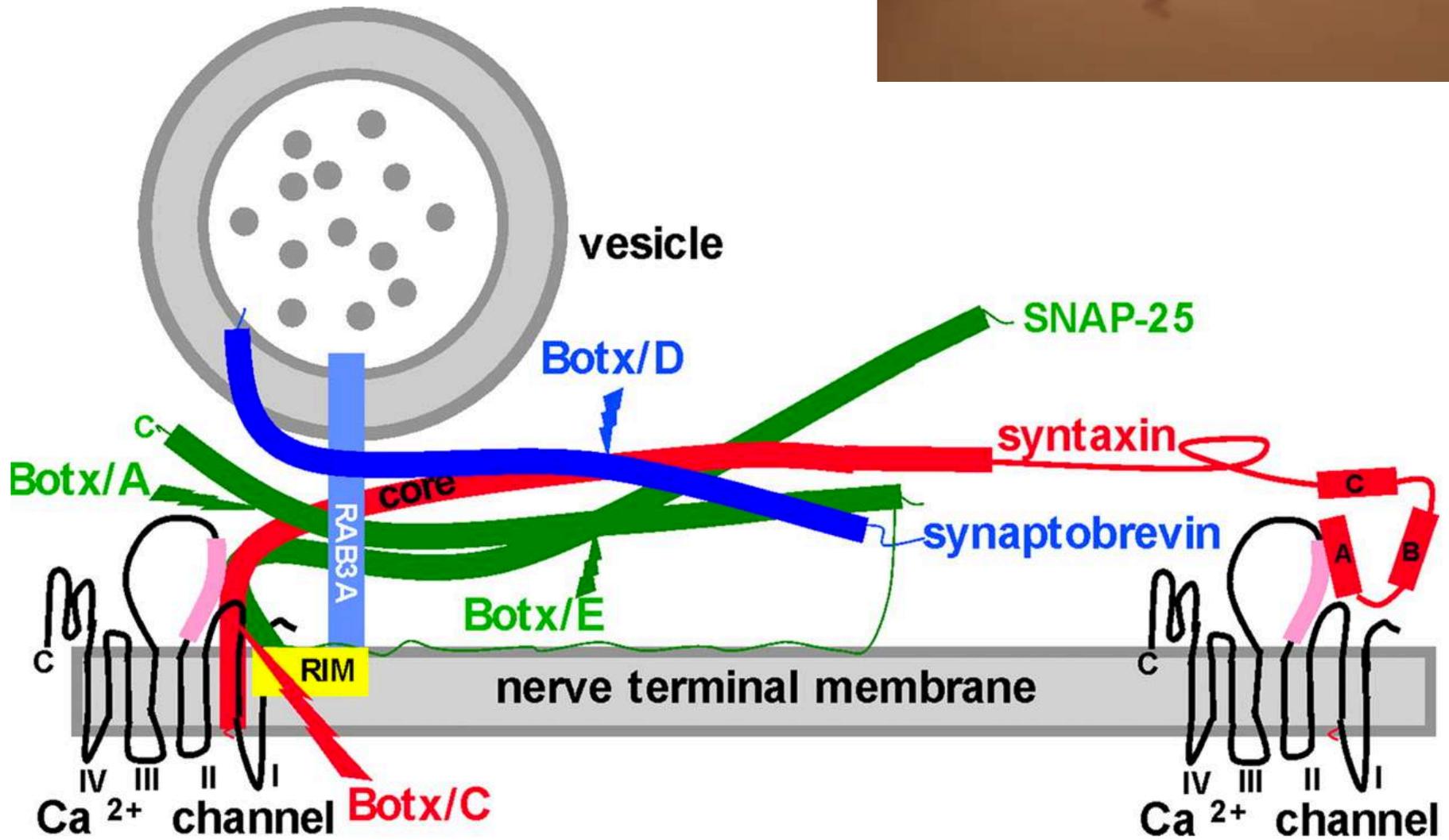
# NEURON



# Какие белки расщепляют ботулиновый токсин

Toxin	Synaptic Protein	Location
Botulinum toxins A & E	SNAP-25	Synaptic plasma membrane
Botulinum toxin C1	Syntaxins	Synaptic plasma membrane
Botulinum toxin B, D, F & G & tetanus toxin	Синаптобrevин	Synaptic vesicle

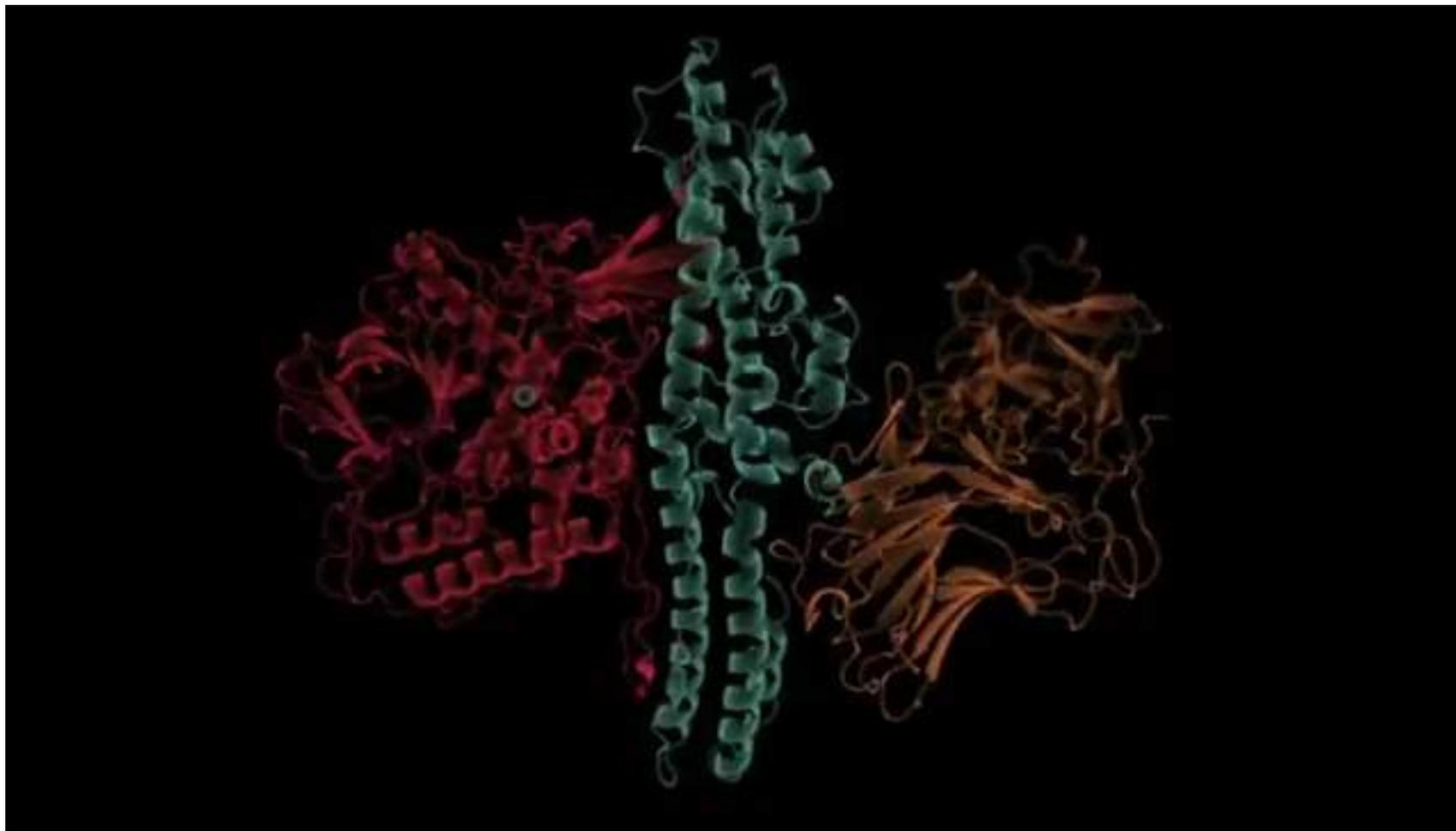
# На память



# Нейропатологии, связанные с нарушением выброса нейромедиаторов

- **Тонический блефароспазм** – неконтролируемые сокращения век глаз.
- **Цервикальная дистония** – болезненное состояние, когда мышцы шеи непроизвольно сокращаются, вызывая резкие движения головы в стороны или назад.
- **Мышечная спастичность** – измененная мышечная деятельность: частые судороги, повышенная рефлексорность сухожилий, гипертония.
- **Гиперактивность мочевого пузыря** - широко распространенное заболевание, 16 % населения США
- **Спастичность конечностей**

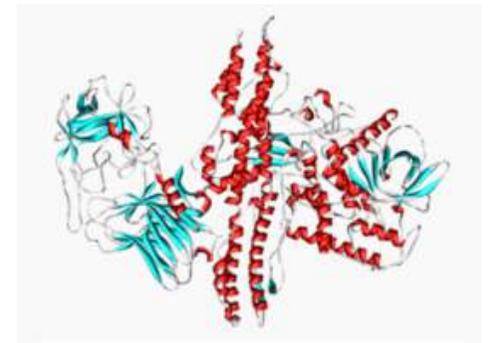
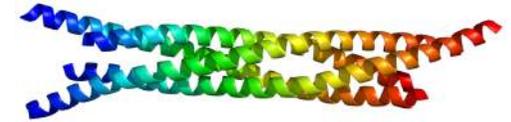
# Болезни синтаксина



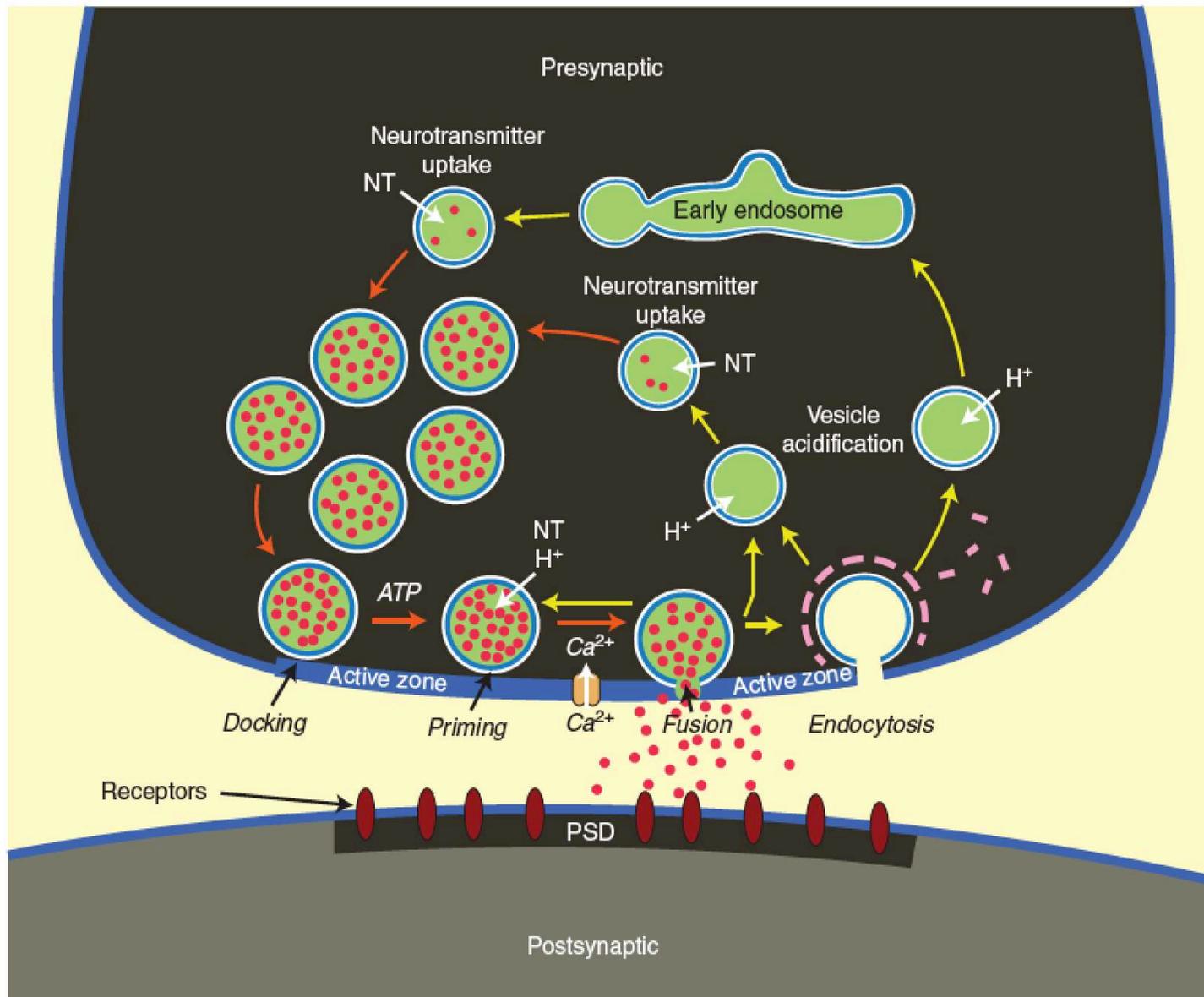
# Словарь

- **synaptobrevin/VAMP** - vesicle membrane protein, part of SNARE complex
- **Syntaxin** – presynaptic membrane protein, part of SNARE complex
- **SNAP-25** - presynaptic membrane protein, part of SNARE complex
- **synaptogagmin** – receptor of  $\text{Ca}^{2+}$
- **RIMs** - Rab3-interacting molecules;
- **RIM-BPs** - RIM-binding proteins;
- **Rab3** - G protein family: four members, Rab3A, 3B, 3C, 3D
- **Rab3A** - regulates  $\text{Ca}^{2+}$ -dependent neurotransmitter release

- **SNAP25** (от англ. synaptosomal-associated protein, **25-kD**) — мембранный белок, компонент белкового комплекса SNARE, осуществляющего стыковку синаптической везикулы с пресинаптической мембраной нейрона и их слияние с последующим высвобождением нейромедиатора.
- **Ботулиновый токсин** типа А, С и Е приводят к расщеплению SNAP25, что вызывает паралич.



# На память



# На память:

## Основные этапы передачи синаптического сигнала

- Синапс – специализированный контакт между клетками, обеспечивающий передачу нервного импульса с пресинаптического окончания на постсинаптическую мембрану
- Нервный импульс (потенциал действия) стимулирует выброс химического вещества (нейромедиатора) в синаптическую щель
- Нейромедиатор пересекает синаптическую щель и взаимодействует с рецепторами на постсинаптической мембране

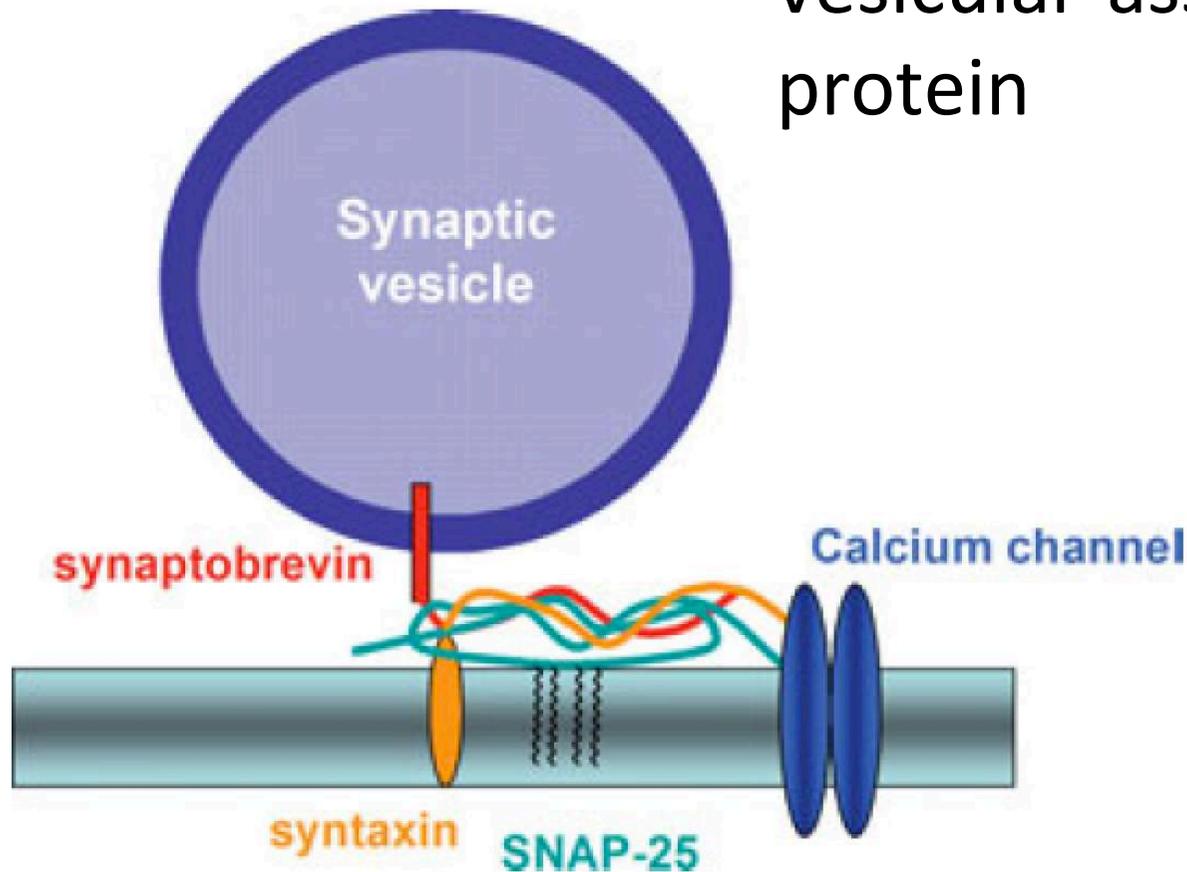
# На память

- Везикула содержит:
  - фосфолипиды и холестерин ( $\approx 13\ 000$  молекул)
  - **Синаптобrevин** ( $\approx 70$  копий): член SNARE комплекса
  - **Синаптотагмин** - рецептор  $\text{Ca}^{2+}$
  - **V-АТФазы**: используют энергию гидролиза АТФ для закачки водорода внутрь
  - **Транспортеры** нейромедиаторов
- Неропатологии
- SNARE комплекс:
  - Синтаксин/SNAP25/Синаптобrevин



# SNAP-25 - synaptosomal-associated protein 25 kDa

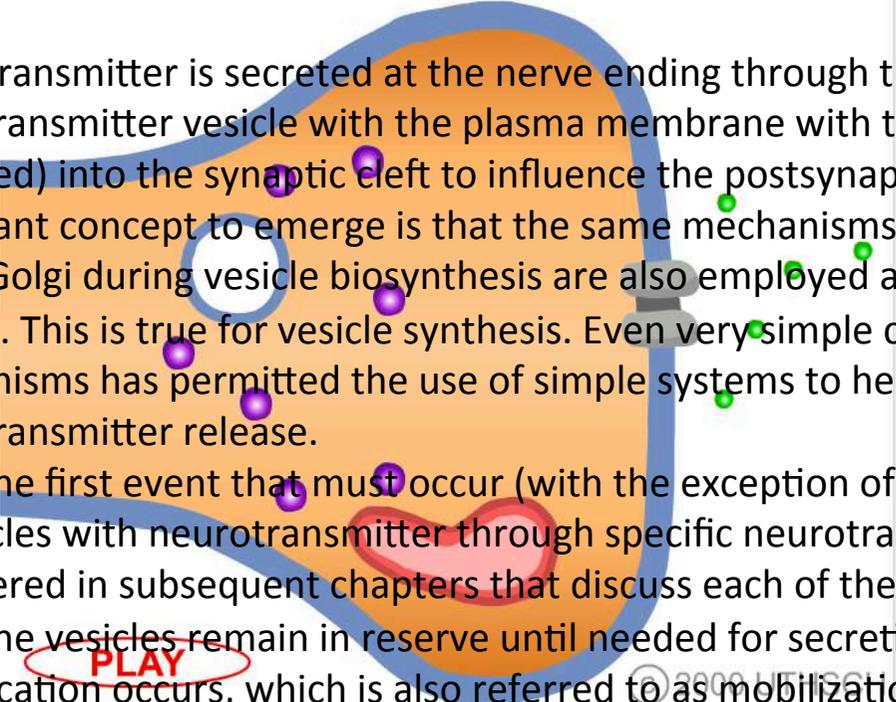
- SNAP-25 - synaptosomal-associated protein 25 kDa
- VAMP/synaptobrevin - vesicular-associated membrane protein



# Словарик

- SNAP-25 - synaptosomal-associated protein 25 kDa
- VAMP/synaptobrevin - vesicular-associated membrane protein
- **SNARE** - Soluble NSF Attachment Protein) **RE**септор") суперсемейство из > 60 белков. Роль – слияние везикул с мембраной (лизосомы, пресинаптическая мембрана)





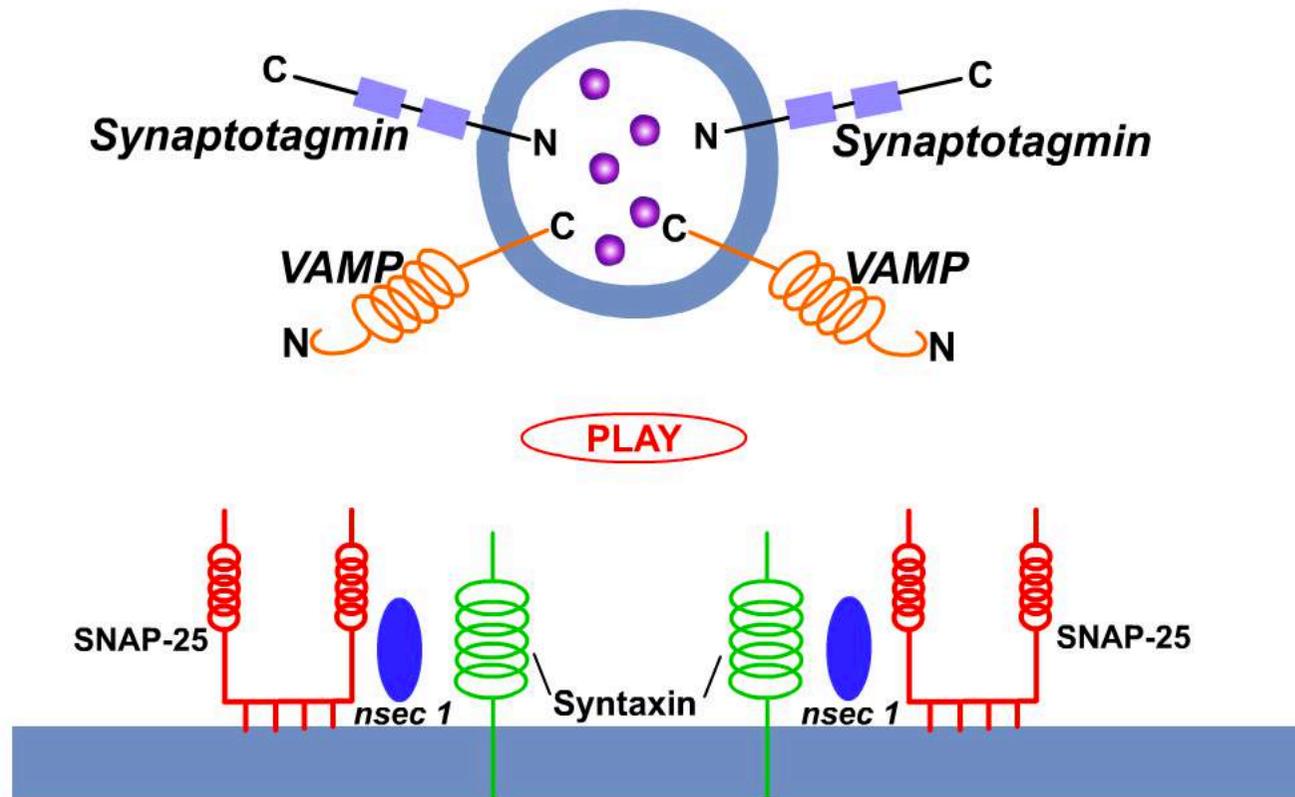
Neurotransmitter is secreted at the nerve ending through the  $\text{Ca}^{2+}$ -dependent fusion of neurotransmitter vesicle with the plasma membrane with the neurotransmitter being secreted (released) into the synaptic cleft to influence the postsynaptic cell. This process is termed exocytosis. An important concept to emerge is that the same mechanisms that occur in vesicle fusion with membranes in the Golgi during vesicle biosynthesis are also employed at the nerve ending for neurotransmitter release. This is true for vesicle synthesis. Even very simple cells like yeast. This conservation of mechanisms has permitted the use of simple systems to help understand the molecular mechanisms of neurotransmitter release.

The first event that must occur (with the exception of neuropeptide neurotransmitters) is the filling of vesicles with neurotransmitter through specific neurotransmitter uptake (NT Uptake). This uptake will be covered in subsequent chapters that discuss each of the specific neurotransmitters.

The vesicles remain in reserve until needed for secretion. When needed for secretion, a translocation occurs, which is also referred to as mobilization. The vesicles move to a region of plasma membrane called the active zone. The active zone is the release site and is characterized by the appearance of dense material adjacent to the plasma membrane. The influx of  $\text{Ca}^{2+}$  is believed to increase translocation by increasing the  $\text{Ca}^{2+}$  dependent phosphorylation of a vesicle binding protein termed synapsin. The theory is that  $\text{Ca}^{2+}$  dependent phosphorylation of synapsin frees the vesicles from binding to actin microfilaments. The vesicles then bind to the active zone of the plasma membrane, where they are in position to undergo release of their neurotransmitter.

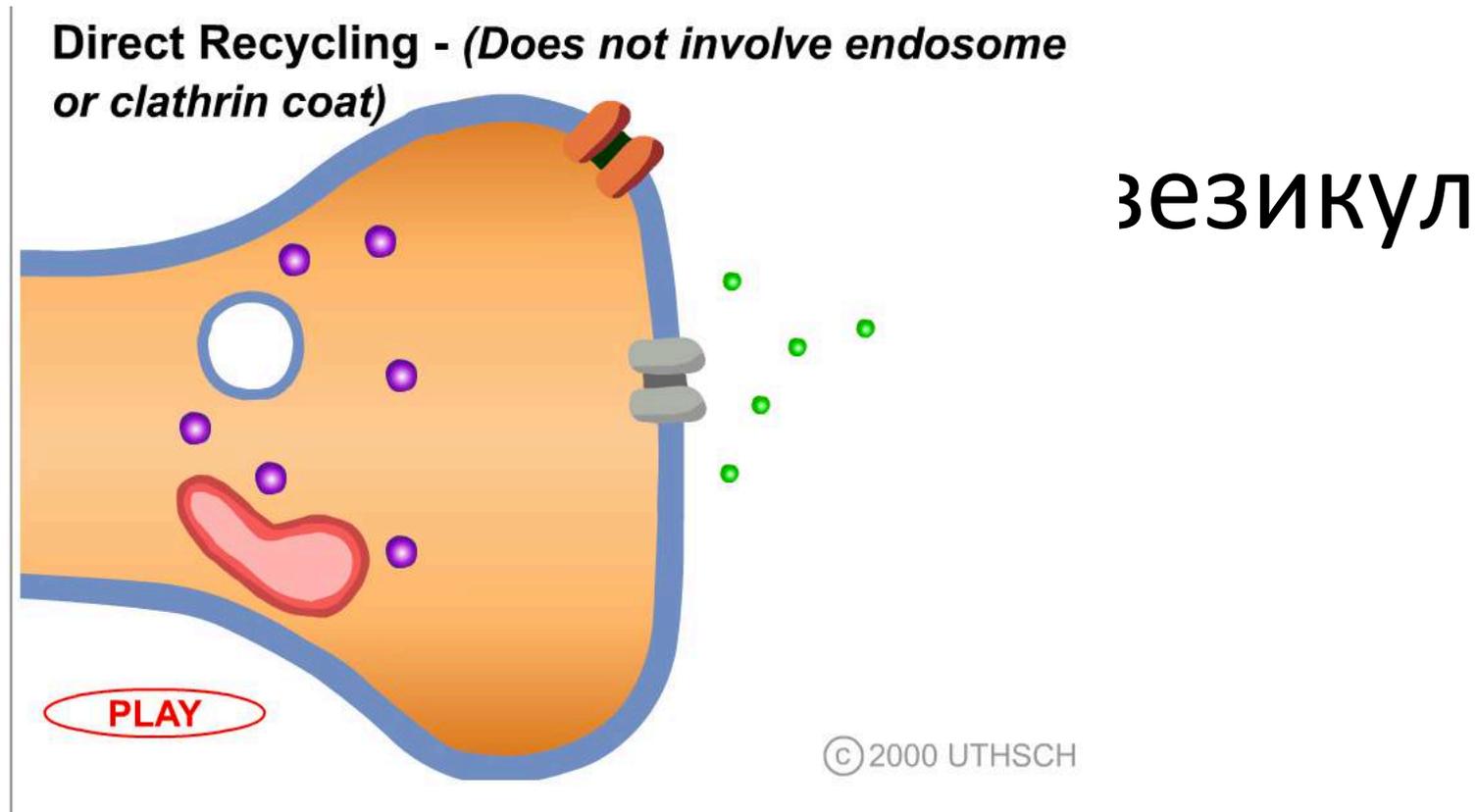
The association of the vesicle with the plasma membrane is termed docking and serves to prime the vesicle for secretion. The docking is believed to occur through the binding of proteins in the vesicle membrane to proteins in the plasma membrane. Several of these proteins have been discovered because they are targets for clostridia bacterial toxins that block synaptic transmission. Several of these toxins and the proteins they detect are shown in Table I. The toxins are so toxic that a single molecule can poison a whole nerve terminal. One of the synaptic vesicle proteins is VAMP, and two of the synaptic plasmal membrane proteins are syntaxin and SNAP-25.

<b>Toxin</b>	<b>Synaptic Protein</b>	<b>Location</b>
Botulinum toxins A & E	SNAP-25	Synaptic plasma membrane
Botulinum toxin C1	Syntaxins	Synaptic plasma membrane
Botulinum toxin B, D, F & G & tetanus toxin	VAMPs	Synaptic vesicle



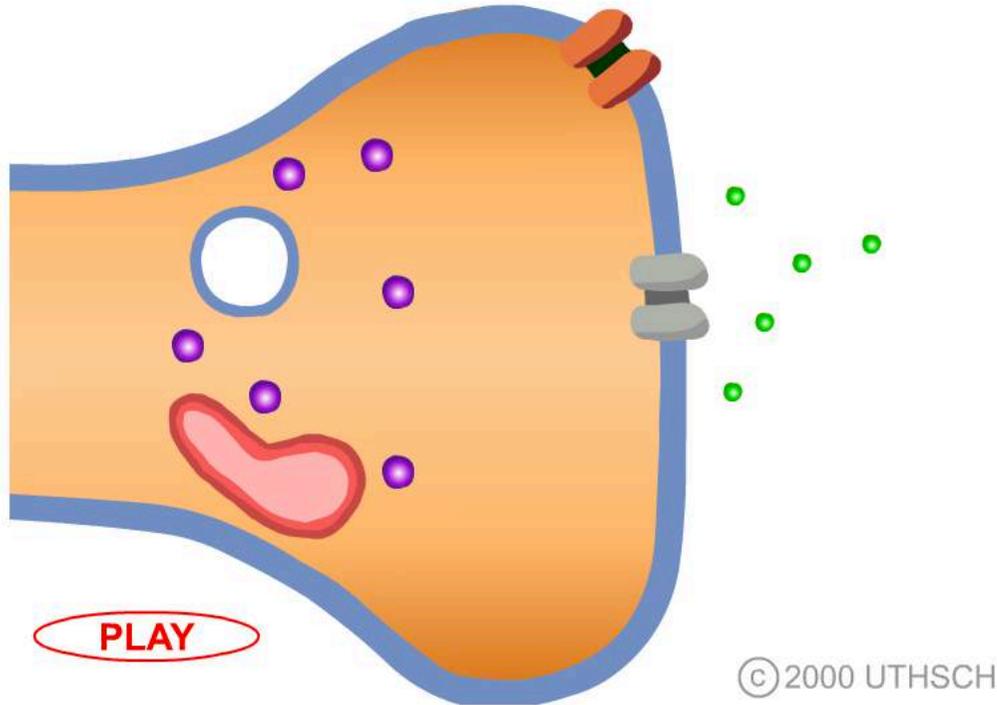
A third plasma membrane protein, n-sec-1, is important because its loose association with the plasma membrane prevents the binding of the neurotransmitter vesicle proteins until n-sec-1 is displaced (the mechanism of n-sec-1 displacement is currently not understood). This and subsequent steps in the secretory process are shown in Figure 10.8. The vesicle and plasma membrane proteins are hypothesized to complex with one another upon the displacement of n-sec-1 to form a "trimeric complex" (SNAP-25, syntaxin and VAMP). This three-member complex has been isolated, intact, from the nerve endings of animals. This association of the proteins initiates fusion. Vesicles at this stage are primed for release.

The final stage of release, also shown in Figure 10.8, is the fission of the membrane at the point of contact between the vesicle and the plasma membrane. Exocytosis of neurotransmitter into the synaptic cleft occurs when this fission takes place. This step is  $\text{Ca}^{2+}$  stimulated, but the mechanism of the  $\text{Ca}^{2+}$  trigger is unknown. One hypothesis is that a vesicle protein called synaptotagmin binds  $\text{Ca}^{2+}$  to initiate fission. Support for synaptotagmin, as the  $\text{Ca}^{2+}$  sensor, is that it possesses two binding sites for  $\text{Ca}^{2+}$ . Additional evidence comes from studies of mice in which synaptotagmin has been knocked out. In these mice fast  $\text{Ca}^{2+}$ -triggered synaptic vesicle exocytosis is severely limited. Many aspects of the fusion-fission mechanism remain to be understood, including: what causes the dissociation of n-sec-1 from the complex, how  $\text{Ca}^{2+}$  functions in the release process and how all the proteins that are involved in release become reassociated with the proper membrane following release as the vesicle membrane is recycled.

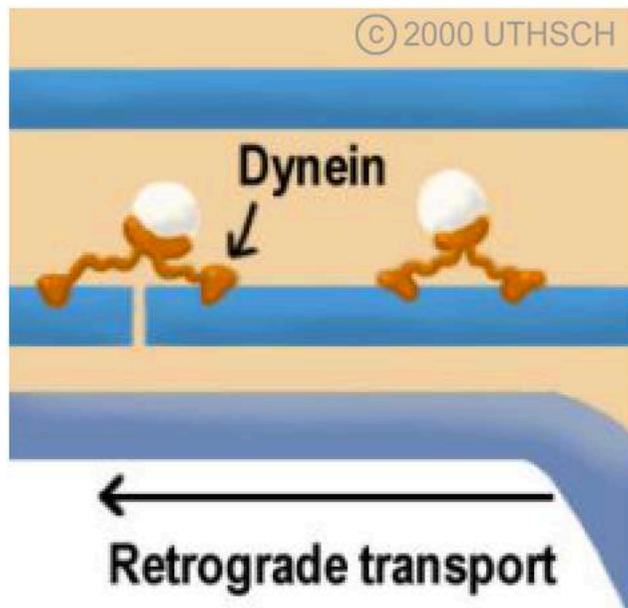


Although the source of vesicles for neurotransmitter secretion comes from biosynthesis in the Golgi apparatus at the cell body, evidence indicates that local re-synthesis of synaptic vesicle is an important aspect of neurotransmitter secretion. Figures 10.9 and 10.10 provides two schematic summaries of how vesicles are locally reused. Both utilize the recapture of vesicle membrane from the nerve ending. In one, vesicles bud off the plasma membrane through the formation of pits that migrate directly to become a neurotransmitter vesicle as soon as it can be refilled with neurotransmitter through the neurotransmitter uptake process. This is shown in Figure 10.9. This mechanism is referred to as the "kiss and run" hypothesis.

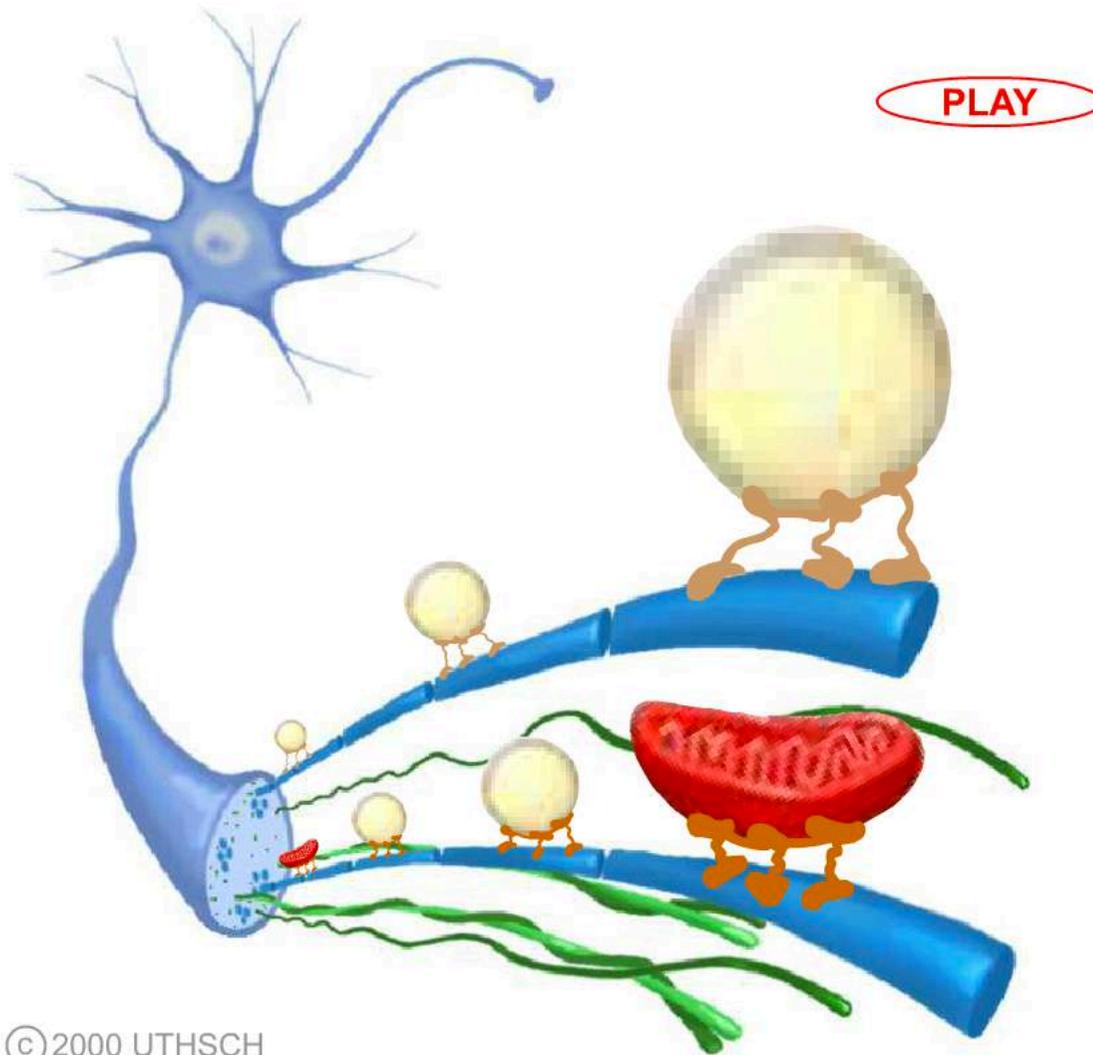
## Indirect Recycling - (*Does involve endosome*)



The second mechanism involves the formation of clathrin coated pits to recapture vesicle membrane, and the vesicles cycle through the endosomal compartment in the nerve ending before becoming functional synaptic vesicles. The vesicles then bud off the endosome to form the neurotransmitter vesicle. This is shown in Figure 10.10. It is believed that both mechanisms can exist in the same nerve ending or only one of the two can be present. Both are important in that they recover vesicle protein to permit a plentiful supply for synaptic transmission. This mechanism also prevents the enlargement of the nerve ending that would occur if vesicle membrane were not recaptured. No matter which mechanism is involved, the supply and resupply of vesicles can only keep pace with a high rate of synaptic transmission for a few minutes.



Eventually the proteins utilized for synaptic transmission in the nerve ending are targeted and returned to the cell body of the neuron to be recycled to make new protein and vesicles. The proteins are returned to the soma through a retrograde axoplasmic transport that is analogous to anterograde transport but uses a different motor protein, dynein. Transport is mediated by the interaction of dynein with microtubules and proceeds at a rate somewhat slower than that of fast anterograde transport (0.2-1 cm per day). In addition to returning proteins to the soma, retrograde transport serves as a means of communication between the nerve endings and the cell soma. This is a mechanism to transport signaling molecules to regulate the development and maintenance of axonal contacts with postsynaptic cells



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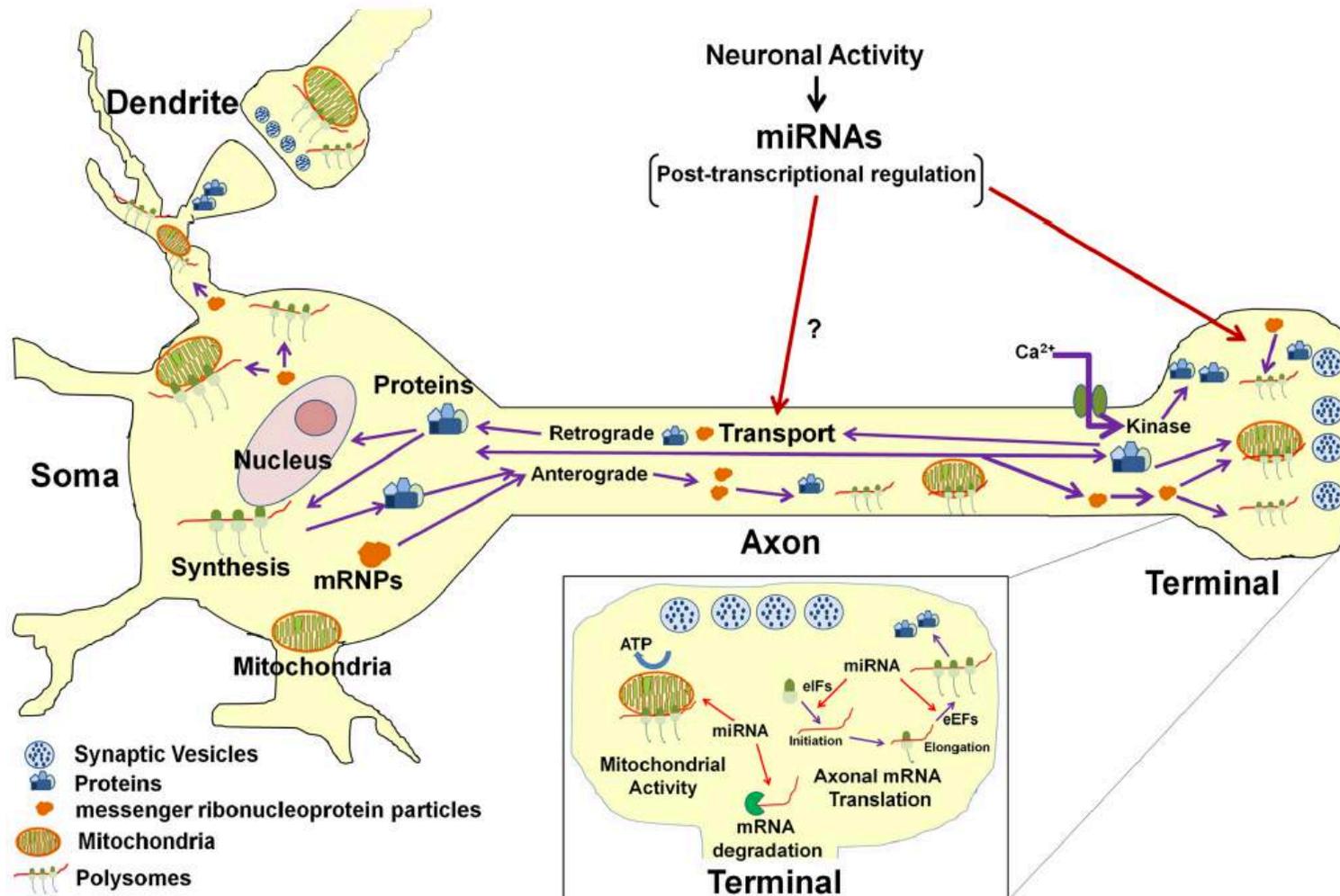
summary of retrograde and anterograde axoplasmic transport. It shows the motor proteins, kinesin and dynein, mediating the movement of vesicles and mitochondria anterogradely and vesicles retrogradely along microtubules. The animation shows the motor proteins as a part of the organelle that is transported. The other possible relationship between the motor protein and microtubules is that the motor proteins are a part of the microtubule and pass the vesicles along the microtubule.

# Question:

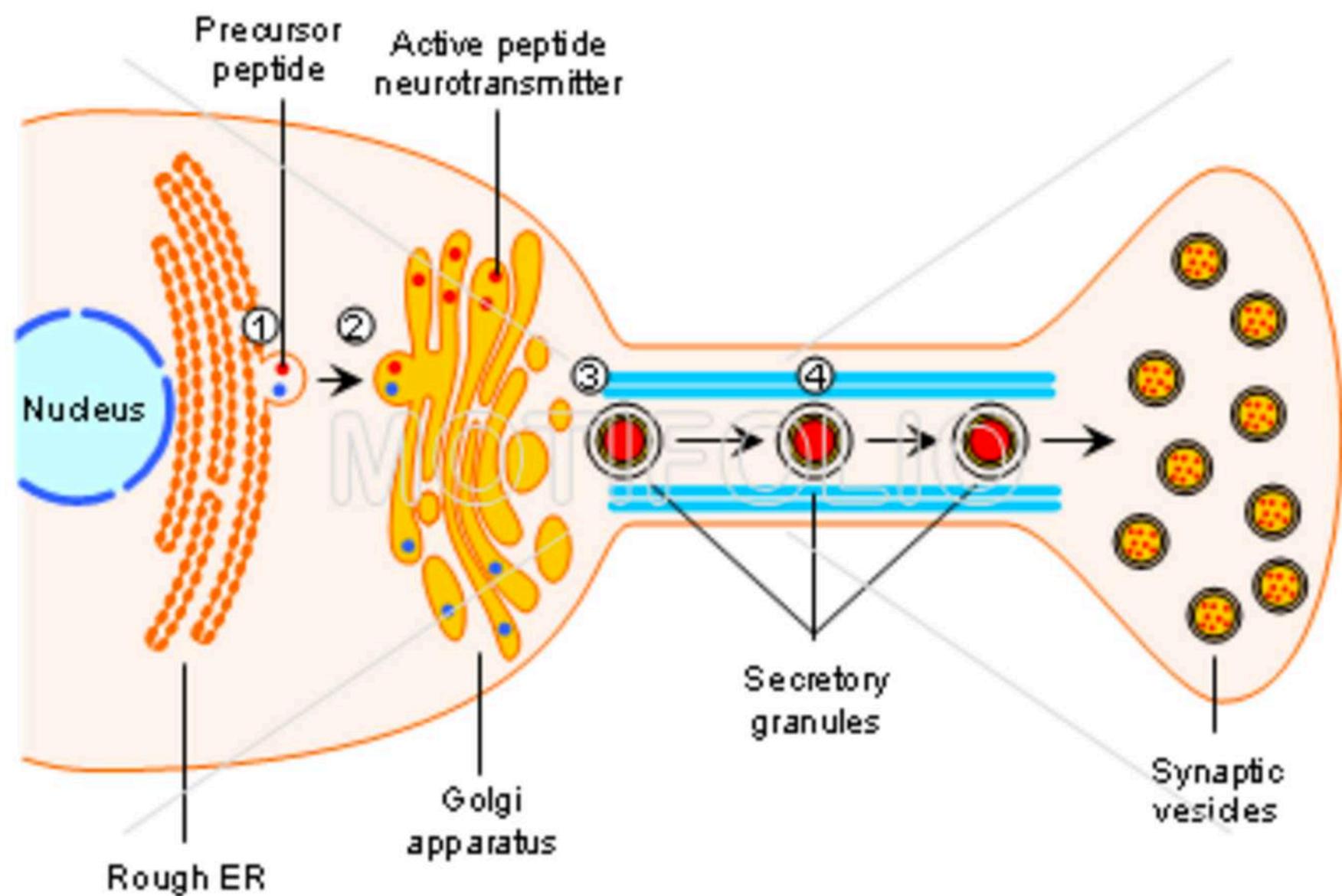
Which of the following processes dictate the amount of neurotransmitter released from a nerve ending on a short-term, minute-to-minute, basis? (NOTE: there is more than one correct answer.)

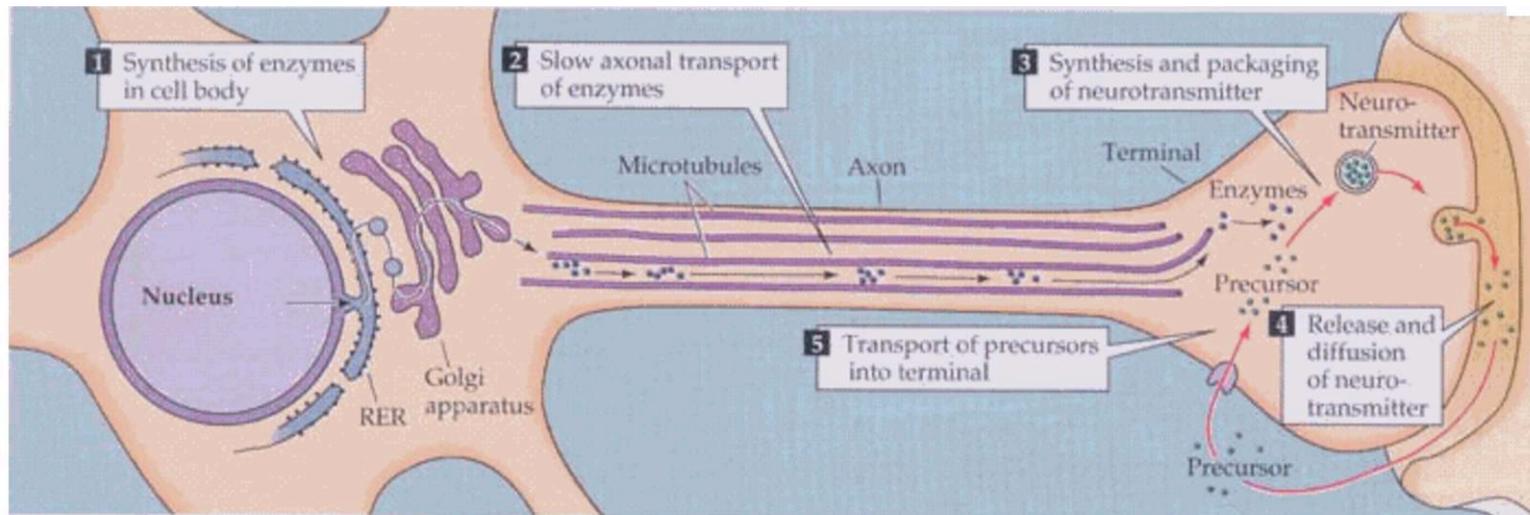
- A. Neurotransmitter synthesis
- B. Vesicle synthesis in the cell soma
- C. Vesicle recycling in the nerve ending
- D. Axoplasmic transport
- E. Calcium availability

# Синтез и транспорт нейромедиаторов

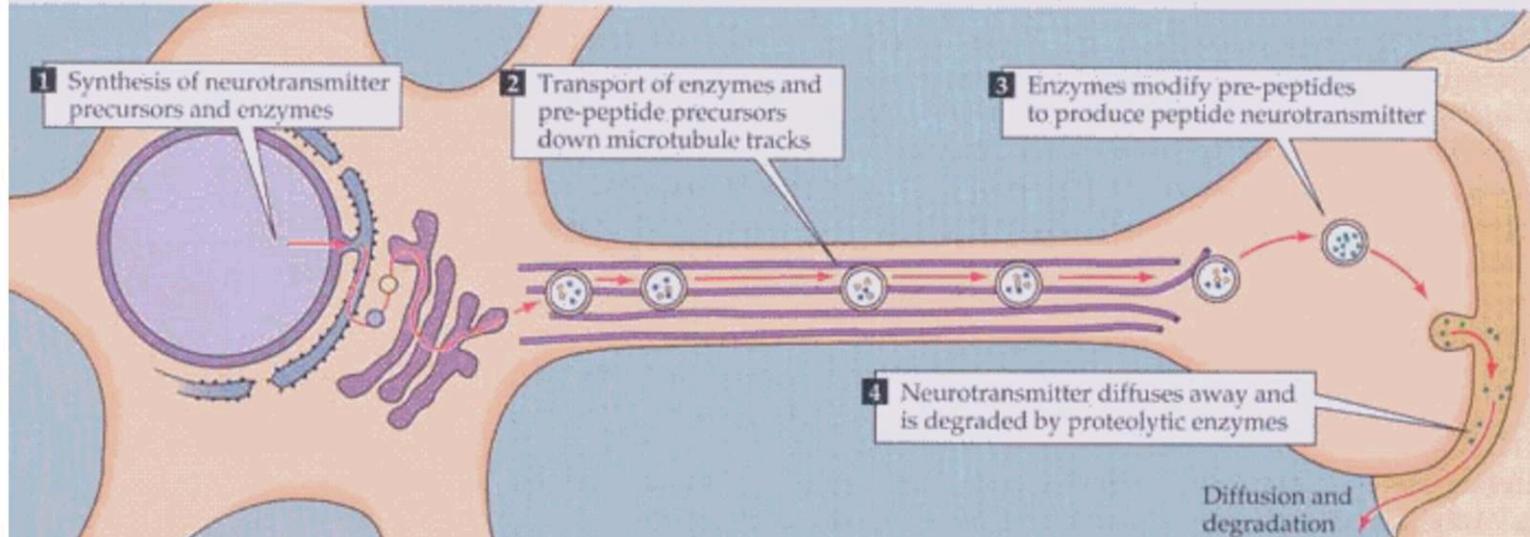


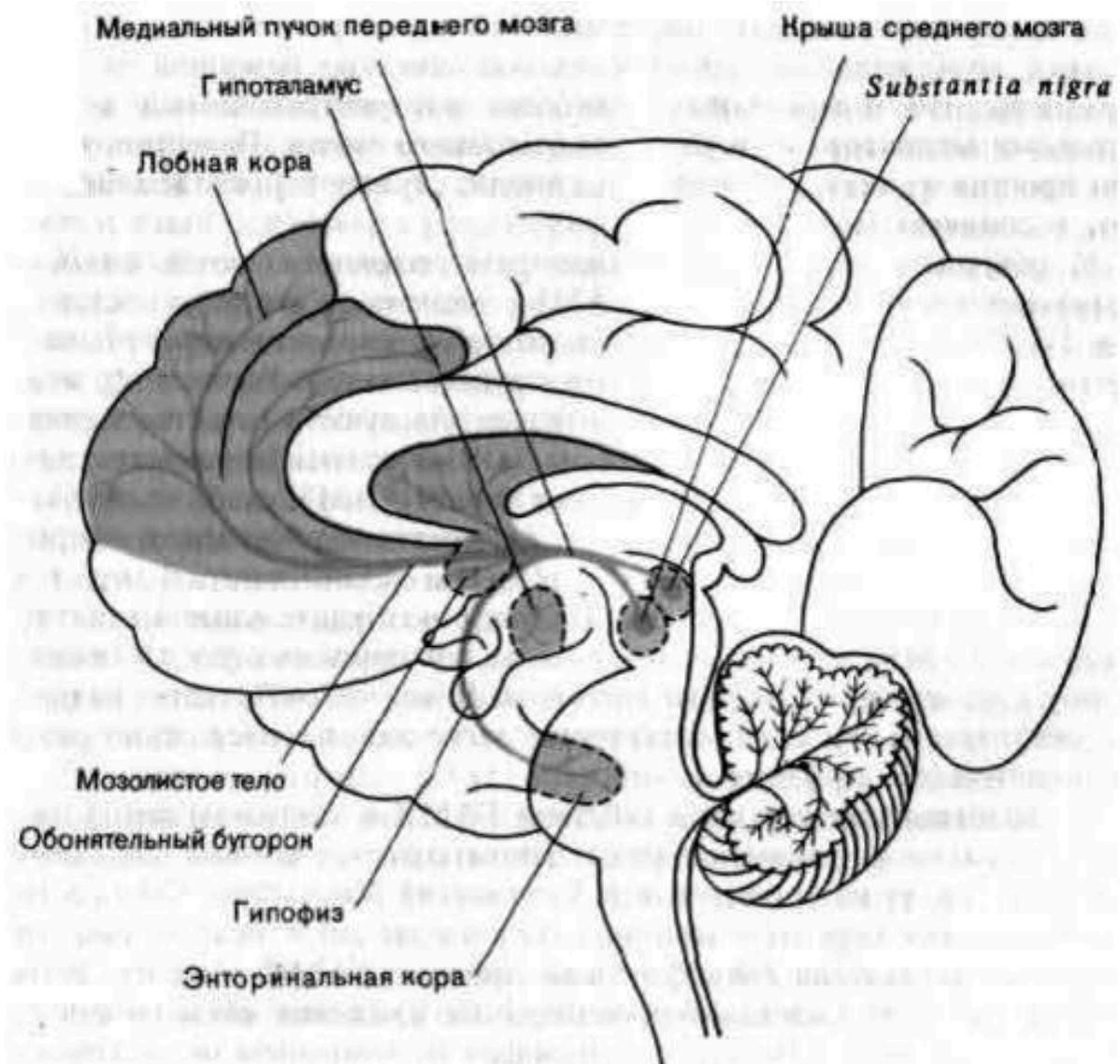
## Synthesis and storage of neurotransmitter (peptides)

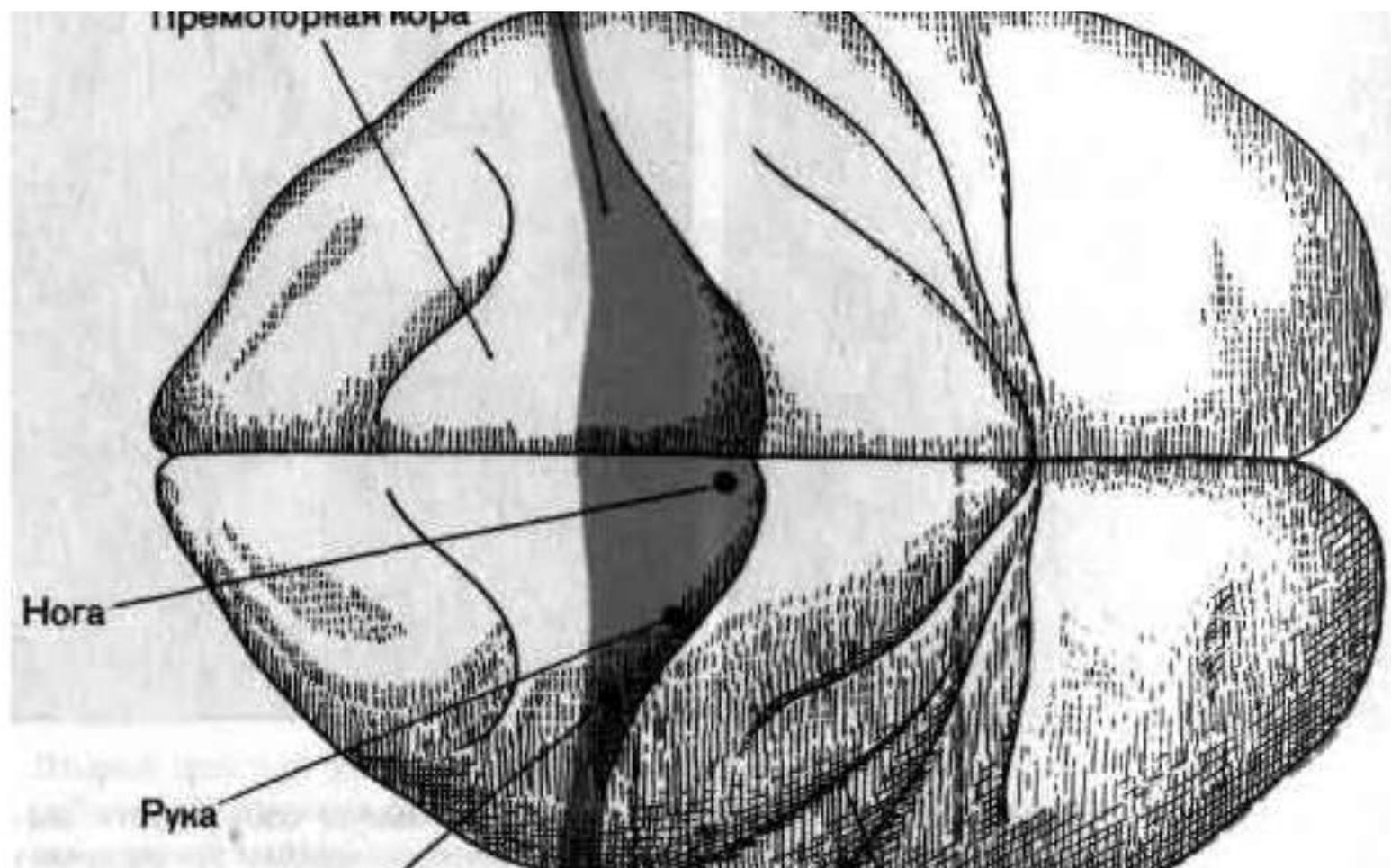


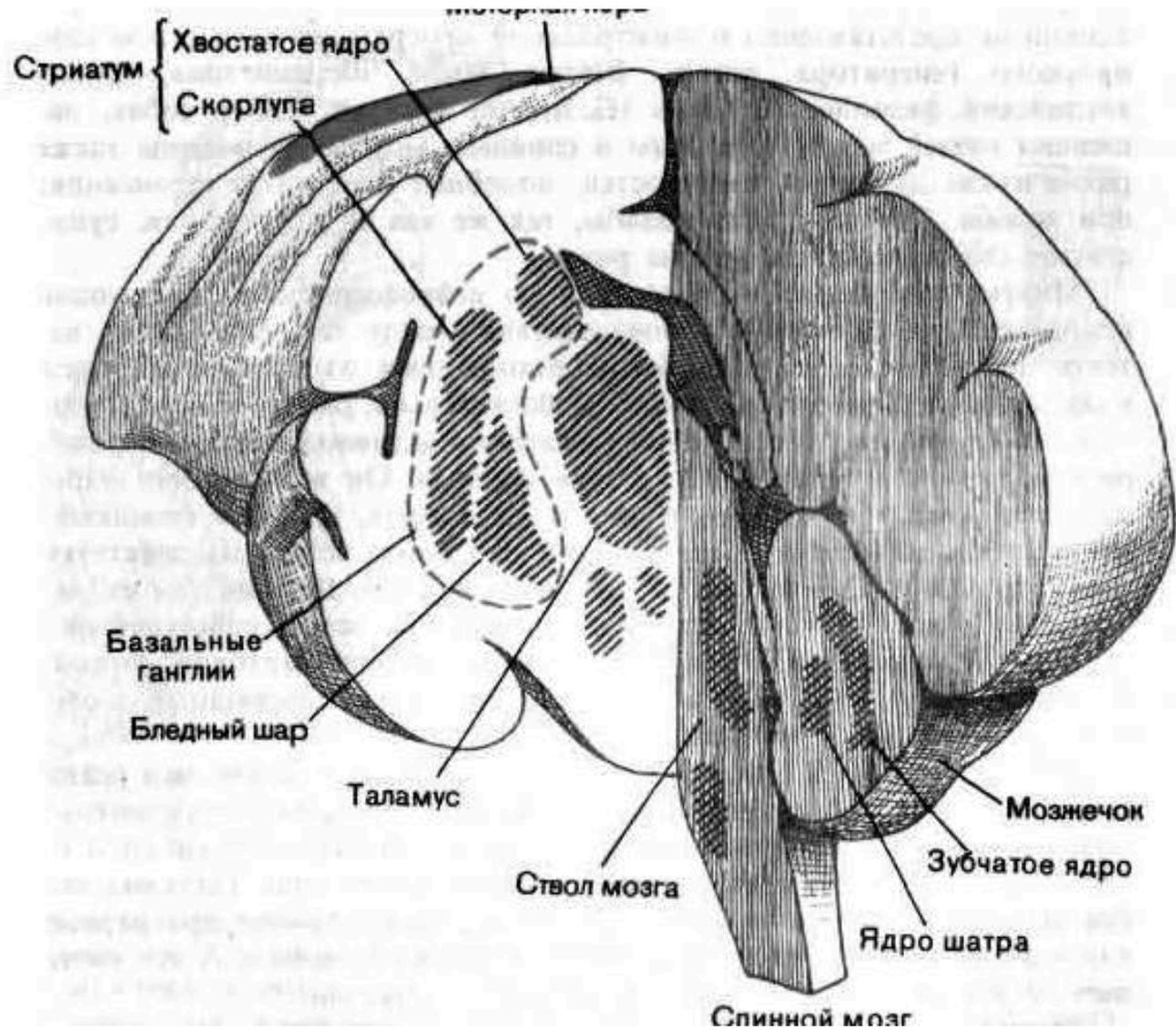


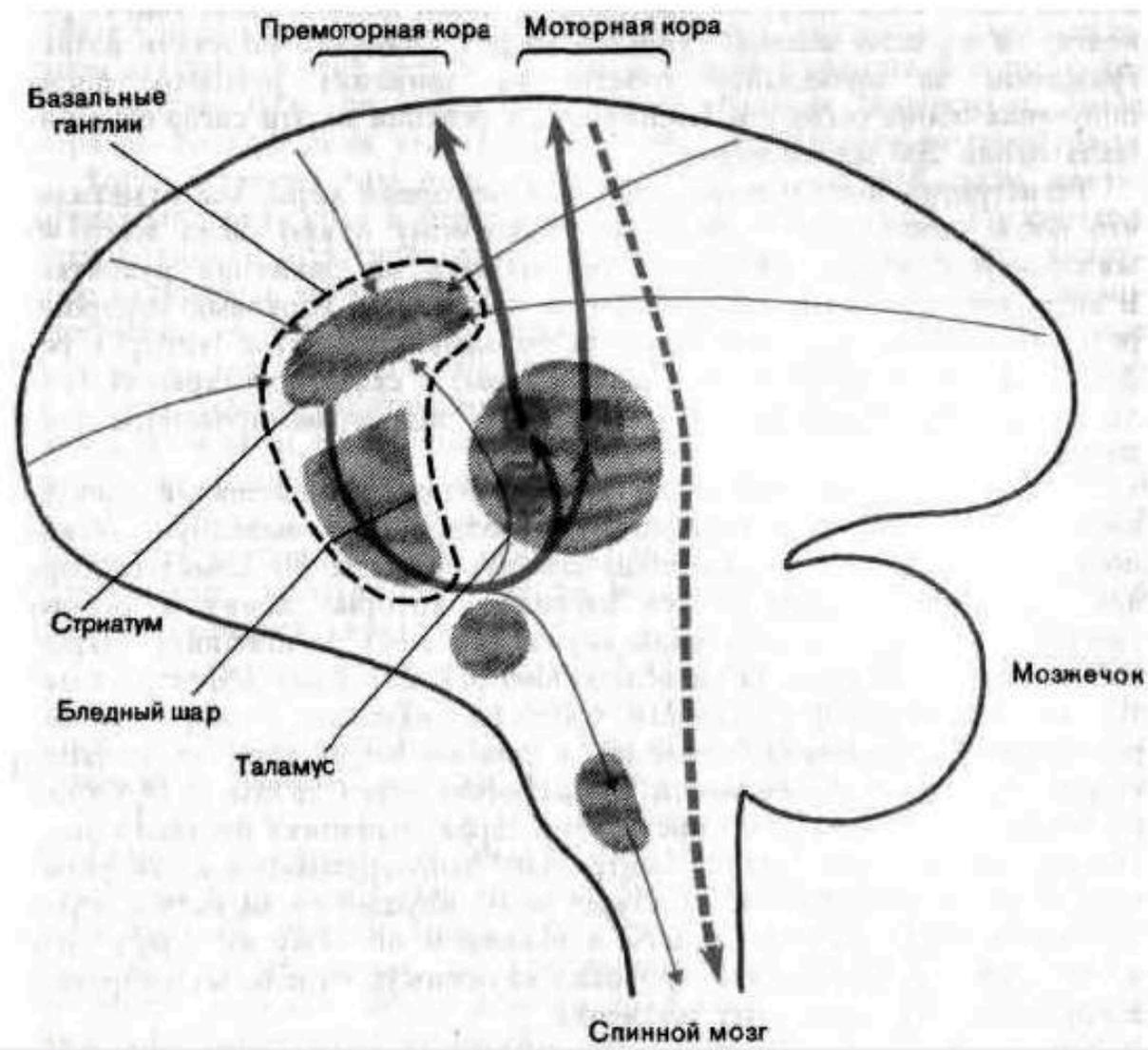
(C) PEPTIDE TRANSMITTERS





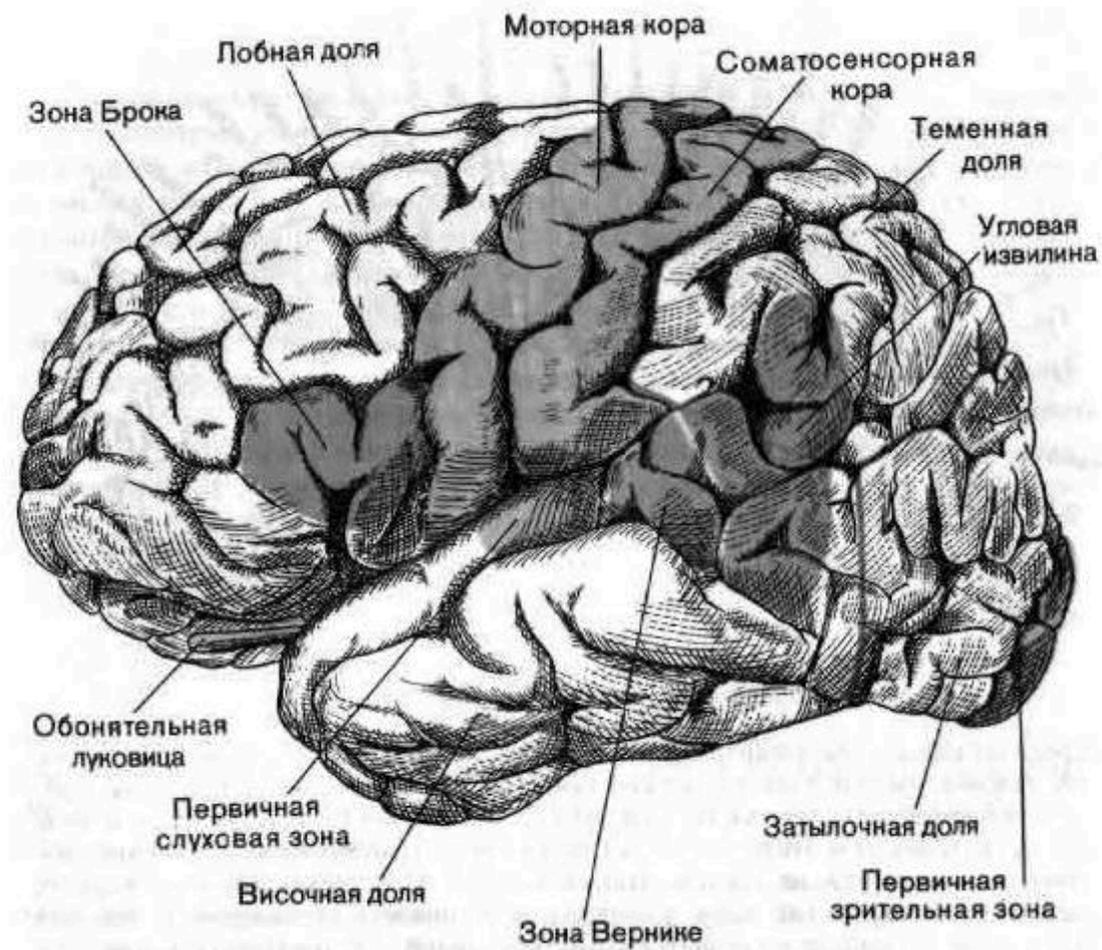


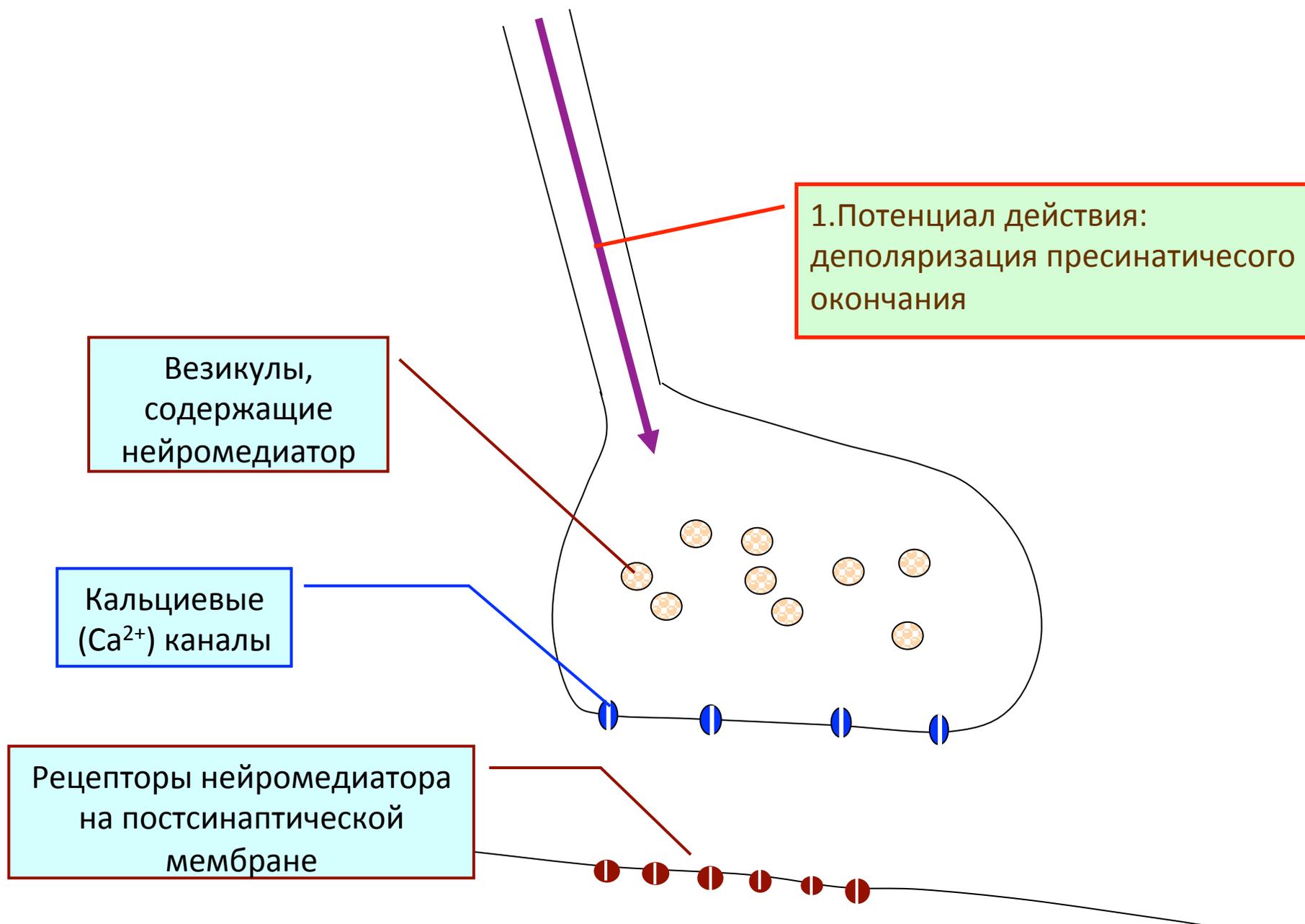


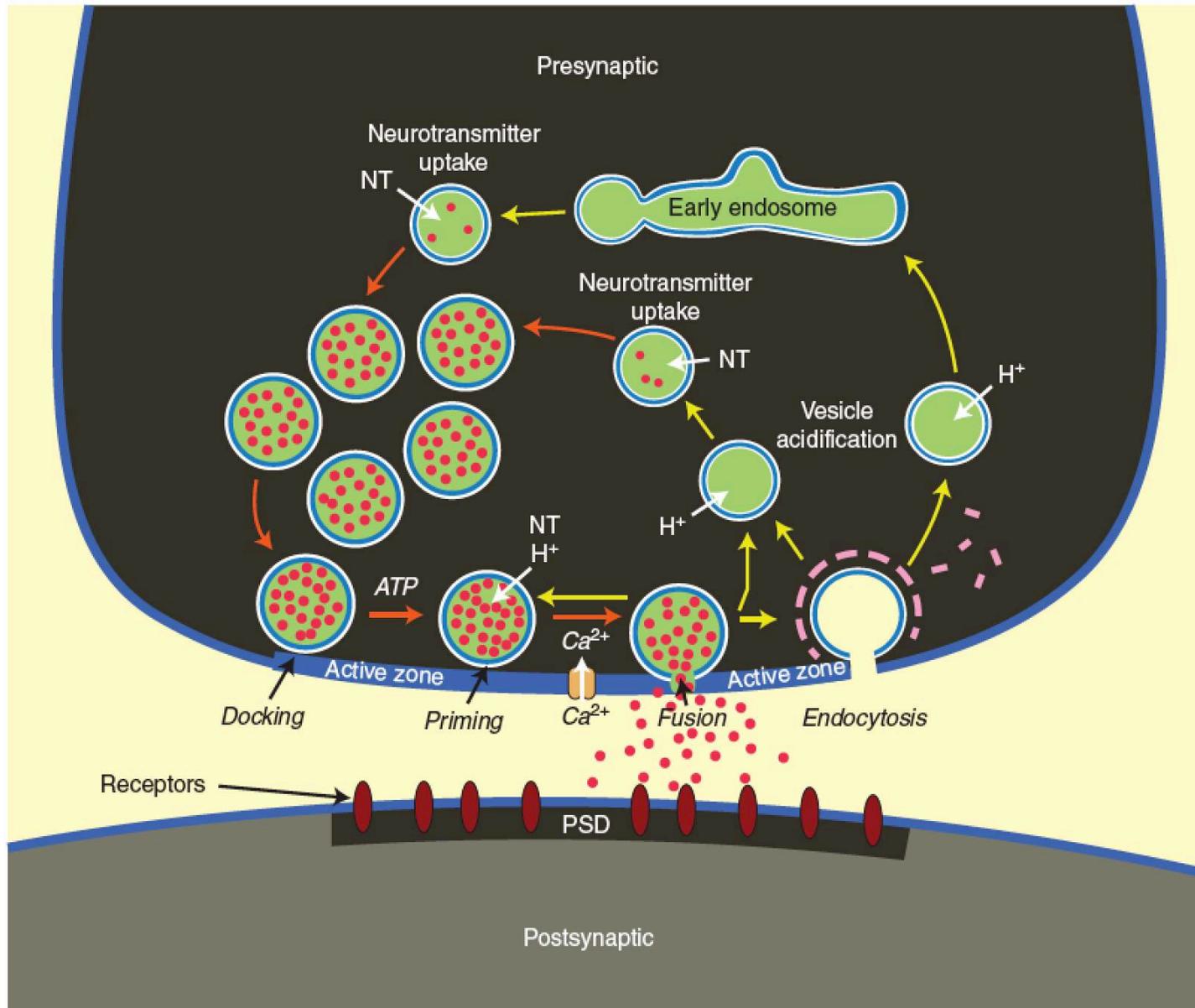


18 СМЫСЛОВЫЕ ЛИЦА.



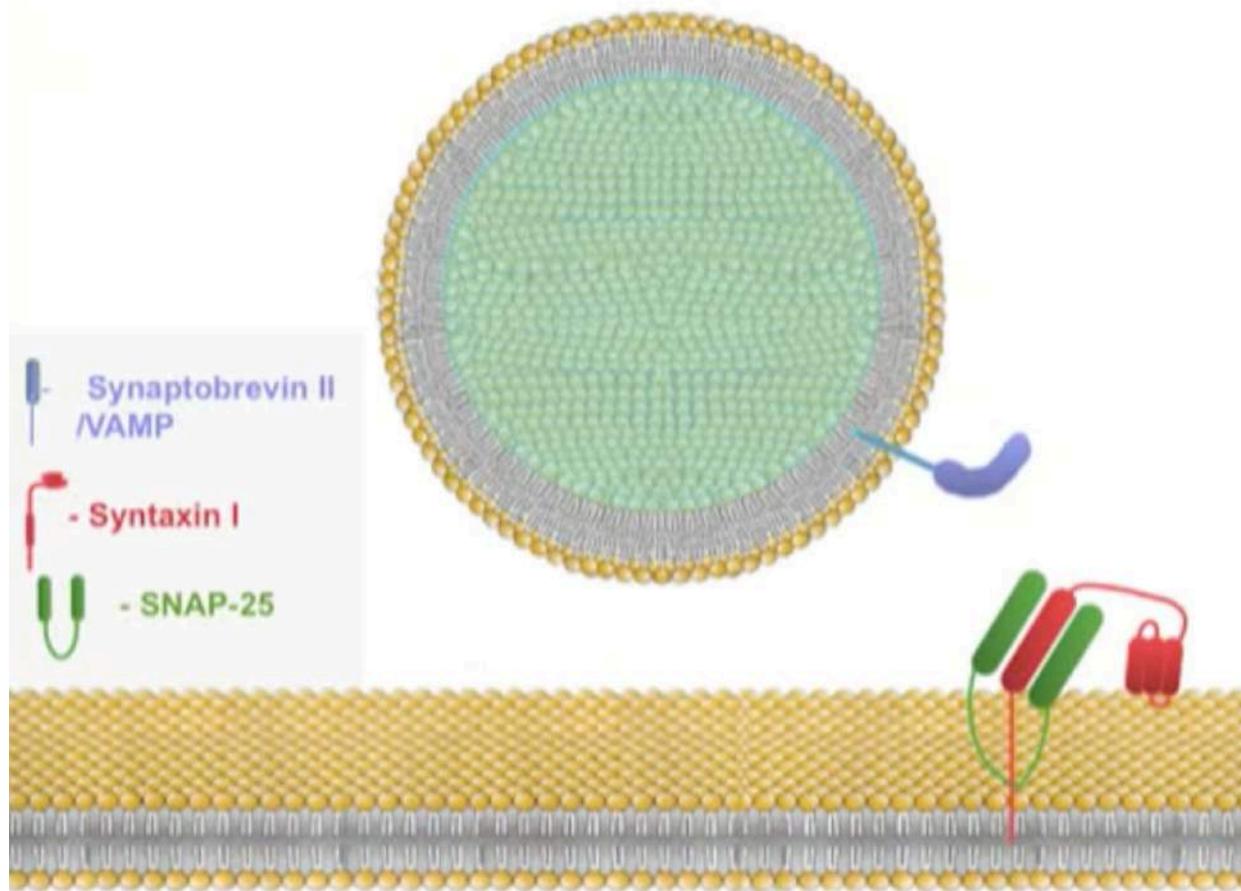


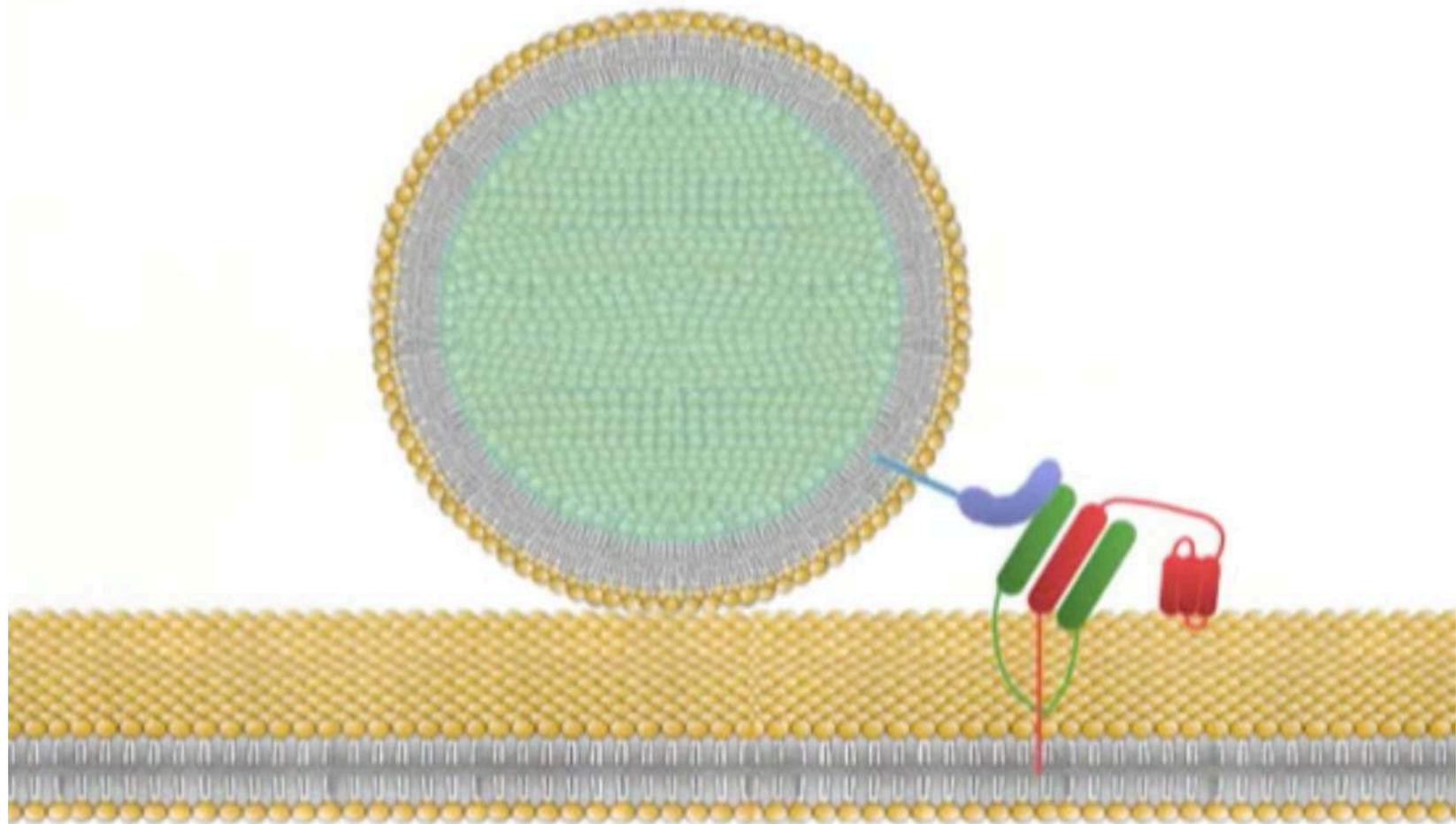


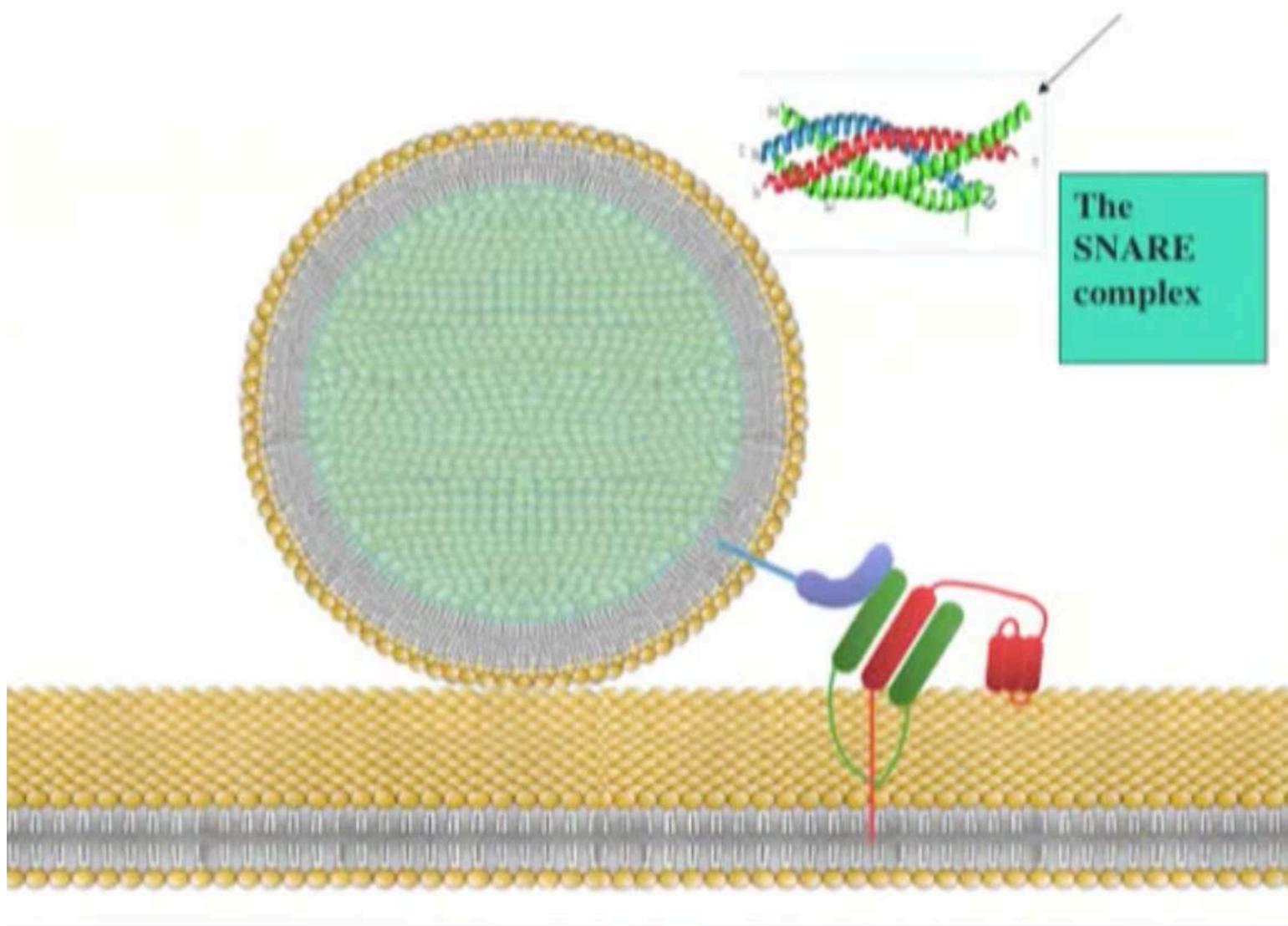


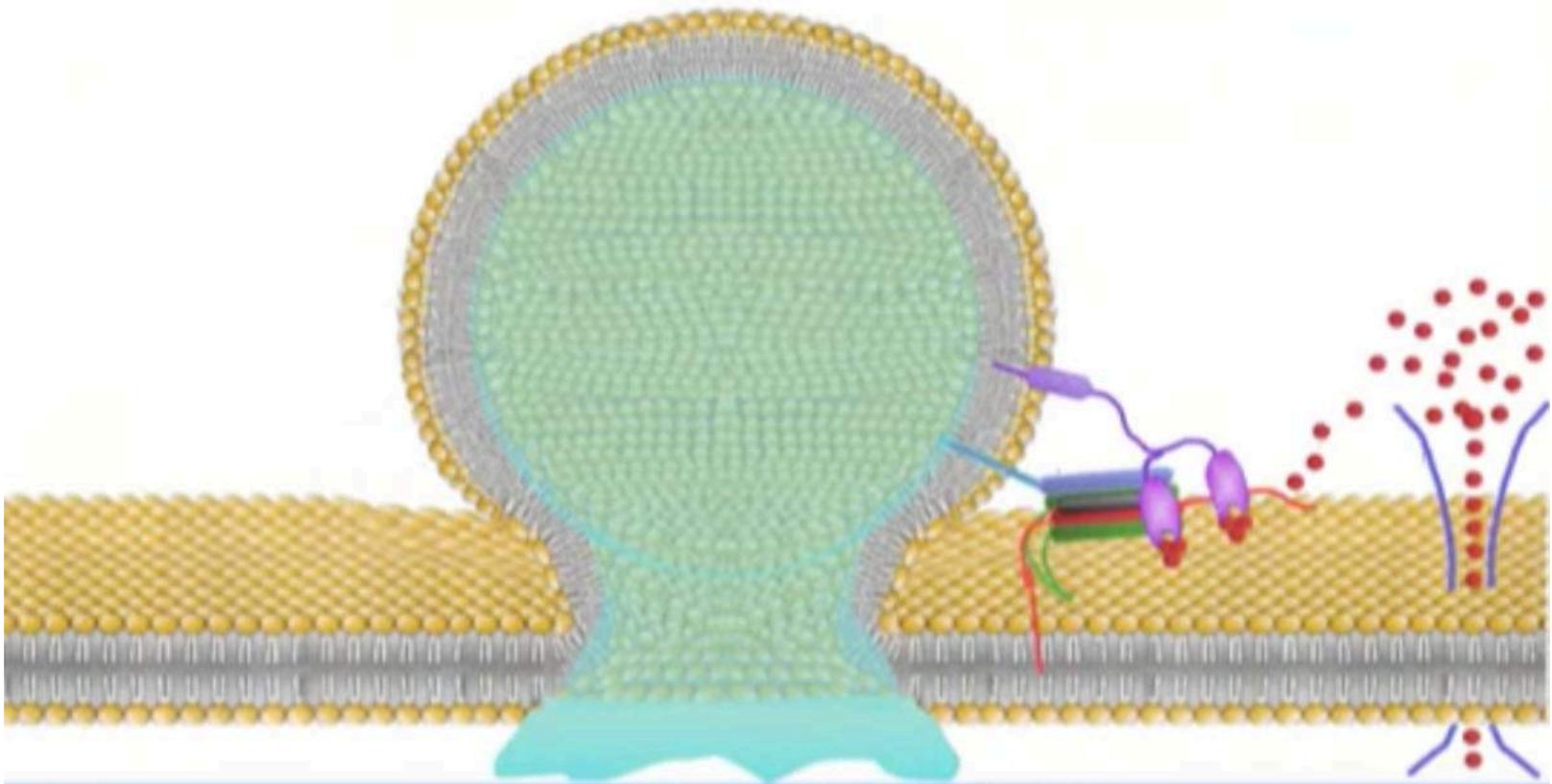
# На память

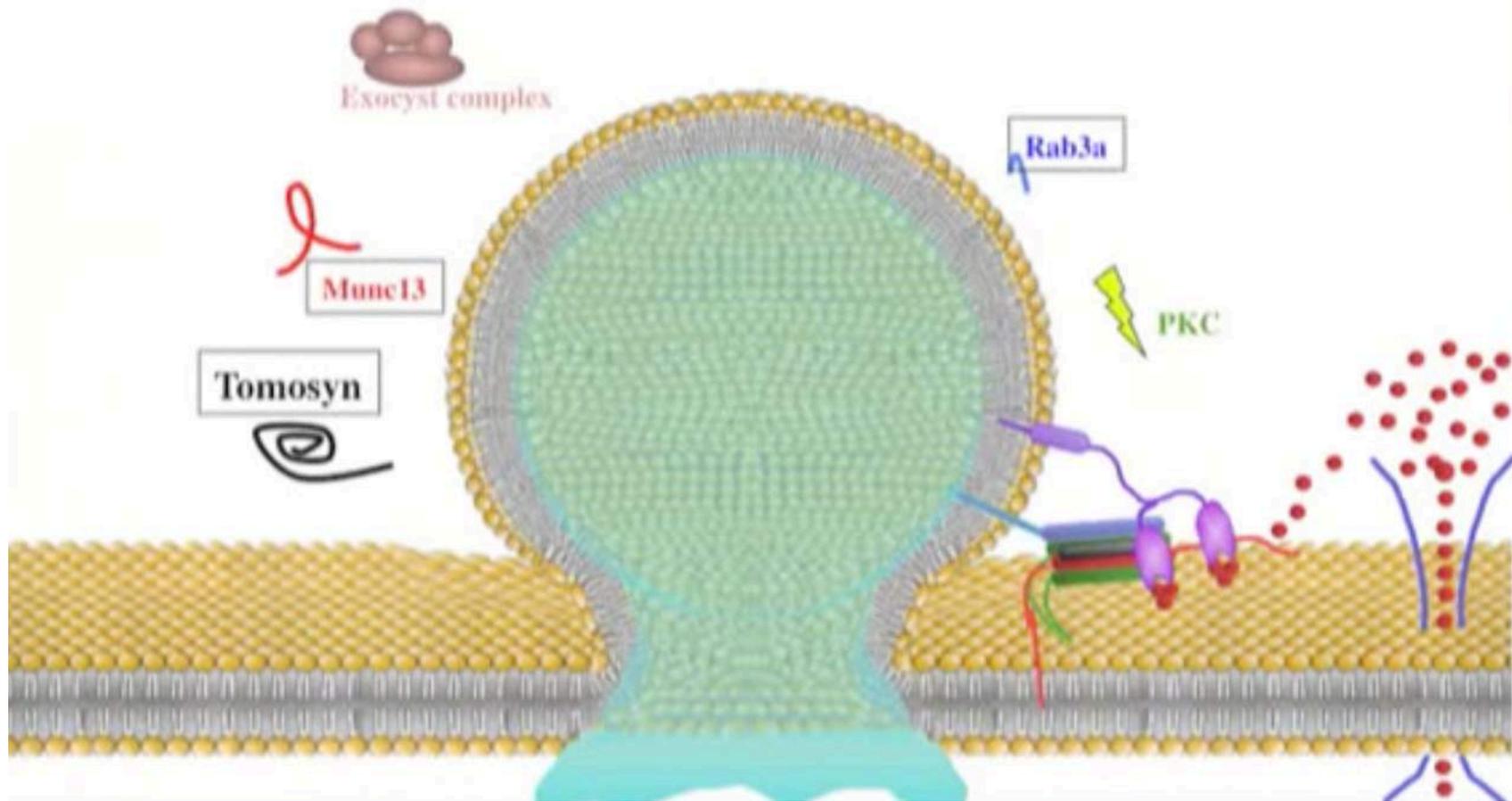
- Кальций связывается с везикулярным белком Синаптоагмином (synaptotagmin), который стимулирует взаимодействие трех белков комплекса SNARE и взаимодействие везикулы с синаптической мембраной, что приводит к её разрыву и выбросу нейромедиатора.

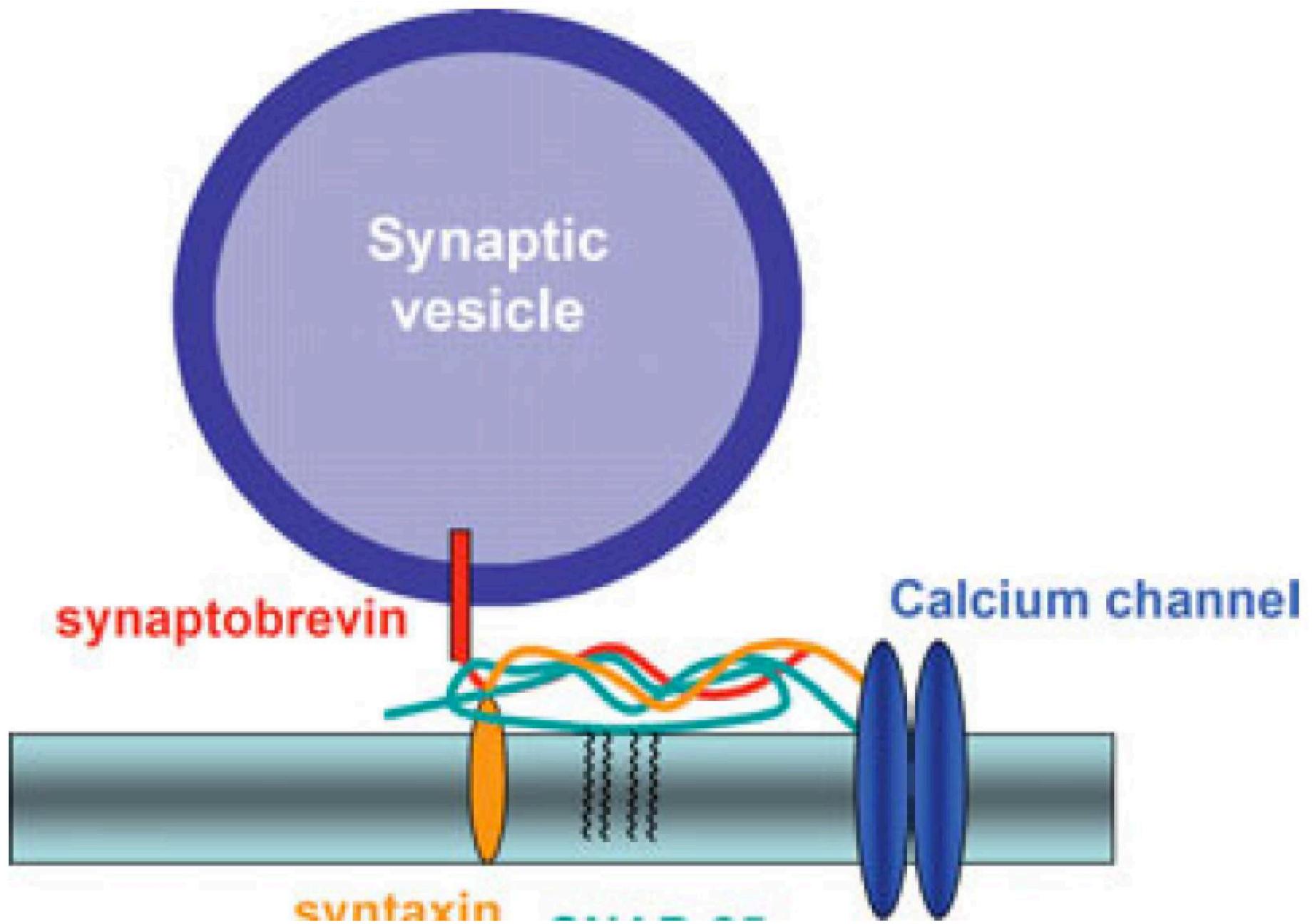




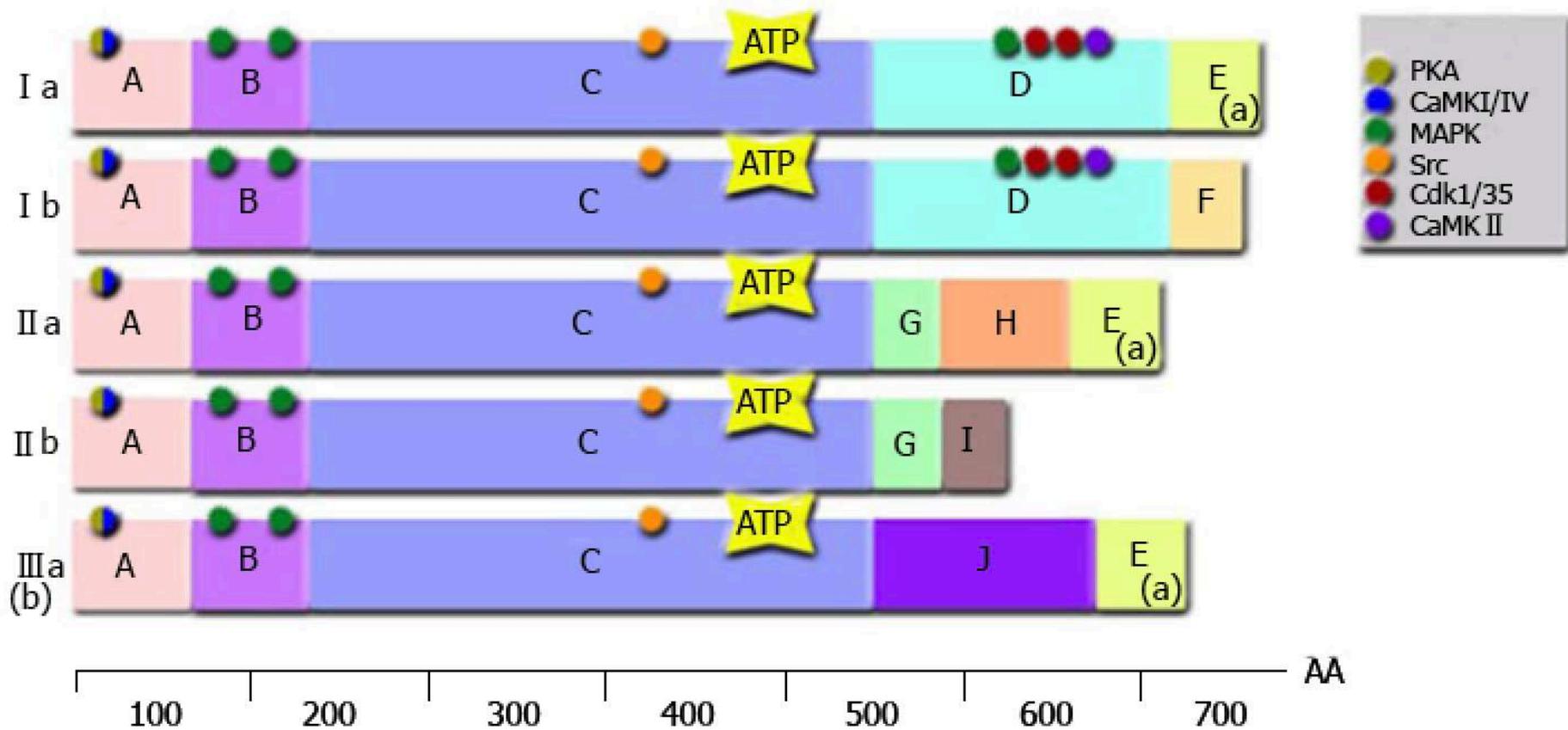








# Синапсы: доменная организация

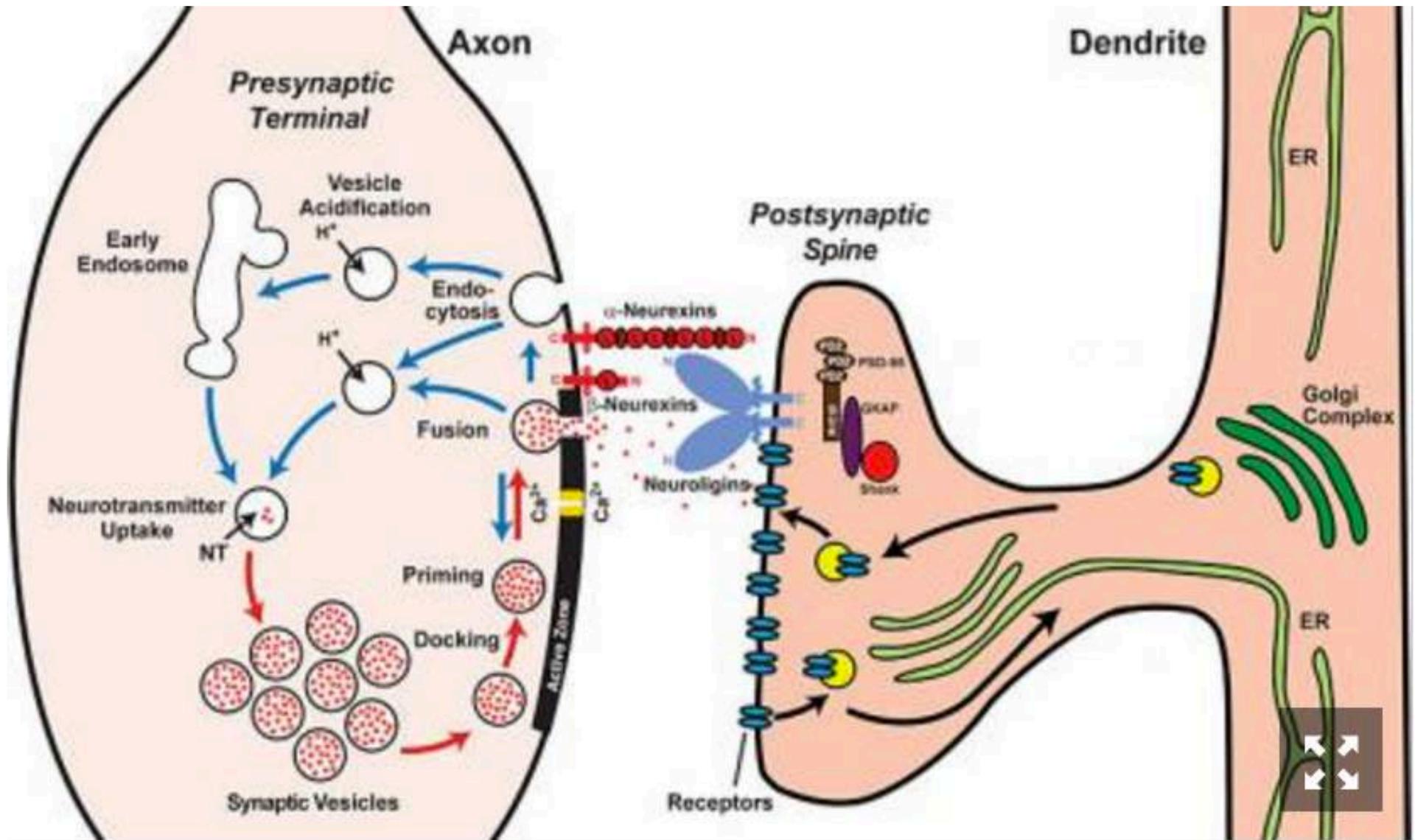


# Синапсины

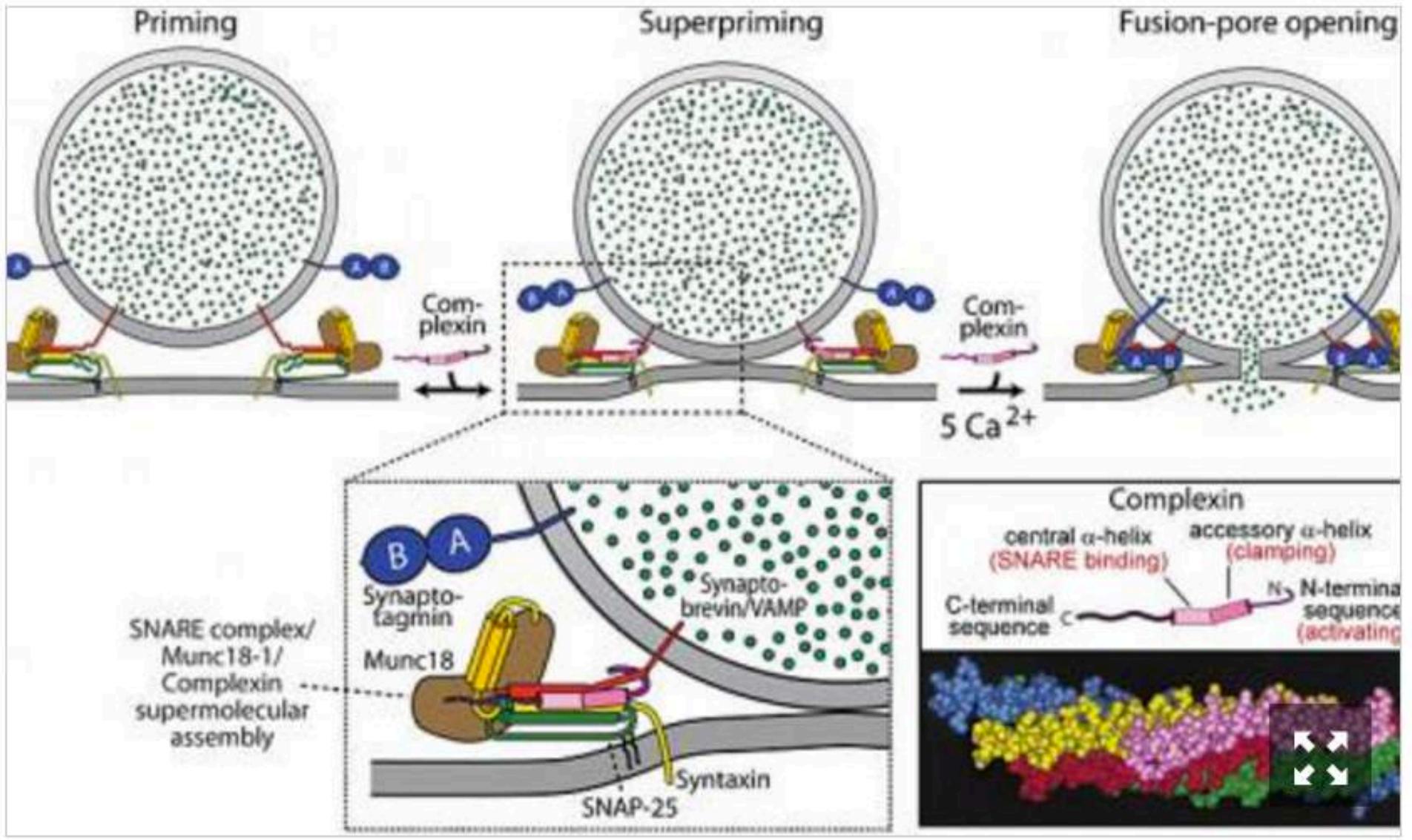
- 9% of the total amount of all vesicle proteins
- presynaptic proteins
- integral for many functional roles, including: synaptogenesis, synapse function, synapse maintenance and synaptic plasticity
- 3 genes: *synapsin I* , *synapsin II* , and *synapsin III*
- synapsins I and II – in mature synapses, while synapsin III - developing synapses
- at least 10 different isoforms of synapsin

- Domain A contains a conserved phosphorylation site for protein kinase A
- (PKA) and Ca<sup>2+</sup>/calmodulin-dependent protein kinase I (CaM kinase I)[1].
- Domain B functions to link the N-terminus to the large central C domain (approximately 300 residues)
- Functionally, synapsins bind to the lipid surface of vesicles *via* the N-terminus, while the variable, hydrophilic C-terminus often facilitates the stabilization of synapsin on phospholipid bilayers and cytoskeletal elements *via* domain E

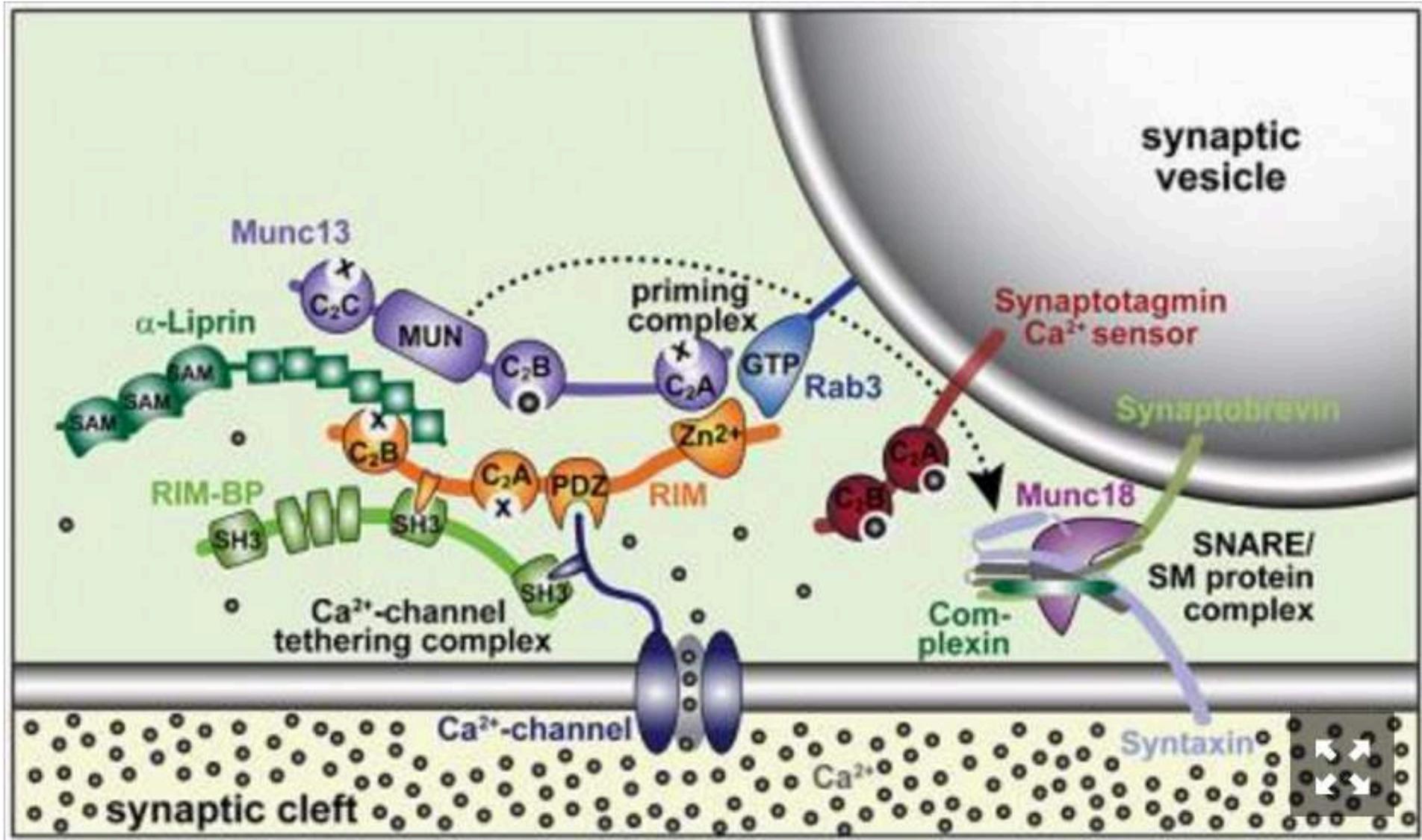




Südhof, T.C., Nature 2008



Gerber, S.H., et al. 2008. *Science* 321:1507–1510

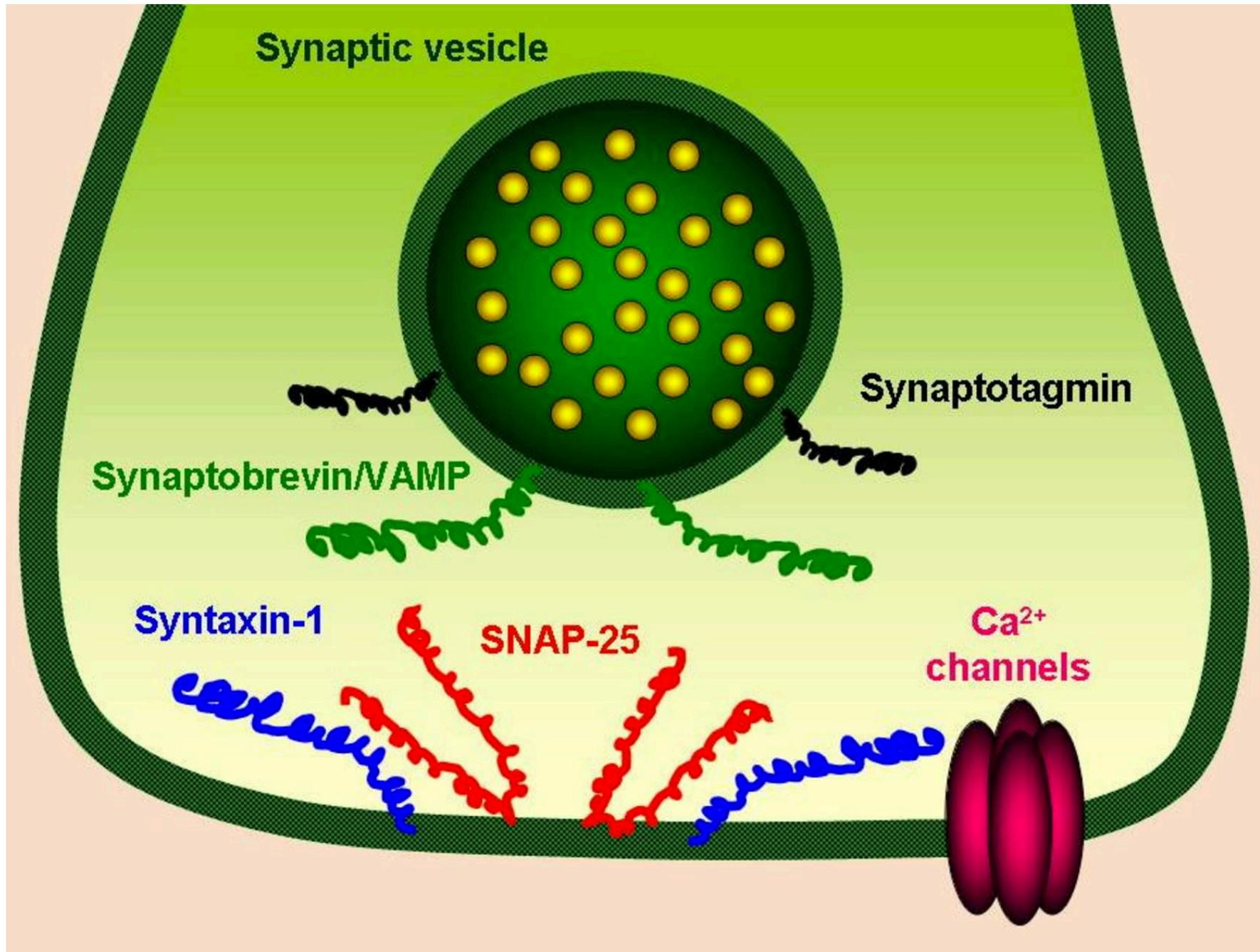


Kaesler, P.S., et al. 2011. *Cell* 144:282-295









# Выброс нейромедиатора:

Строго регулируемый процесс слияния везикул с пресинаптической мембраной под контролем:

- Потенциал-управляемых **Ca<sup>2+</sup> каналов**, которые активируются в ответ на деполяризацию нервного окончания, вызванную нервным импульсом (активация Na<sup>+</sup> каналов)
- **Синаптотагмина ( synaptotagmin)** – сенсора Ca<sup>2+</sup>
- Трех белков с общим названием **SNAREs**:
  - **синаптовревин/VAMP** (расположен в мембране везикулы)
  - **синтаксин (syntaxin-1** - в пресинаптической мембране)
  - **SNAP-25** ( в пресинаптической мембране)

# Выброс нейромедиатора:

- Строго регулируемый процесс слияния везикулы с пресинаптической мембраной под контролем:
  - потенциал-управляемых **Ca<sup>2+</sup> каналов**, которые активируются в ответ на деполяризацию нервного окончания, вызванную нервным импульсом (активация Na<sup>+</sup> каналов);
  - трех белков с общим названием **SNAREs**:
    - **синаптобrevин/VAMP** (расположен в мембране везикулы)
    - **синтаксин (syntaxin-1** - в пресинаптической мембране)
    - **SNAP-25** ( в пресинаптической мембране)

# Болезни из-за нарушений экспрессии SNAREs белков

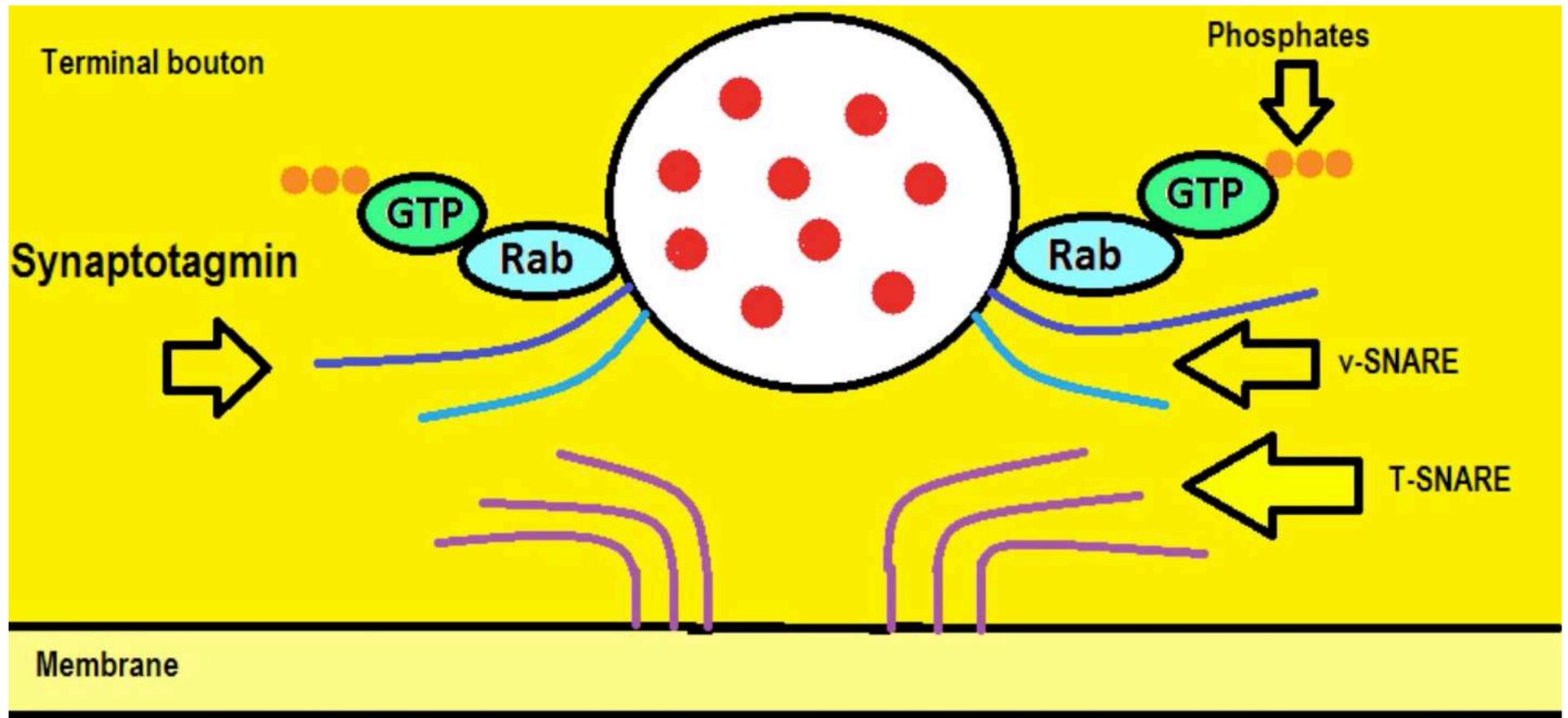


- **Болезнь Хантингтона Huntington disease (HD),**
- **Шизофрения Schizophrenia**
- **Биполярное расстройство (маниакально-депрессивный психоз, МДП)**
- **Болезнь Альцгеймера**
- **Синдром дефицита внимания и гиперактивности (Attention deficit hyperactive disorders(ADHD))**

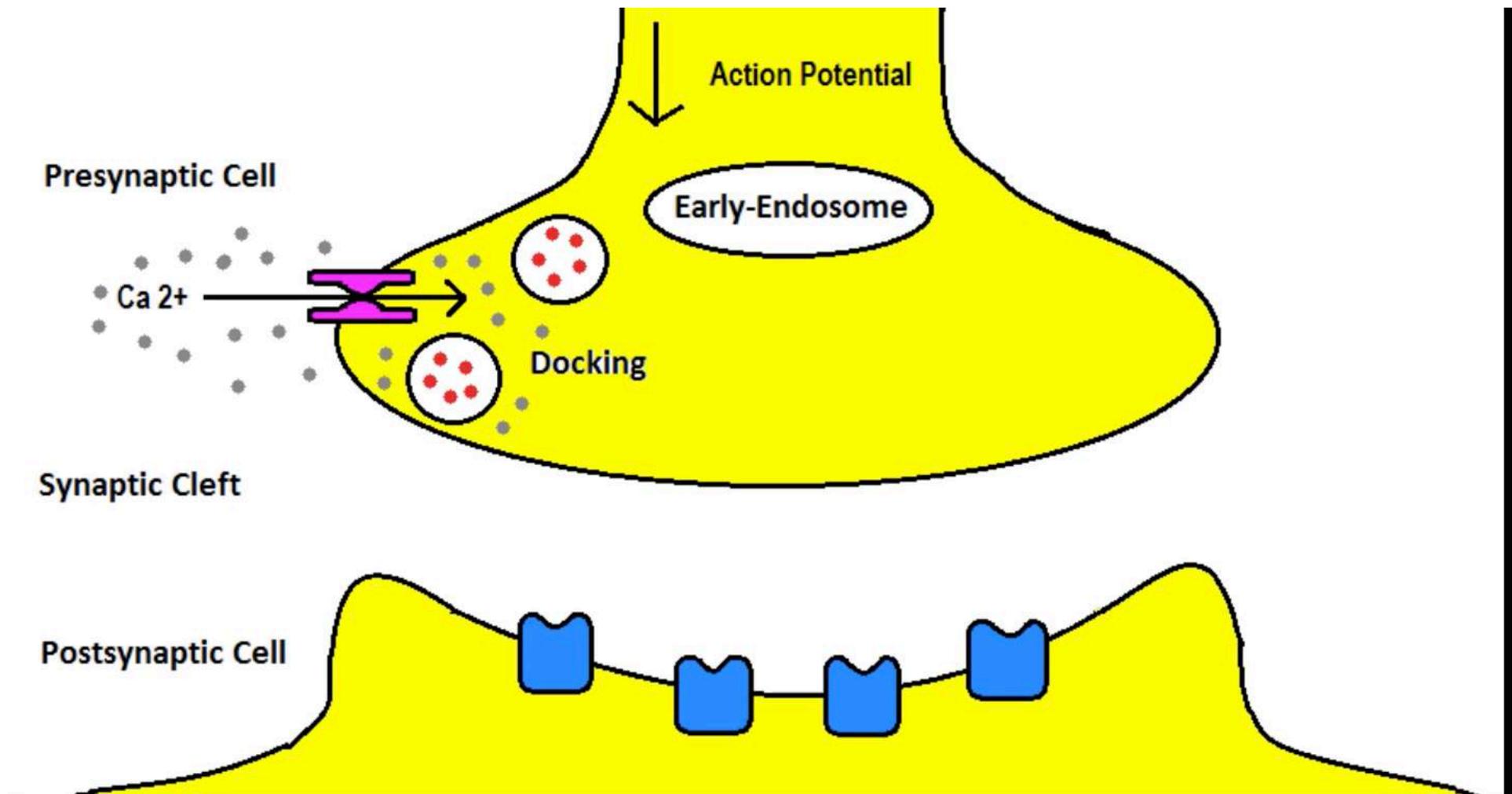
- трех белков с общим названием **SNAREs**:
- **синаптобrevин/VAMP** (расположен в мембране везикулы)
  - **синтаксин (syntaxin-1** - в пресинаптической мембране)
  - **SNAP-25** ( в пресинаптической мембране)

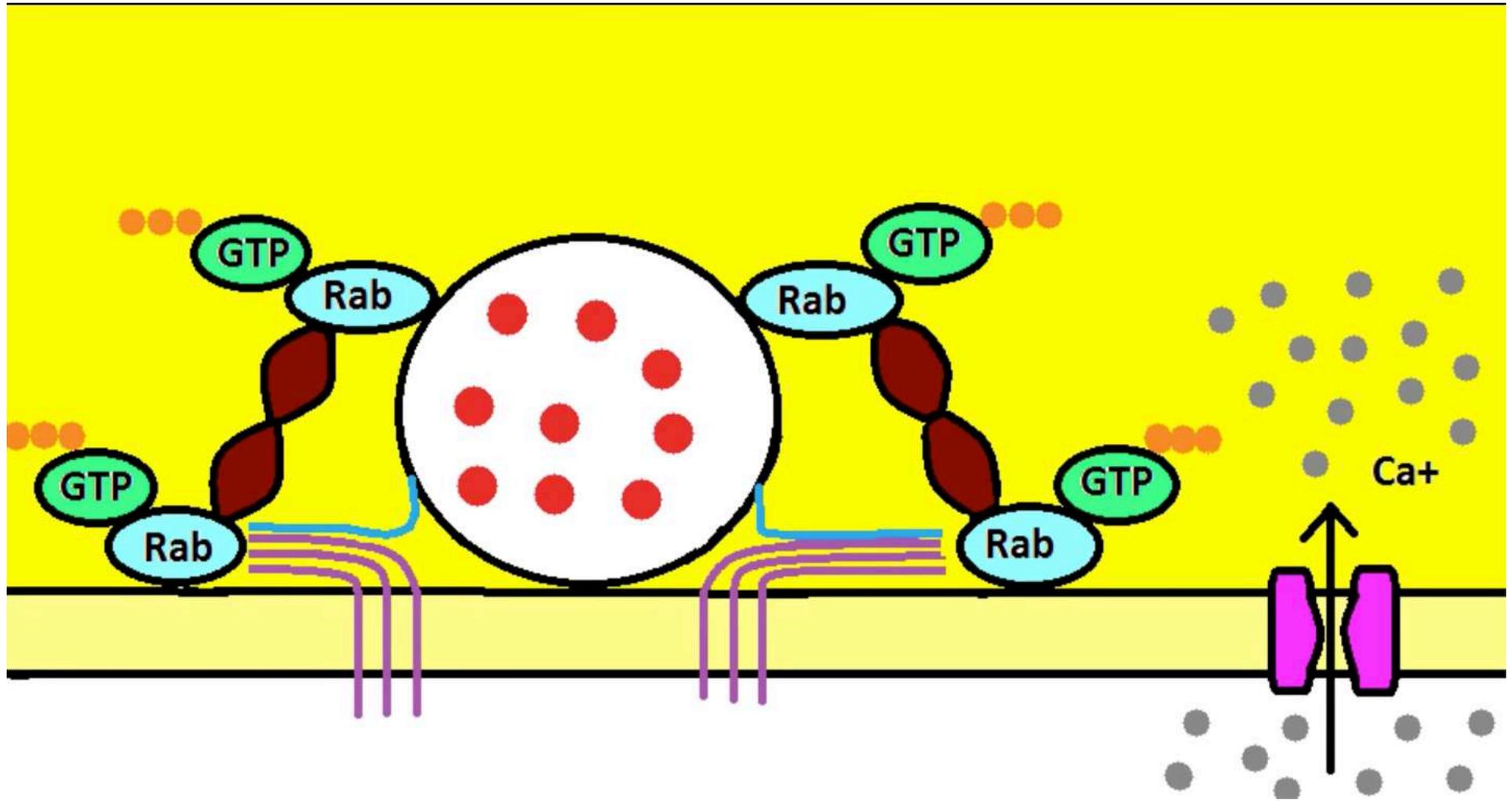


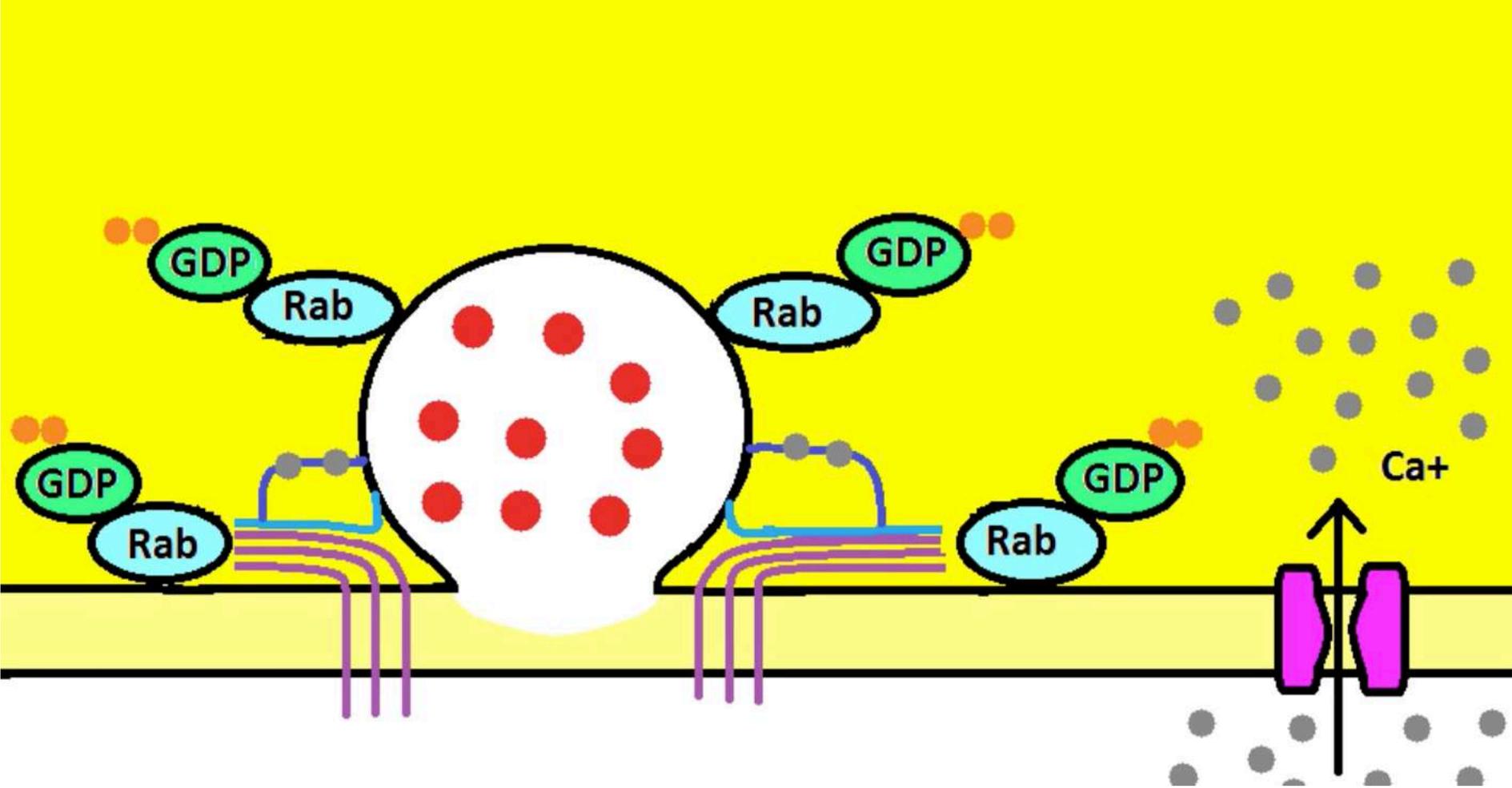


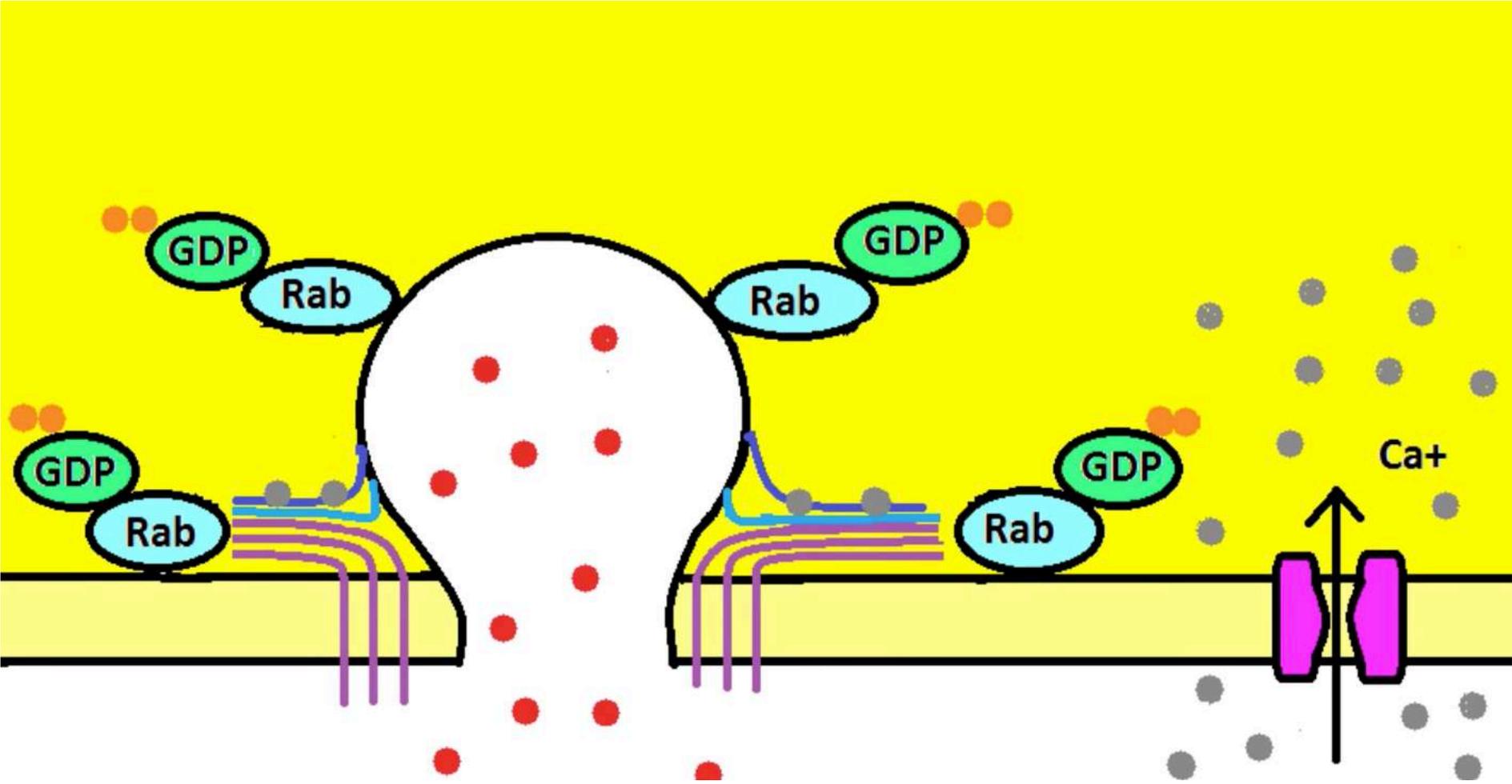


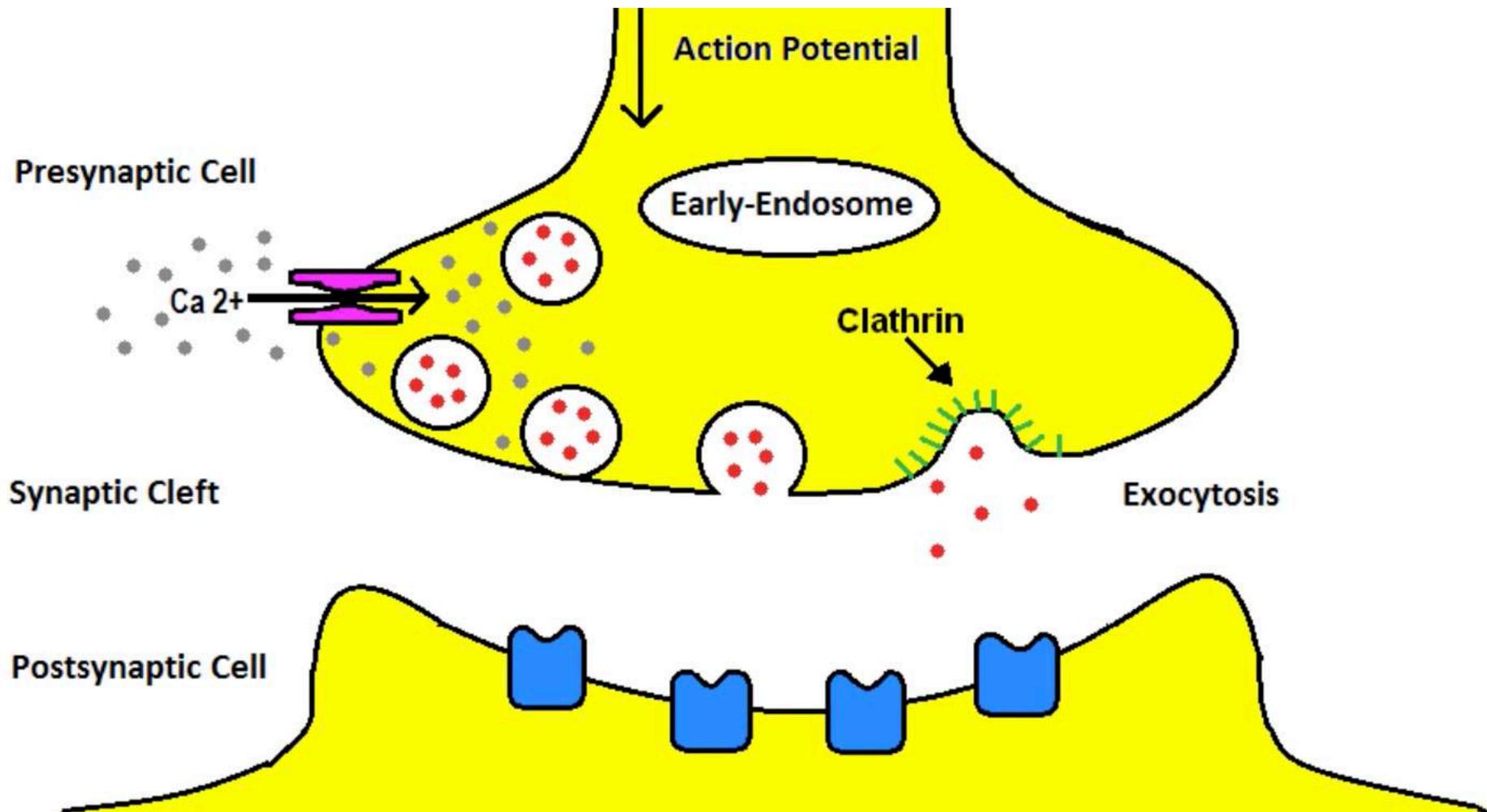
Synaptic Cleft

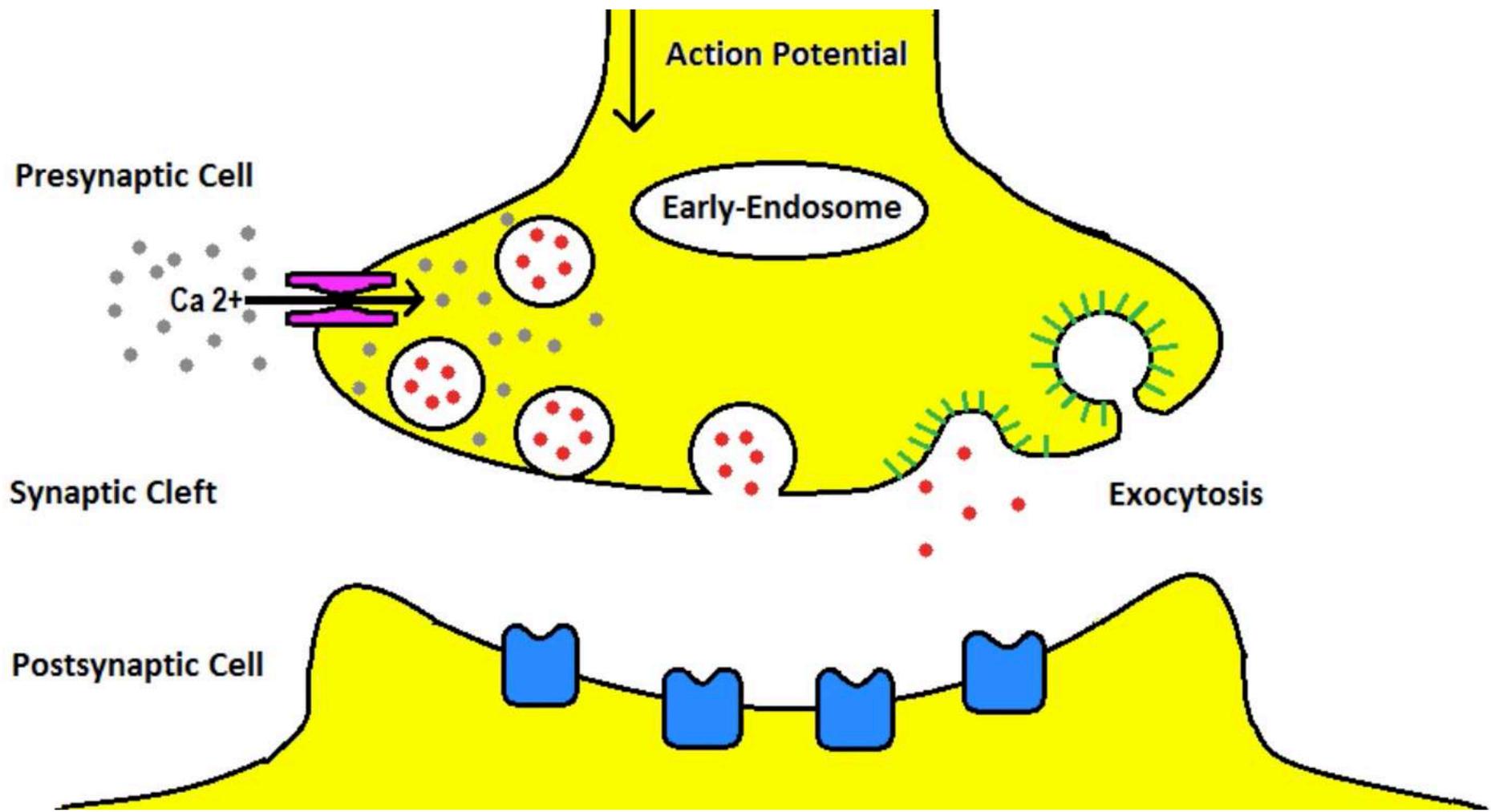


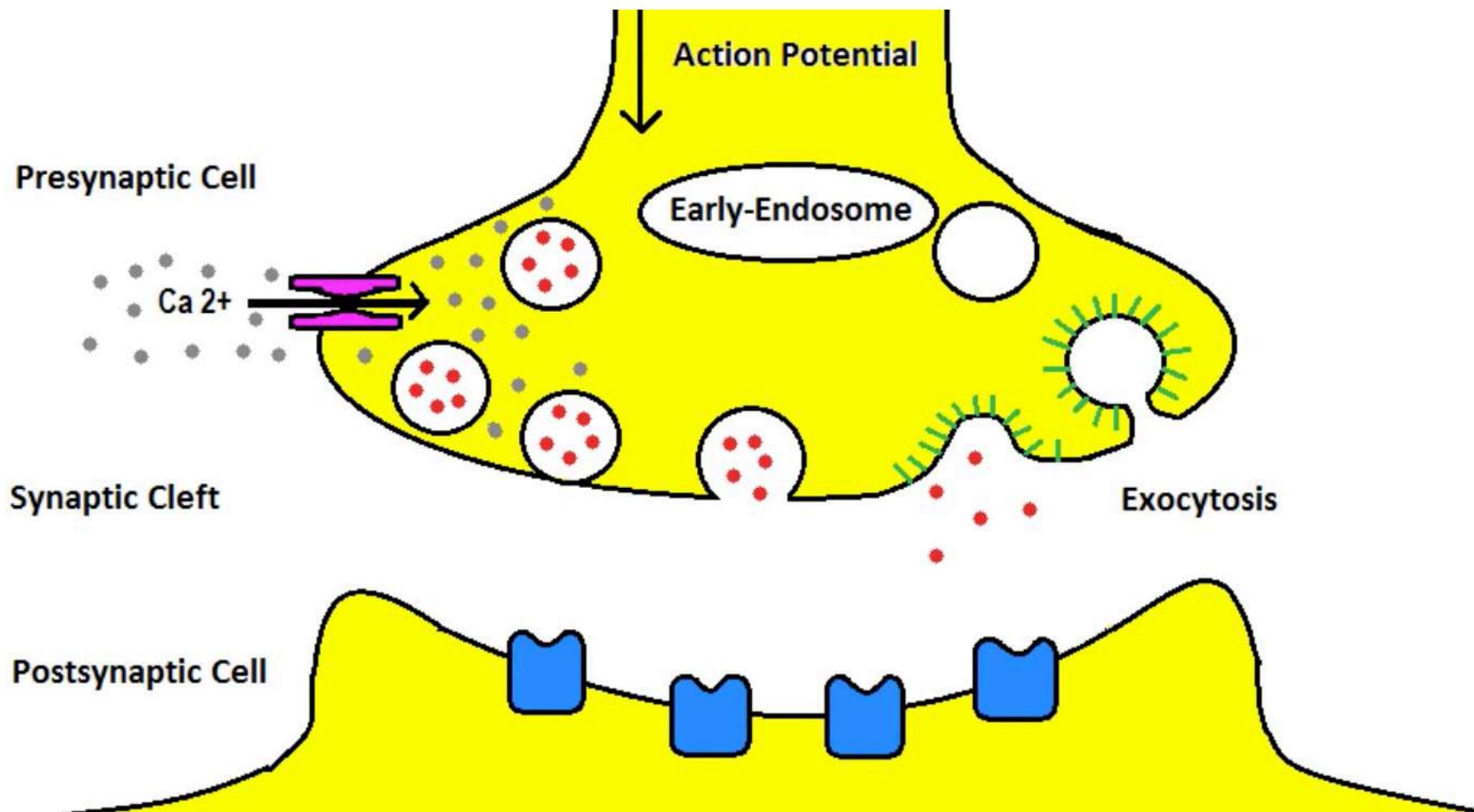


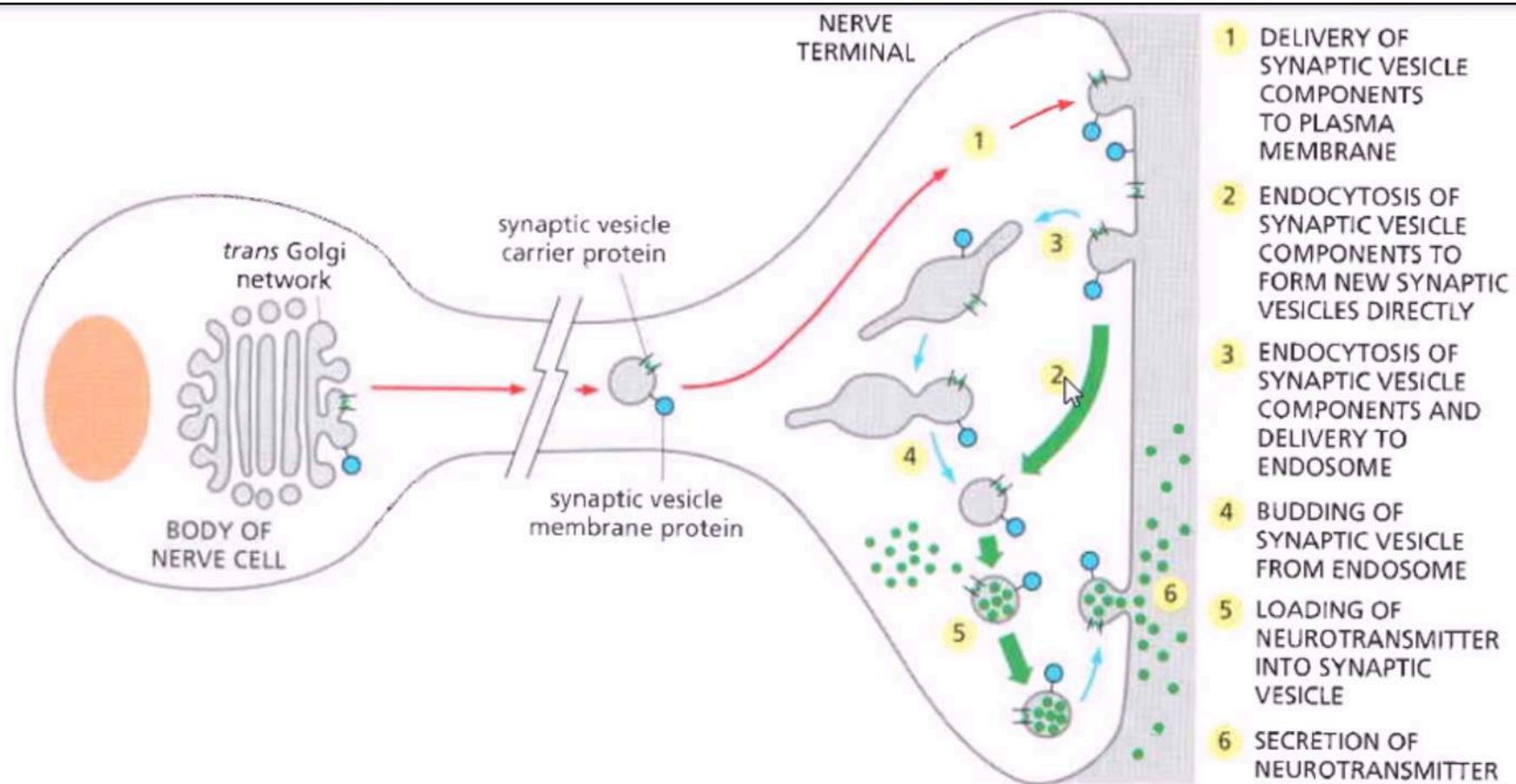




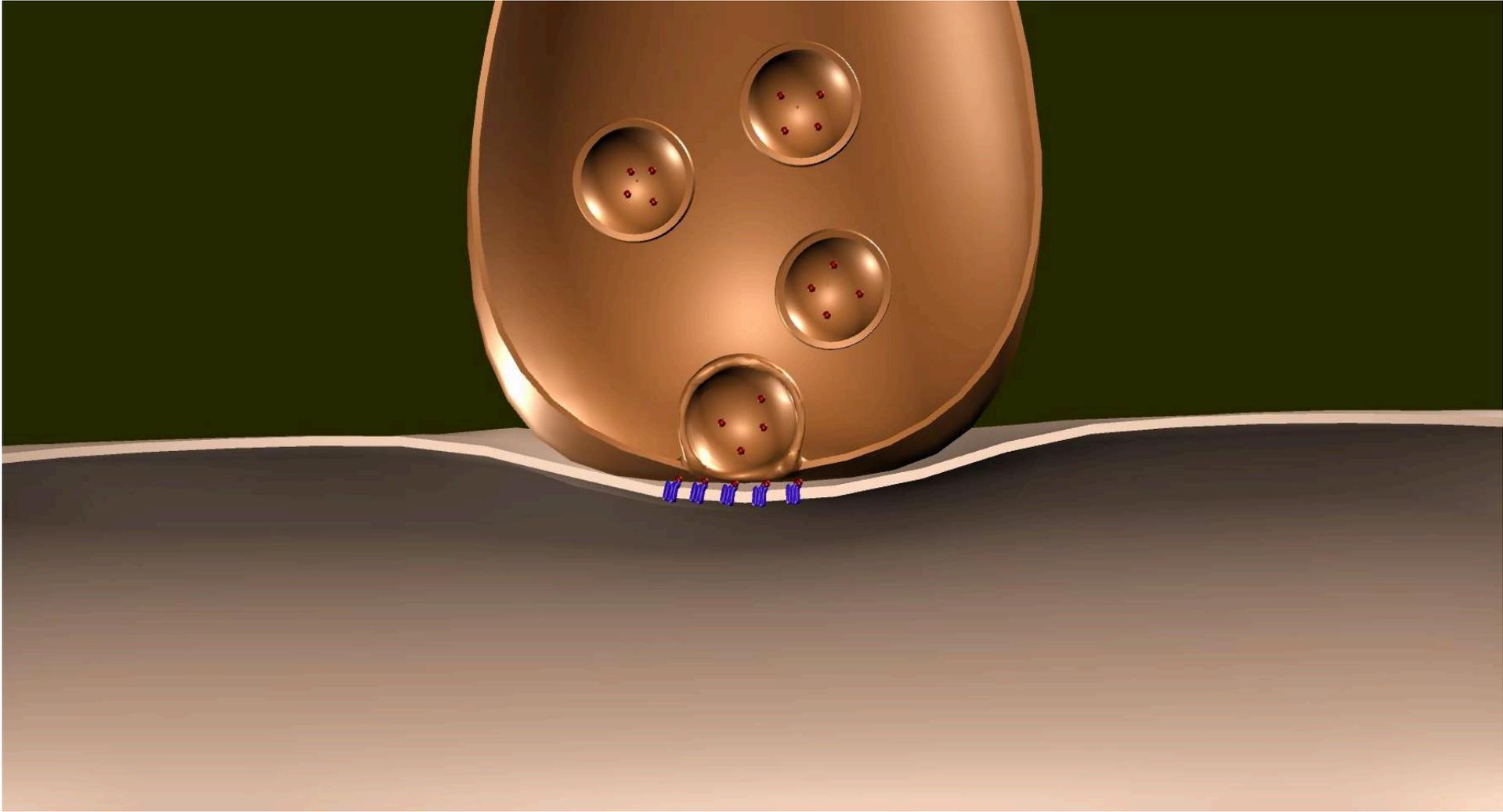




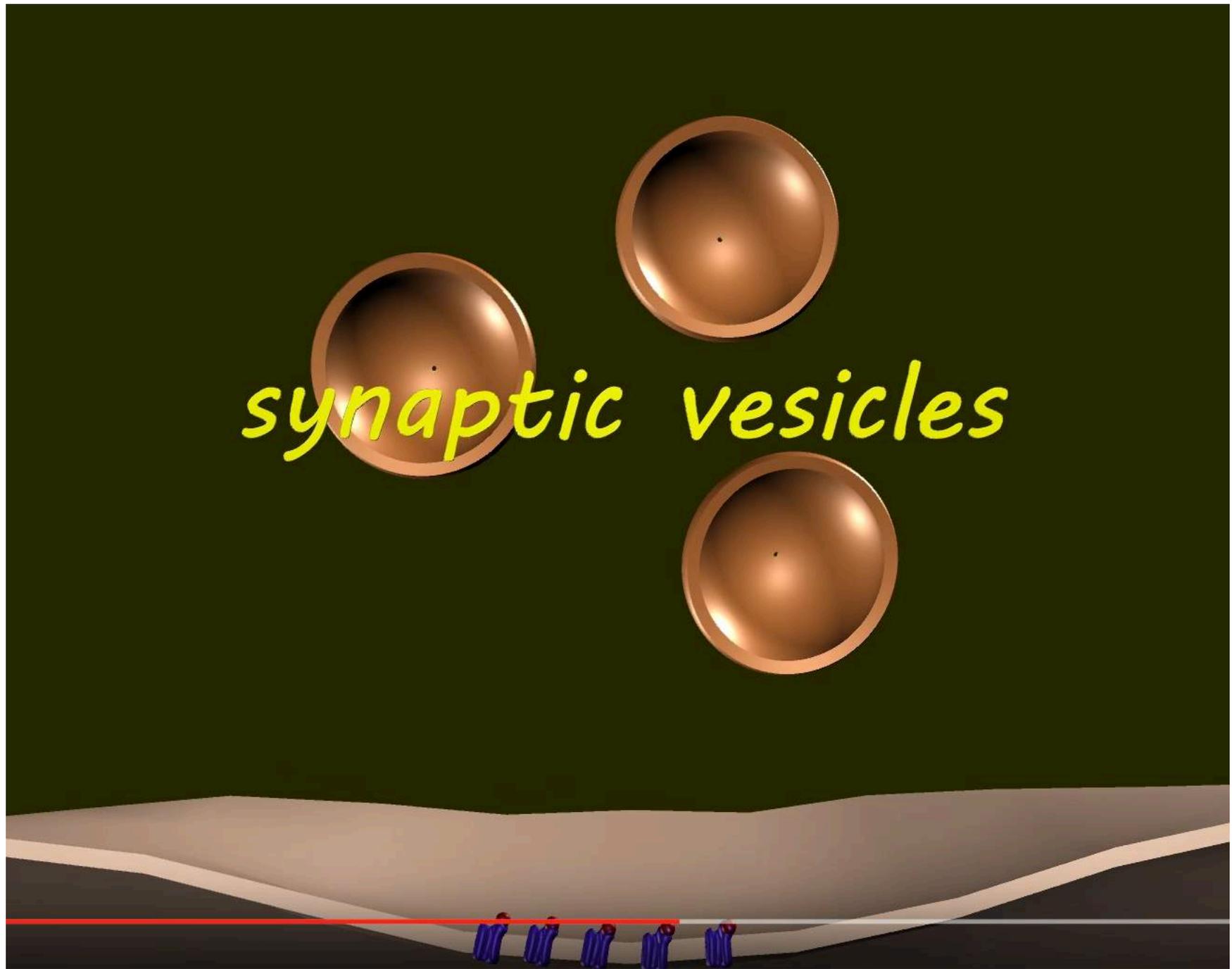


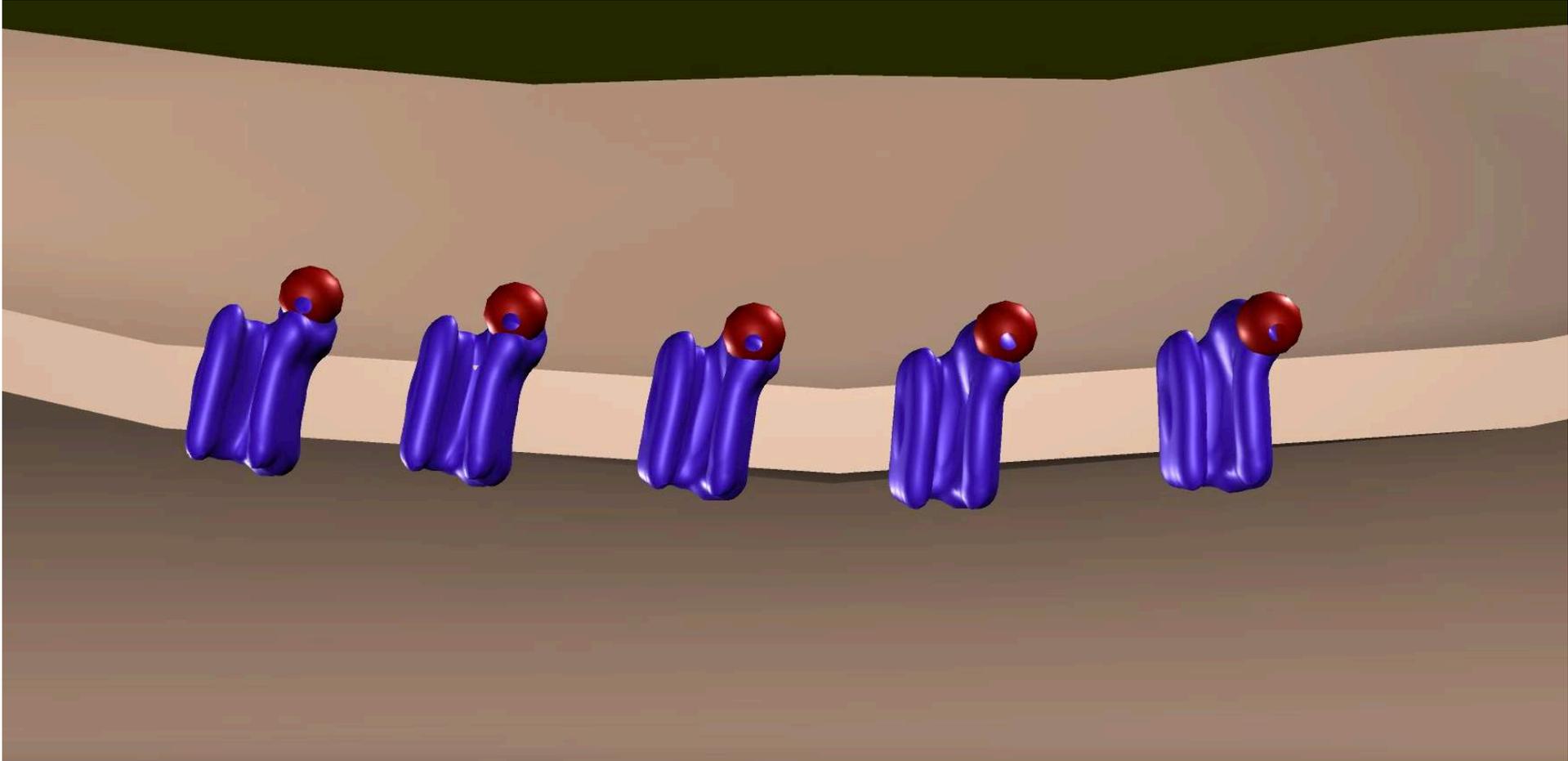


**Figure 13-73 The formation of synaptic vesicles.** These tiny uniform vesicles are found only in nerve cells and in some endocrine cells, where they store and secrete small-molecule neurotransmitters. The import of neurotransmitter directly into the small endocytic vesicles that form from the plasma membrane is mediated by membrane carrier proteins that function as antiports, being driven by a  $H^+$  gradient maintained by proton pumps in the vesicle membrane.



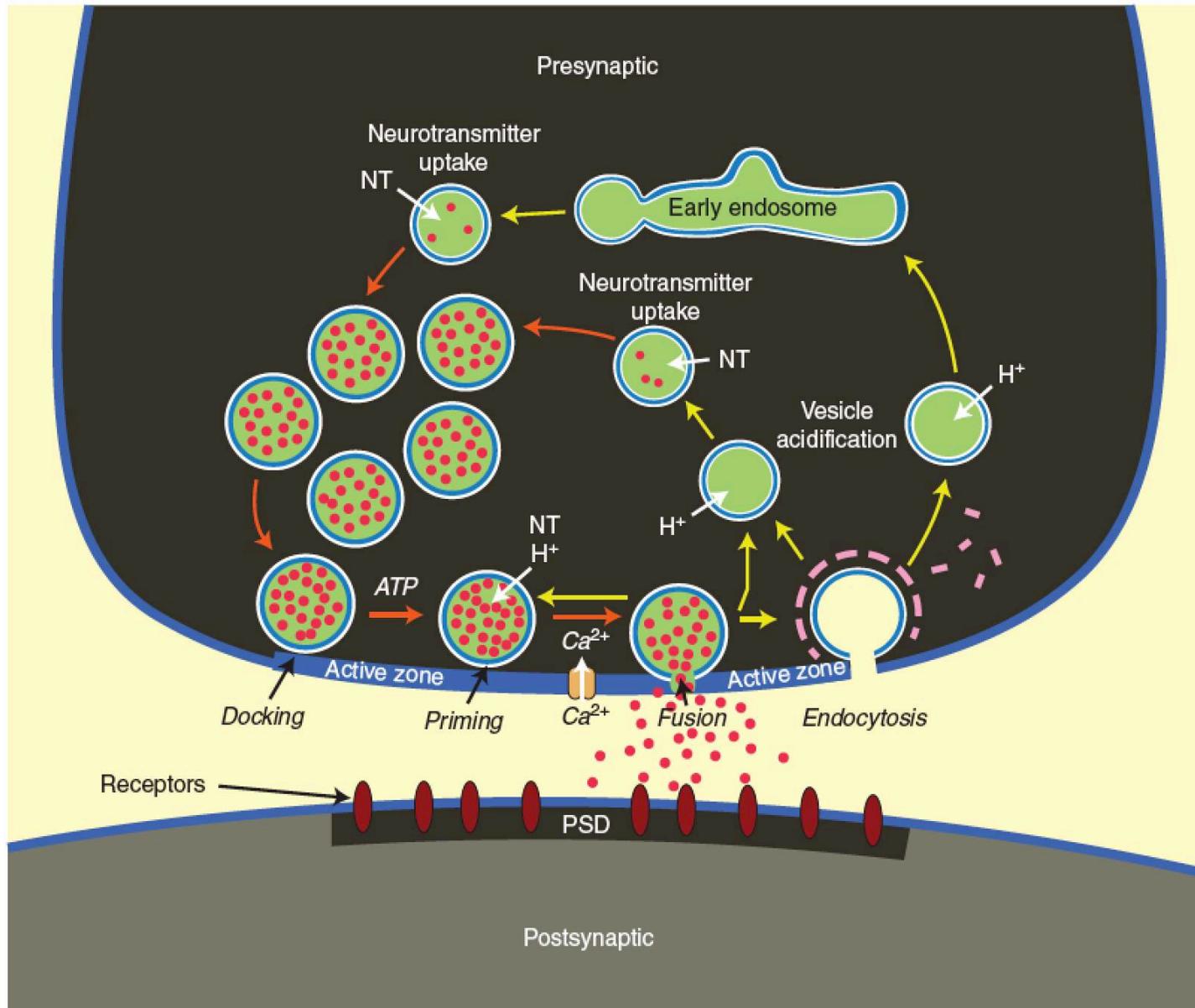
*synaptic vesicles*





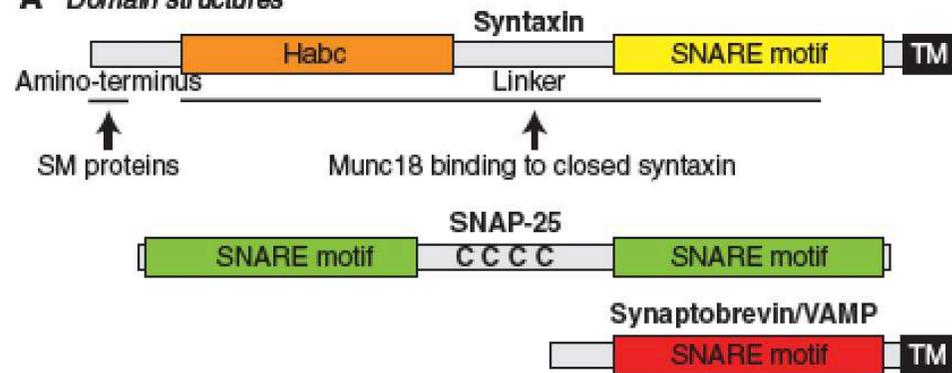




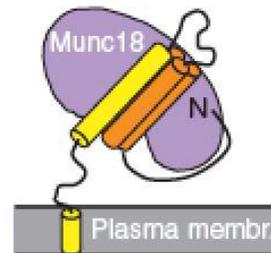




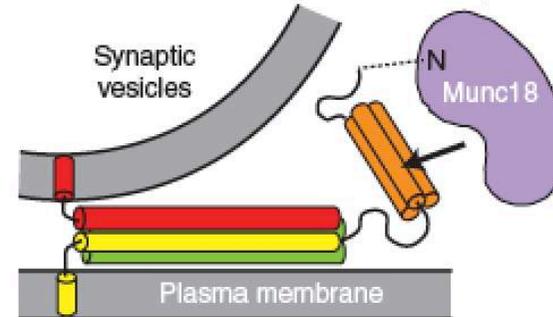
**A Domain structures**



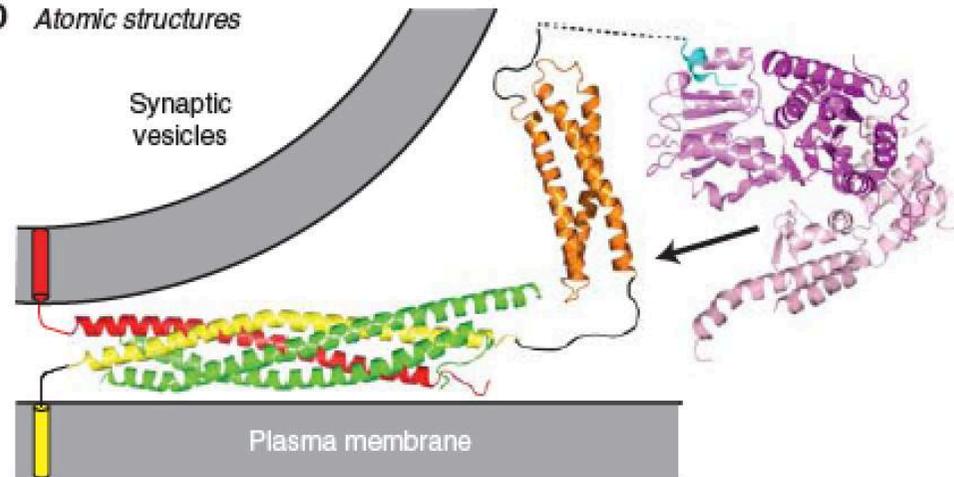
**B Syntaxin/Munc18 heterodimer**

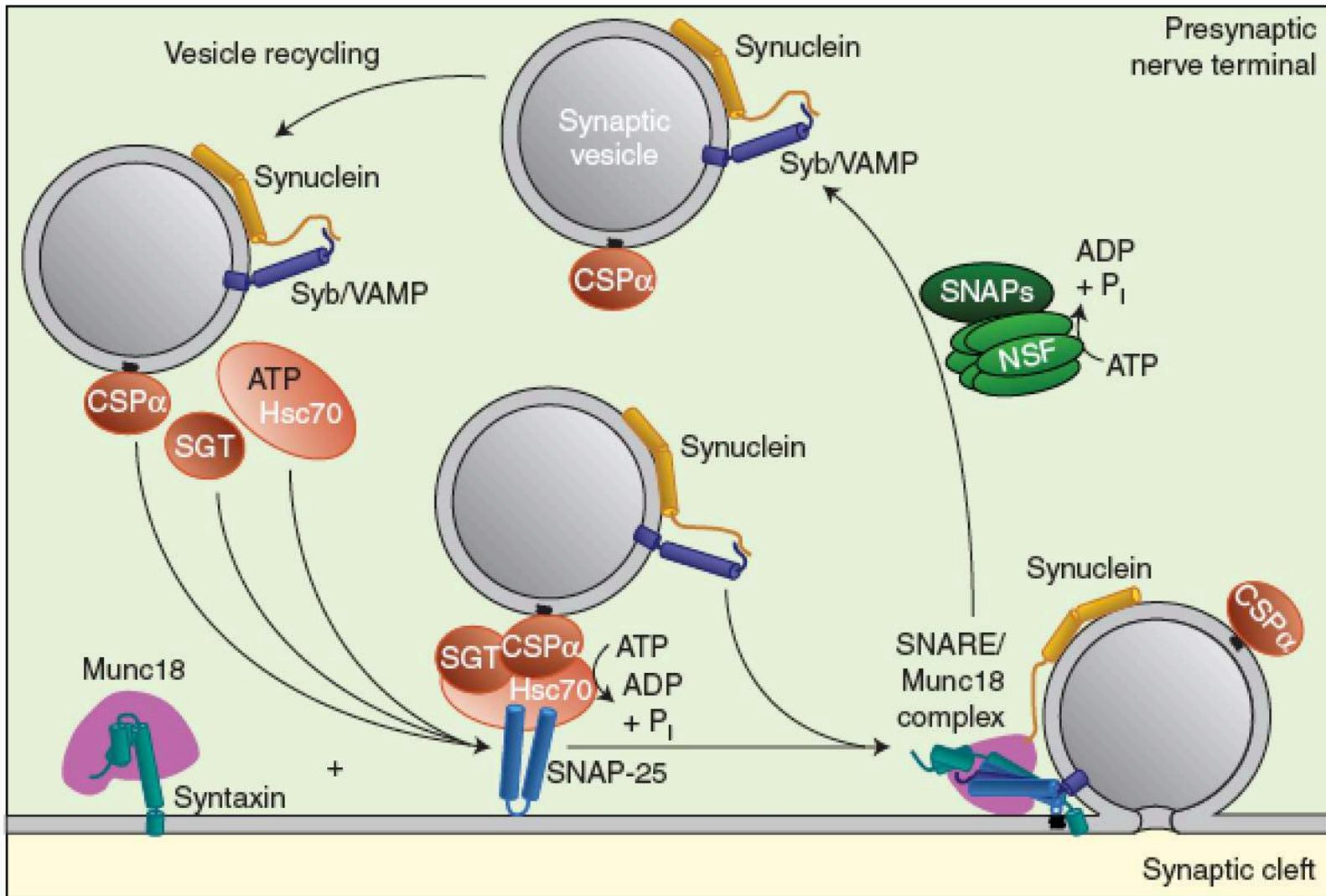


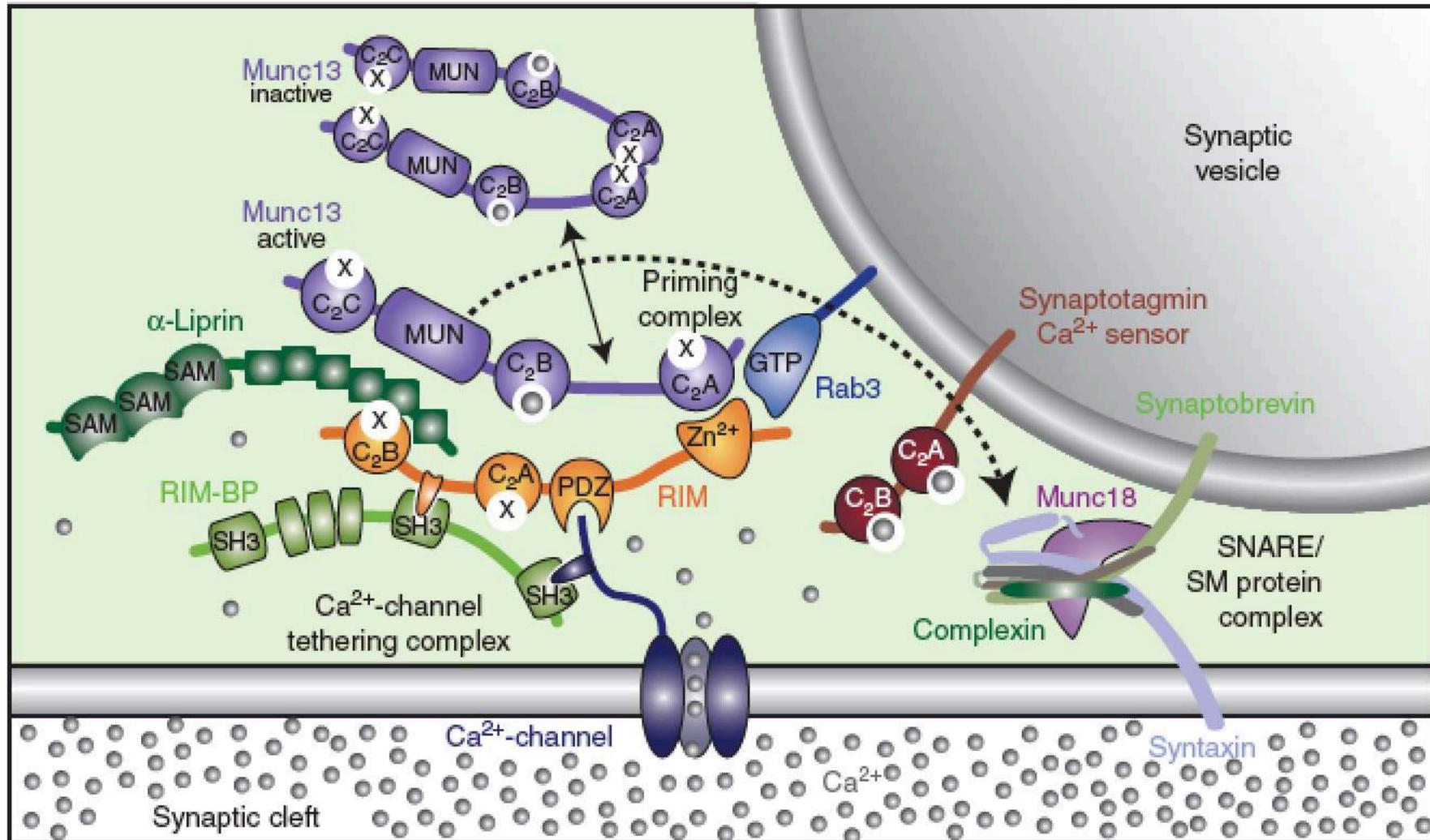
**C SNARE complex/Munc18 assembly**



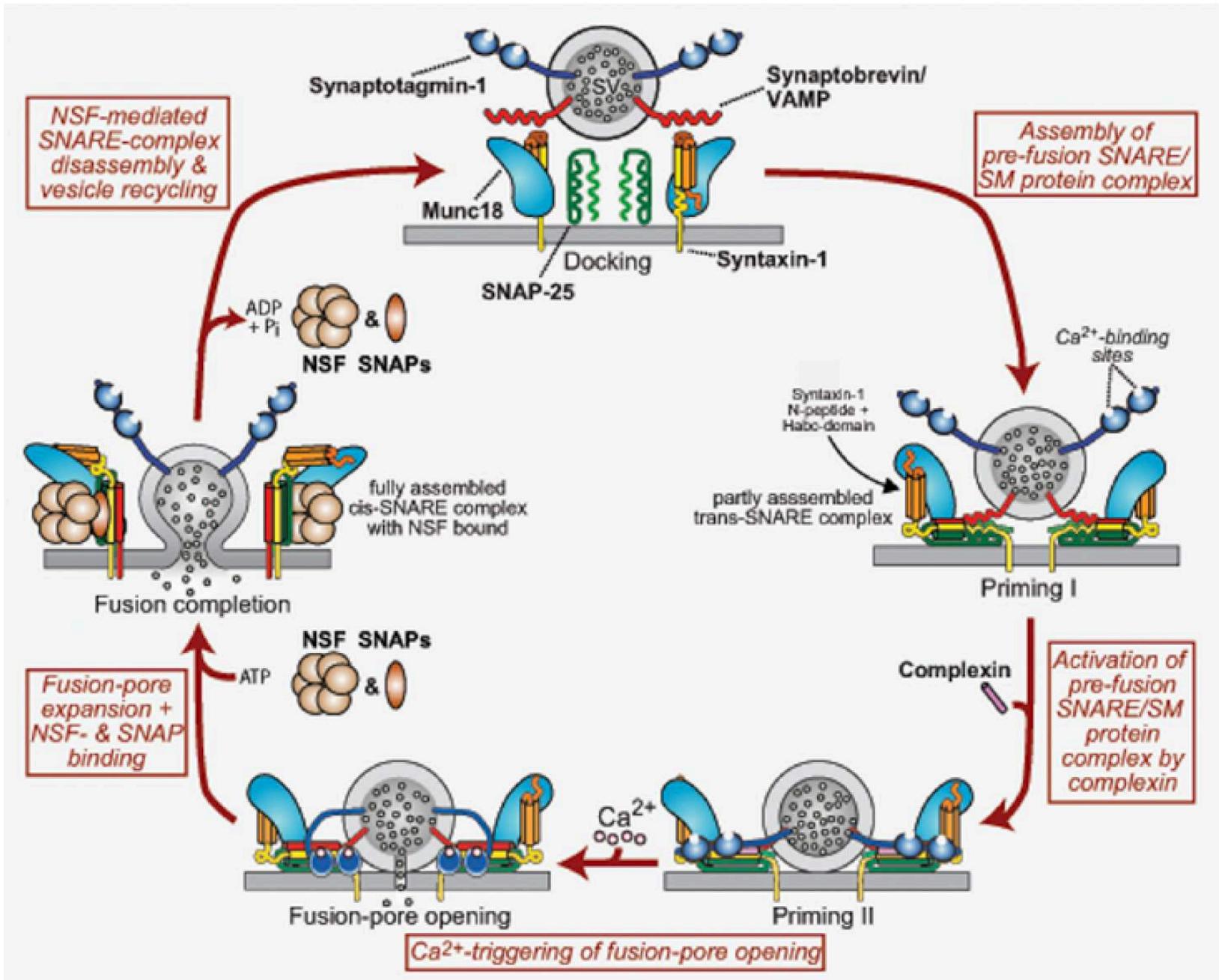
**D Atomic structures**











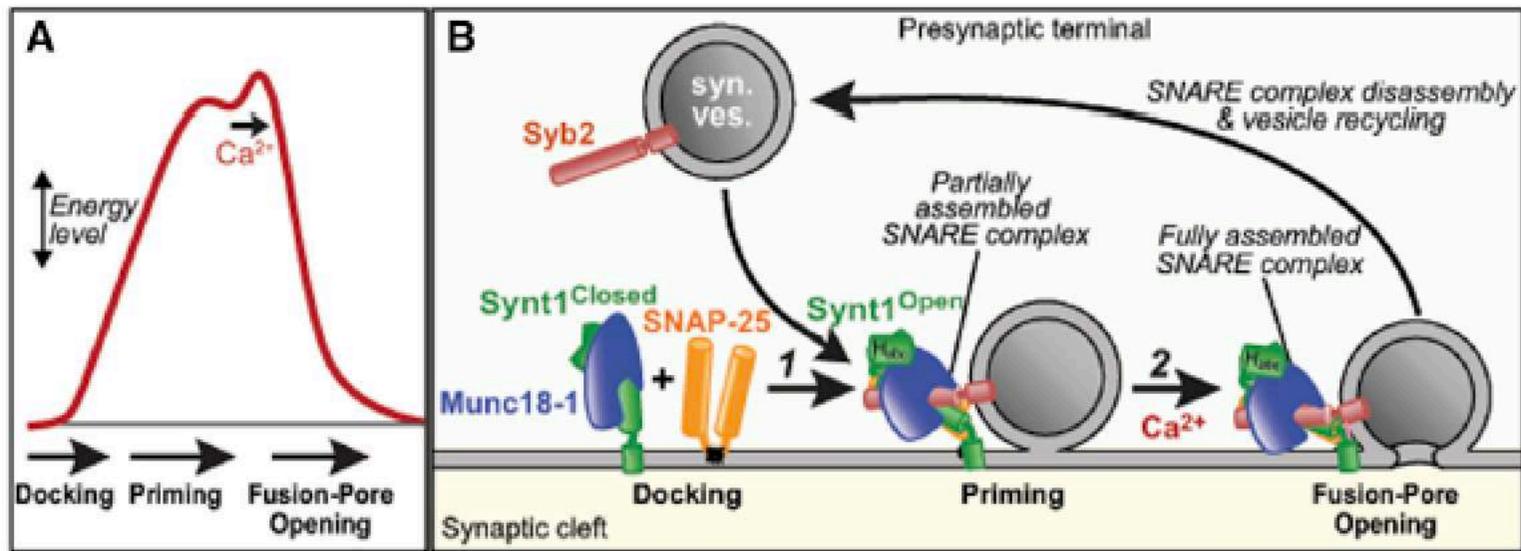
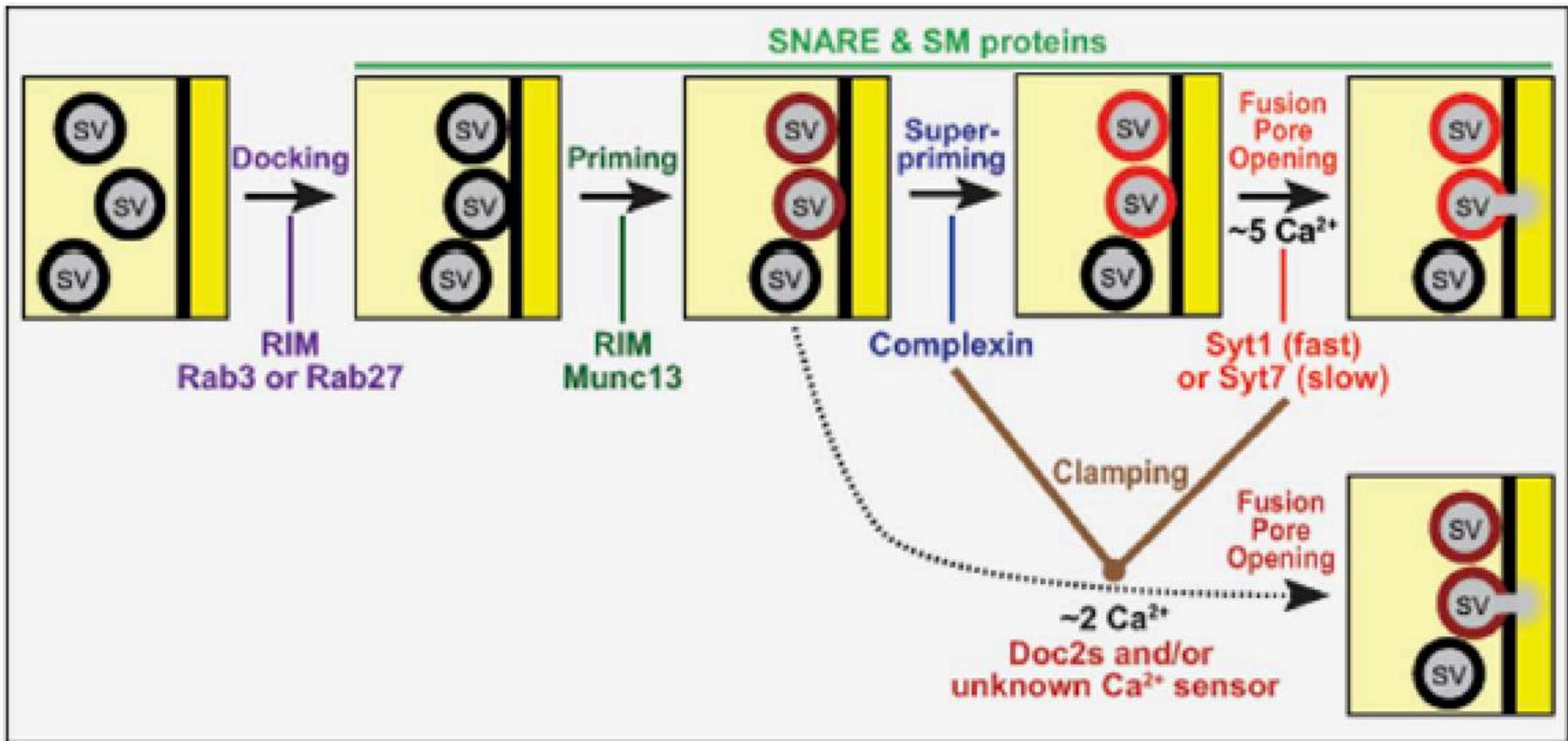
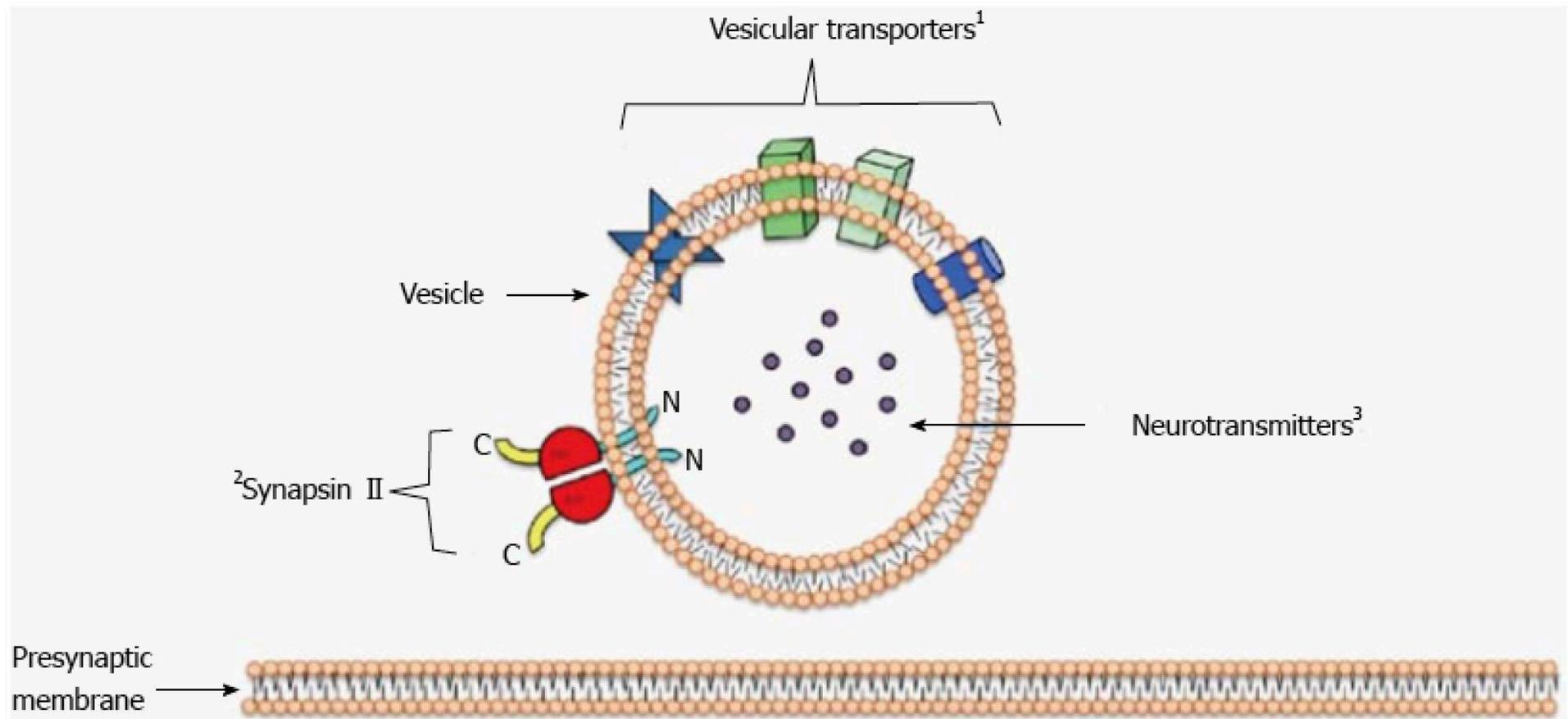


Figure 3. Energy Landscape of Fusion and Proposed Role of the SM Protein Munc18-1 in Promoting Fusion Pore Opening

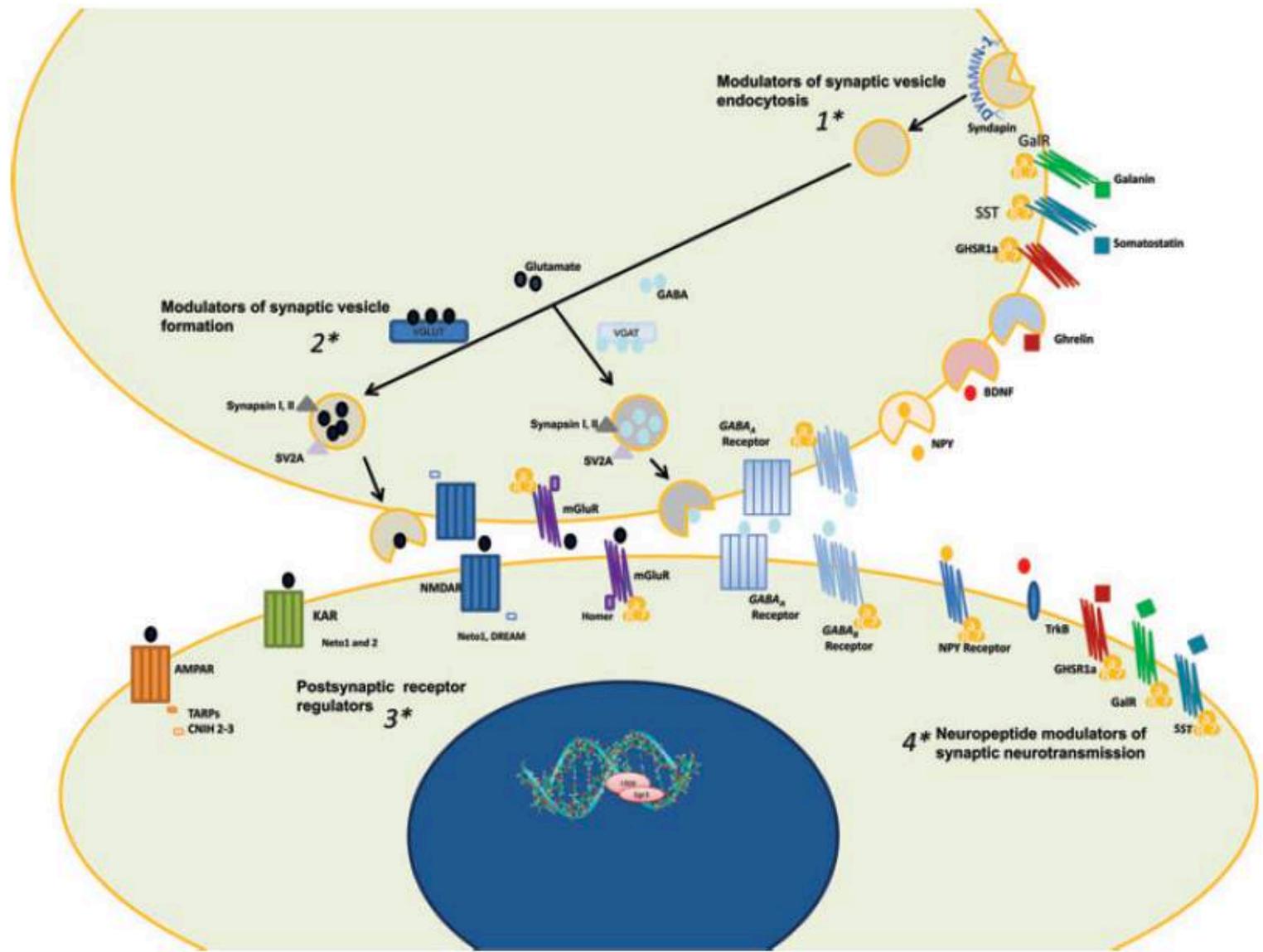
(A) Schematic diagram of the energy level of a vesicle that is docked, primed, and fused. The diagram illustrates that partial SNARE complex assembly during priming is proposed to provide most of the energy required for fusion, such that  $\text{Ca}^{2+}$  triggering only adds a small amount of additional energy to induce fusion pore opening.

(B) Model of Munc18-1 function in fusion. Prior to priming of docked vesicles, Munc18-1 is bound to the closed conformation of syntaxin-1; this interaction is primarily regulatory to maintain a defined rate of entry into the fusion reaction and additionally serves for the mutual stabilization of Munc18-1 and syntaxin-1 for each other (Gerber et al., 2008; Zhou et al., 2013a). Partial SNARE complex assembly during priming (middle) is associated with a dramatic conformational change in syntaxin-1, which has to open, and in Munc18-1, whose binding changes from that to a closed syntaxin-1 conformation to binding to the open syntaxin-1

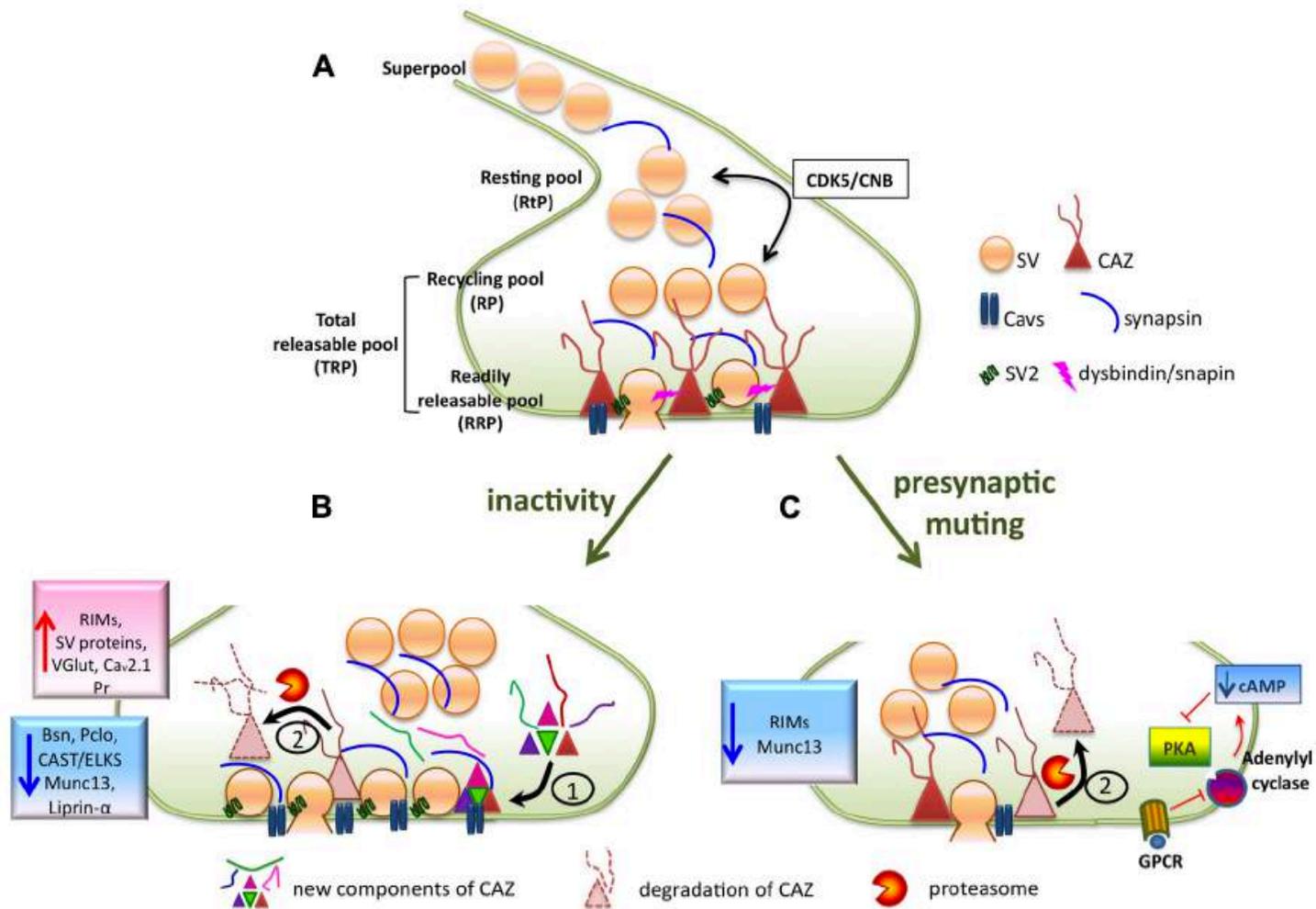








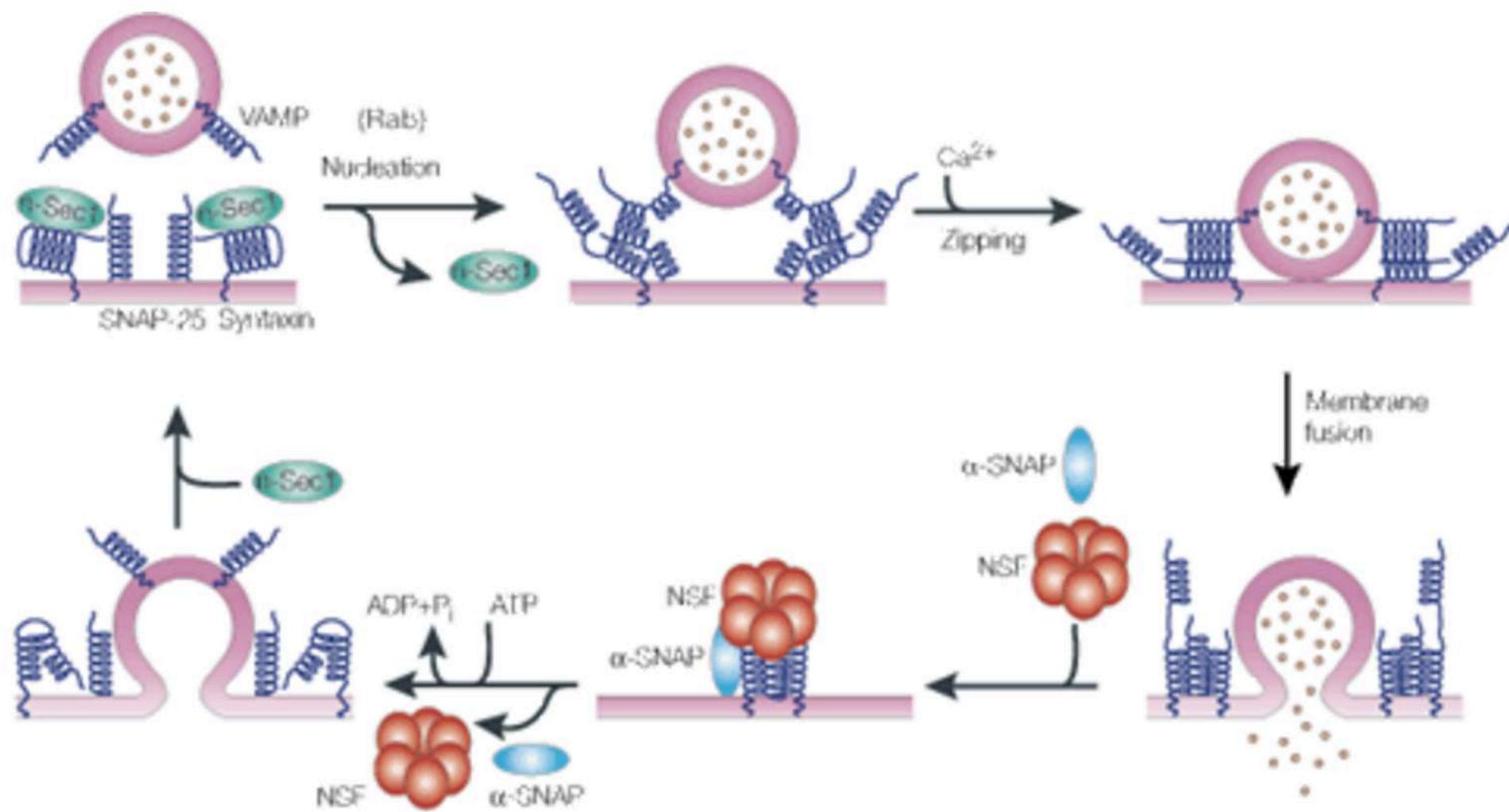




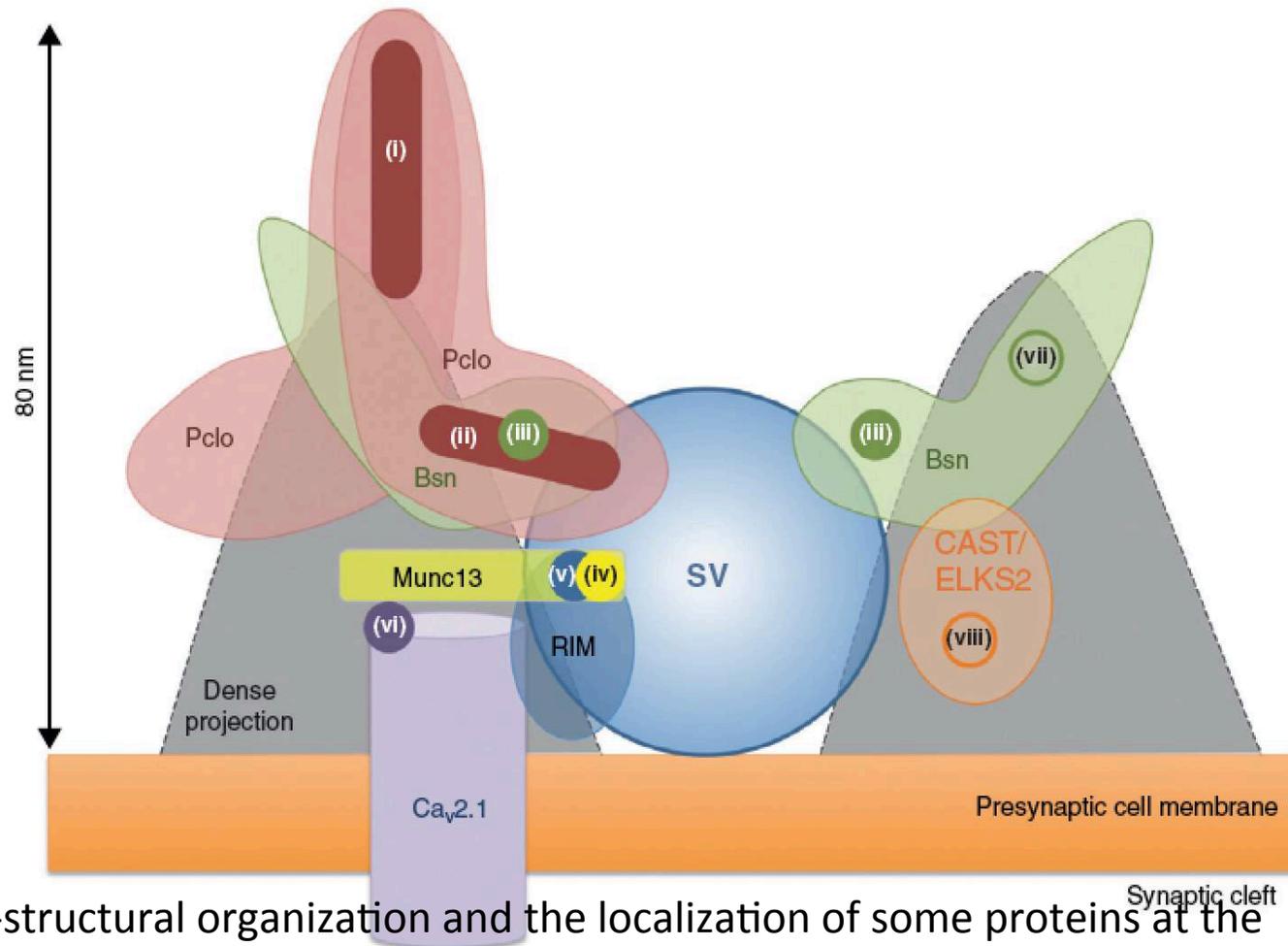


<b>Transmitter Molecule</b>	<b>Derived From</b>	<b>Site of Synthesis</b>
Acetylcholine	Choline	CNS, parasympathetic nerves
Serotonin 5-Hydroxytryptamine (5-HT)	Tryptophan	CNS, chromaffin cells of the gut, enteric cells
GABA	Glutamate	CNS
Glutamate		CNS
Aspartate		CNS
Glycine		spinal cord
Histamine	Histidine	hypothalamus
Epinephrine	Tyrosine	adrenal medulla, some CNS cells
Norpinephrine	Tyrosine	CNS, sympathetic nerves
Dopamine	Tyrosine	CNS
Adenosine	ATP	CNS, periperal nerves

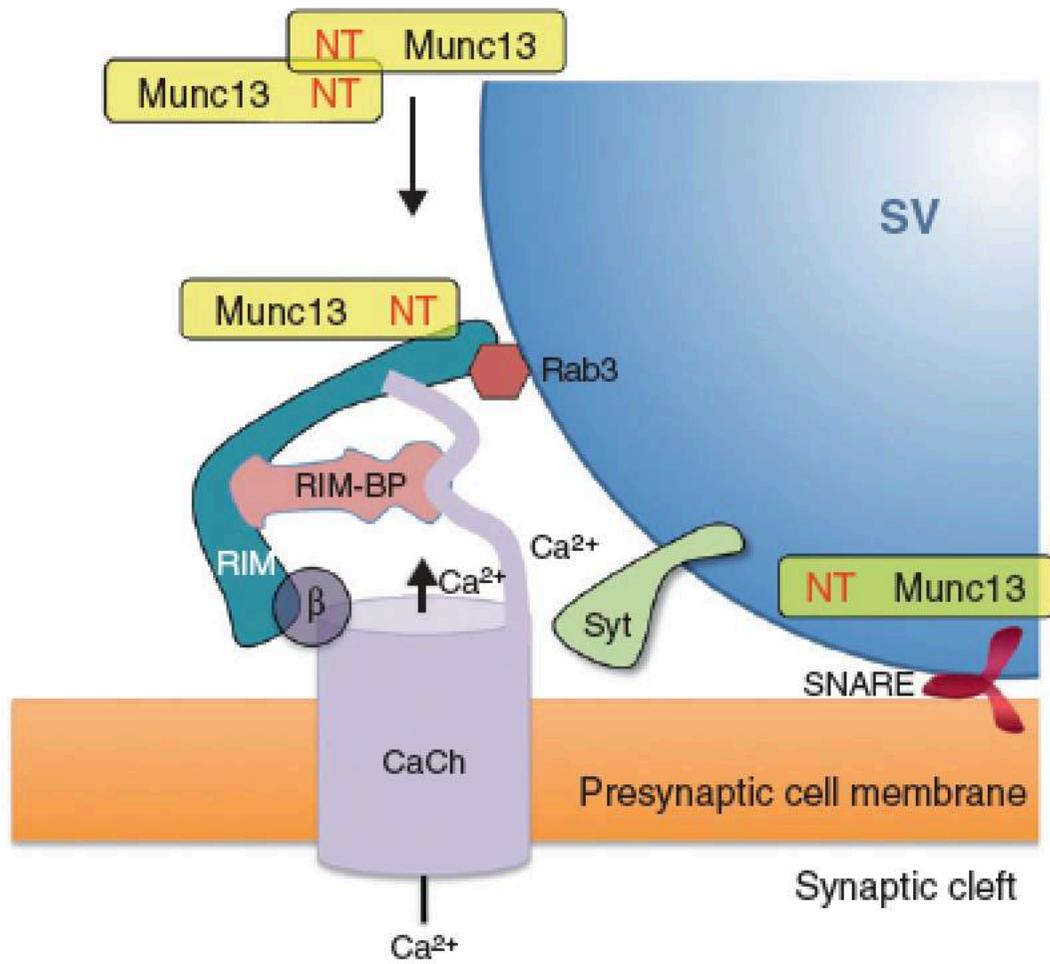
Neurotrans.	Types of receptors	Mode of action	Result in postsynaptic cell	Target
Acetylcholine	Nicotinic Muscarinic	Opens ion channels	EPSP	CNS neurons; skeletal muscle
Serotonin	Two main classes; multiple subclasses	G-protein coupled receptors; both AC and IP3/DAG	Depends on receptor type	Platelet aggregation, smooth muscle contraction, satiety, vomiting
GABA	GABA-A GABA-B	Receptor Cl <sup>-</sup> channel G-linked K <sup>+</sup> channel	IPSP in all cases	Throughout CNS and in retina
Norepinephrine	$\alpha$ Receptor  $\beta$ receptor	G-protein linked to cAMP  G-protein linked to cAMP	IPSP  EPSP	Relaxes smooth muscles of gut, bronchial tree, and vessels to skel. muscle  Increases rate and strength of cardiac contraction; excites smooth muscle in vessels
Dopamine	D1, D2, D3, D4, and D5	G-protein linked to cAMP, direct channel opening, cAMP to K <sup>+</sup> channel opening	EPSP and IPSP	D1-3 are located in the striatum of the CNS, and the basal ganglia D3-5 play a role in mood, psychosis and neuroprotection

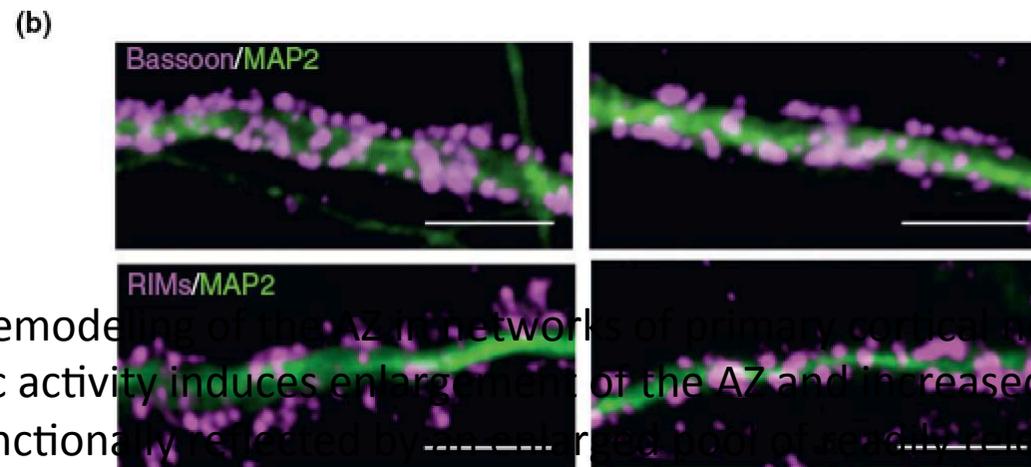
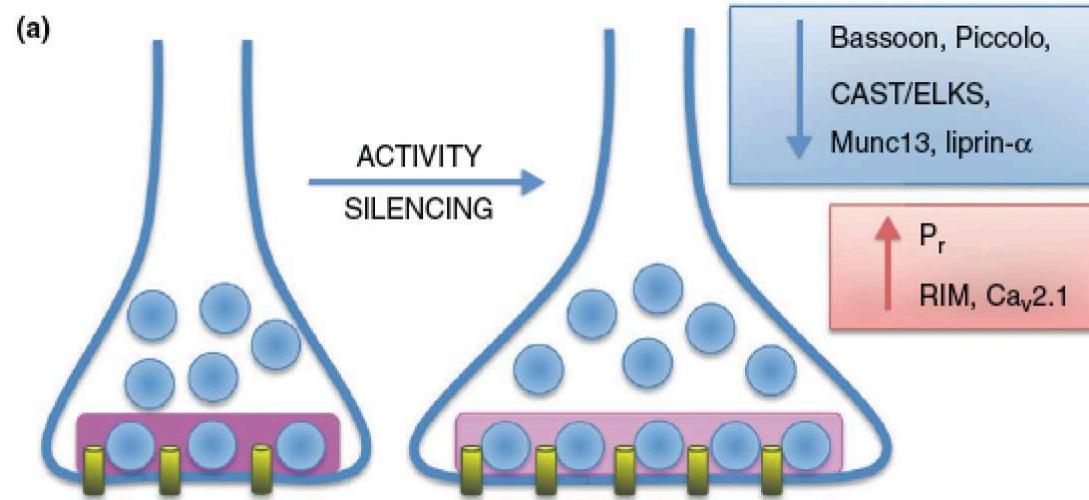






Scheme of the ultra-structural organization and the localization of some proteins at the active zone of conventional brain synapses. (i) Immunogold localization of epitopes against Piccolo/Aczonin (Pclo) N-terminal portion (six epitopes), (ii) C-terminal portion of Pclo (five epitopes), (iii) Bassoon (Bsn) C-terminal portion, (iv) Munc13 N-terminus, (v) RIM N-terminus, (vi) P/Q-type calcium channel Ca<sub>v</sub>2.1, (vii) Bsn N-terminal portion and (viii) CAST/ ELKS2. Note: epitopes (vi)–(viii) and some of (i) and (ii) have only been mapped for their distance from the presynaptic cell membrane and not relative to the dense projections. Epitopes (i)–(vi) were taken from Ref. [17\_\_] and (vii), (viii) from Ref. [21]. Epitope mapping with superresolution light microscopy favorably supports the data for RIM, Bsn and Pclo [20\_].

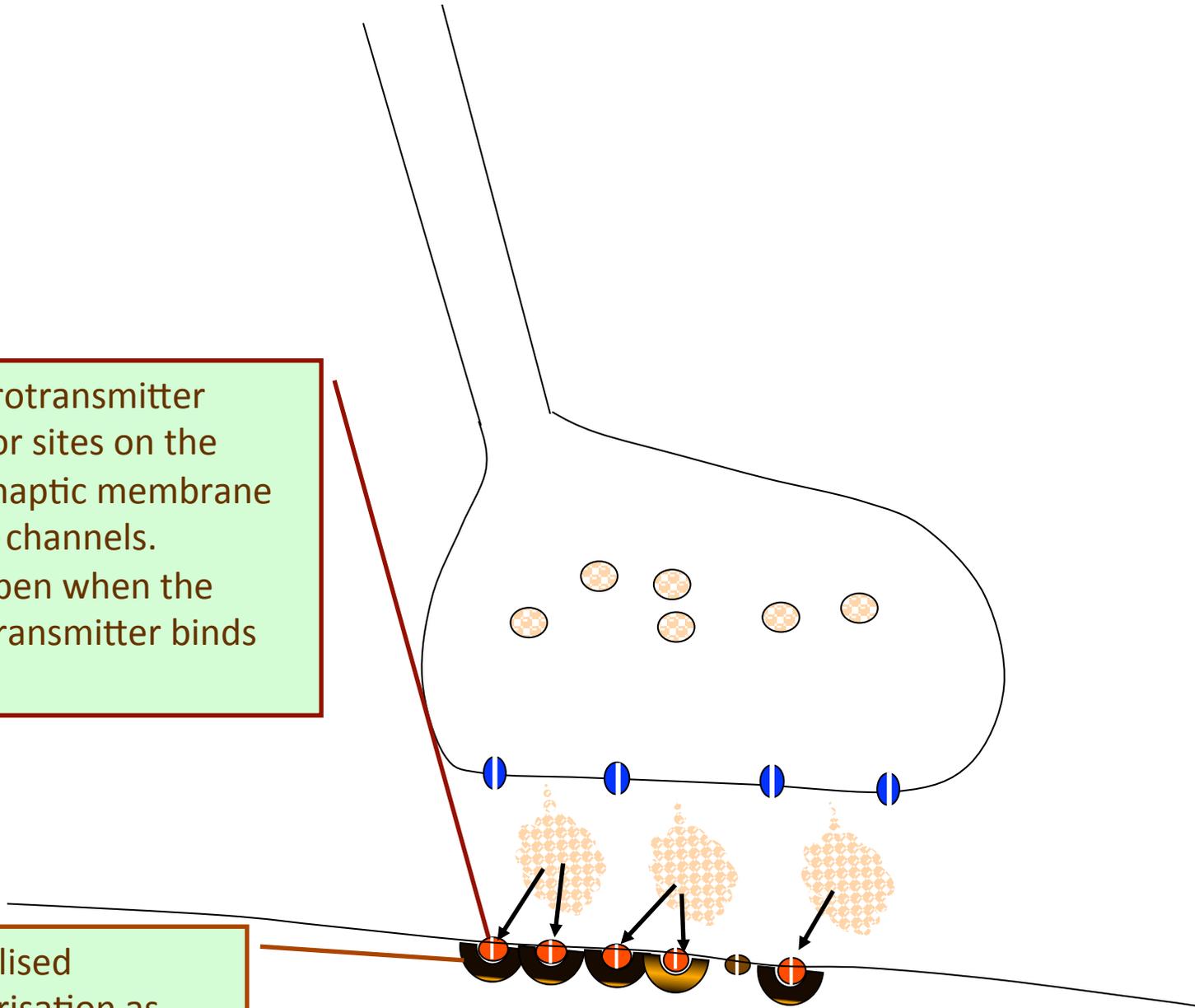




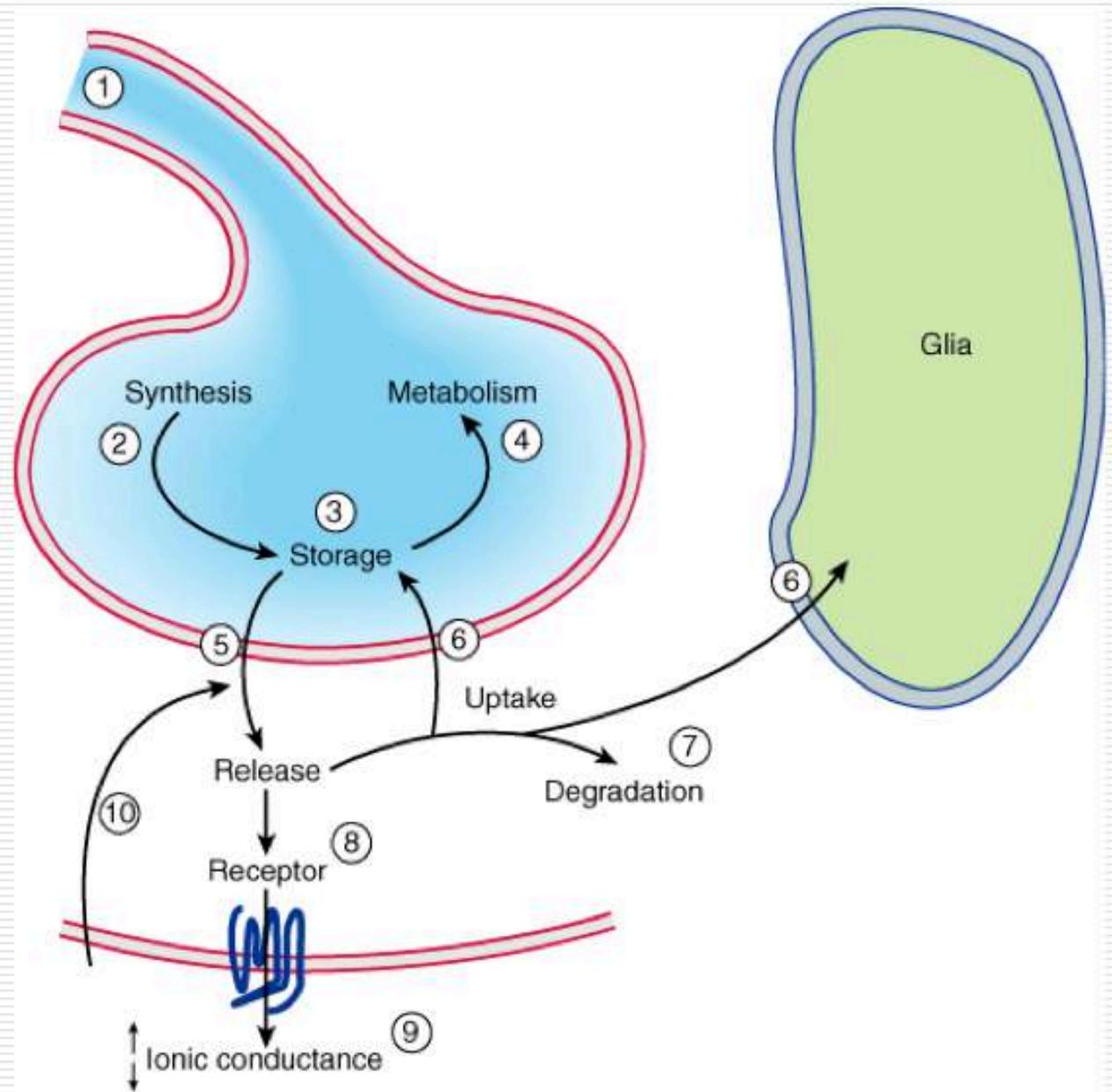
Activity silencing-induced remodeling of the AZ in networks of primary cortical neurons. (a) Chronic silencing of synaptic activity induces enlargement of the AZ and increased number of docked vesicles, which is functionally reflected by an enlarged pool of readily-releasable vesicles and higher synaptic release probability ( $P_r$ ) [58]. This structural and functional remodeling is associated with the reduction of most CAZ proteins in synapses, the redistribution of RIM to the most active synapses and an all-over increase of synaptic P/Q-type CaV2.1 channels. (b) Examples of immunostaining showing depletion of Bassoon and redistribution of RIM at synapses of proximal dendrites of rat cortical neurons (visualized by anti-MAP2 staining) upon 48-h pharmacological inhibition of glutamate receptors using D-AP5 and CNQX (for details see [60\_]). Scale bar, 5  $\mu$ m.

**5.** Neurotransmitter receptor sites on the postsynaptic membrane are ion channels. They open when the neurotransmitter binds

**6.** Localised depolarisation as ions leak in or out of membrane.

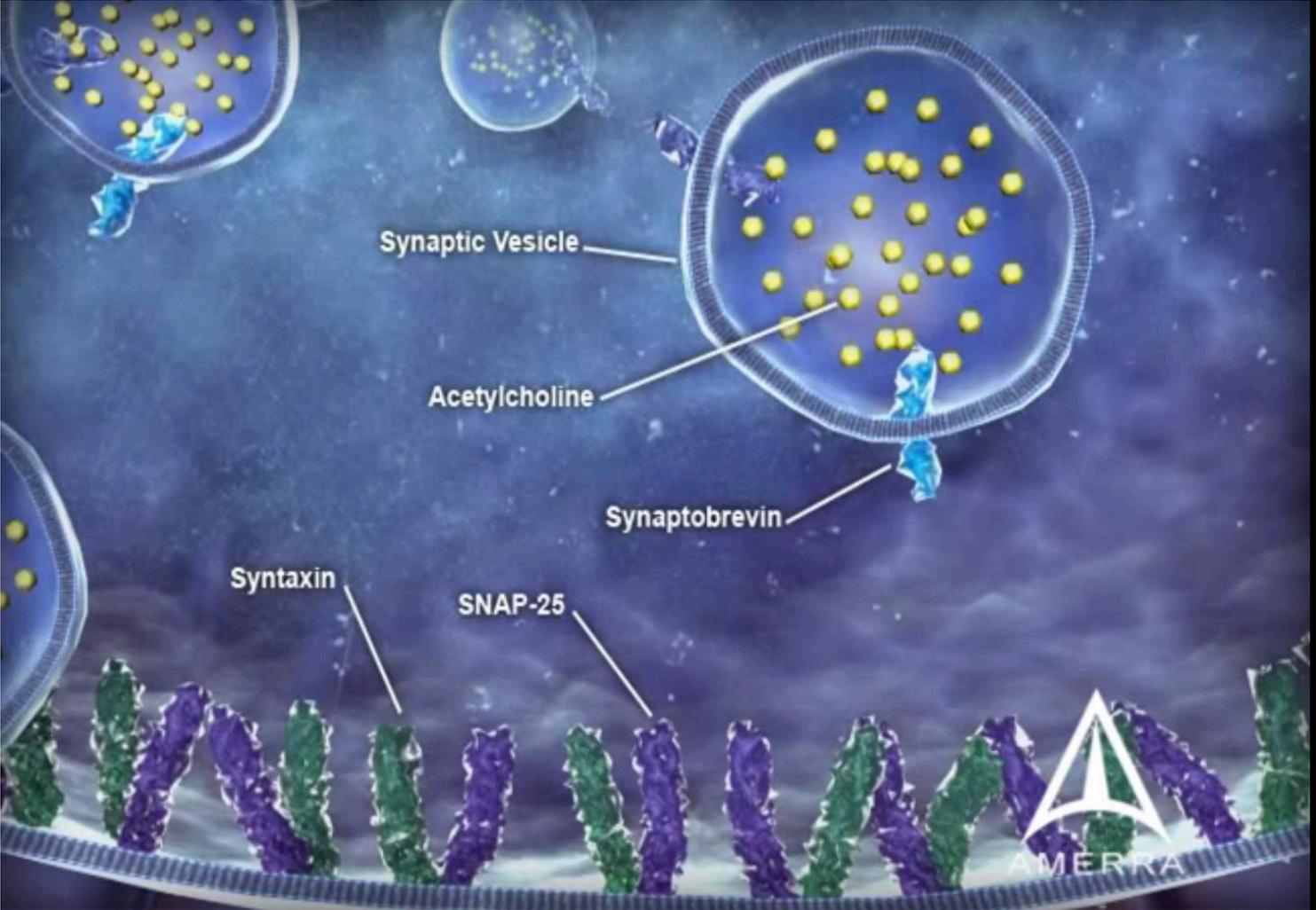


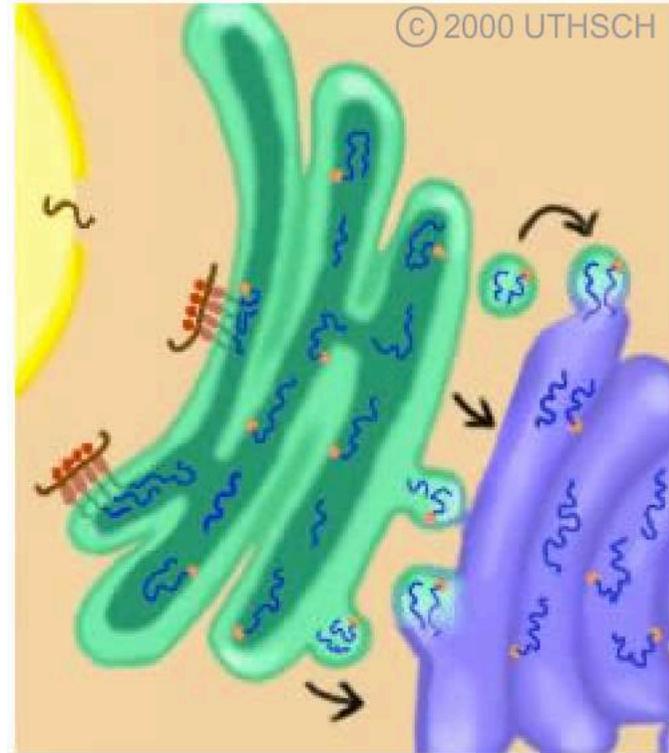
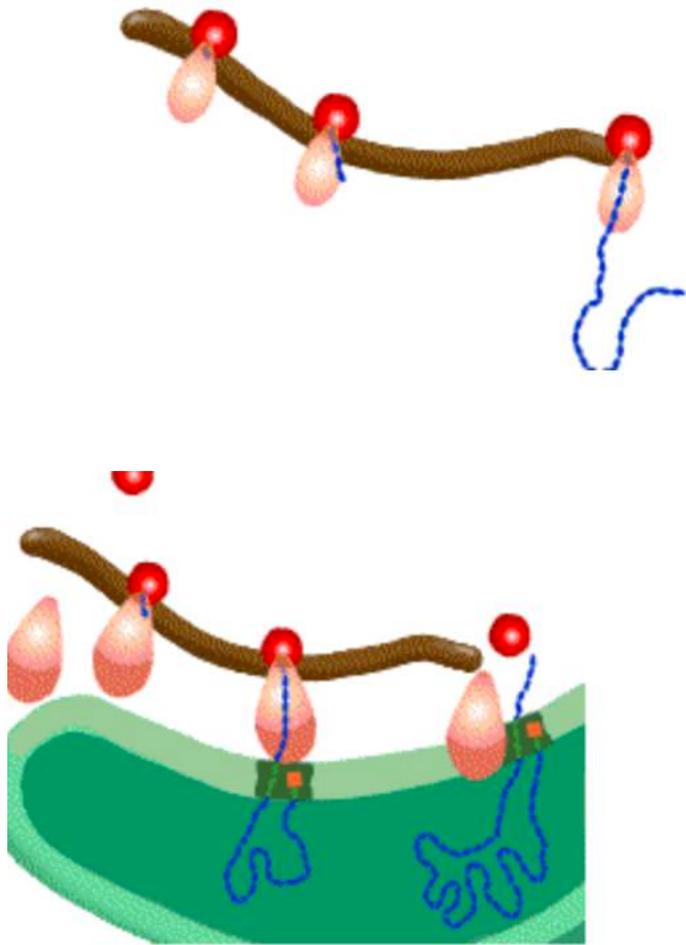
# Site of drug action

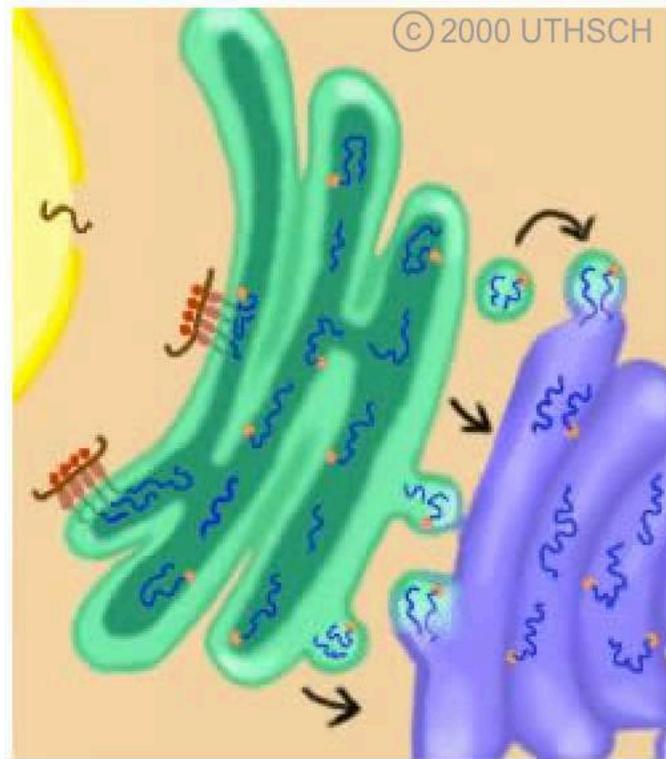


Source: Katzung BG, Masters SB, Trevor AJ; *Basic & Clinical Pharmacology*, 11th Edition; <http://www.accessmedicine.com>

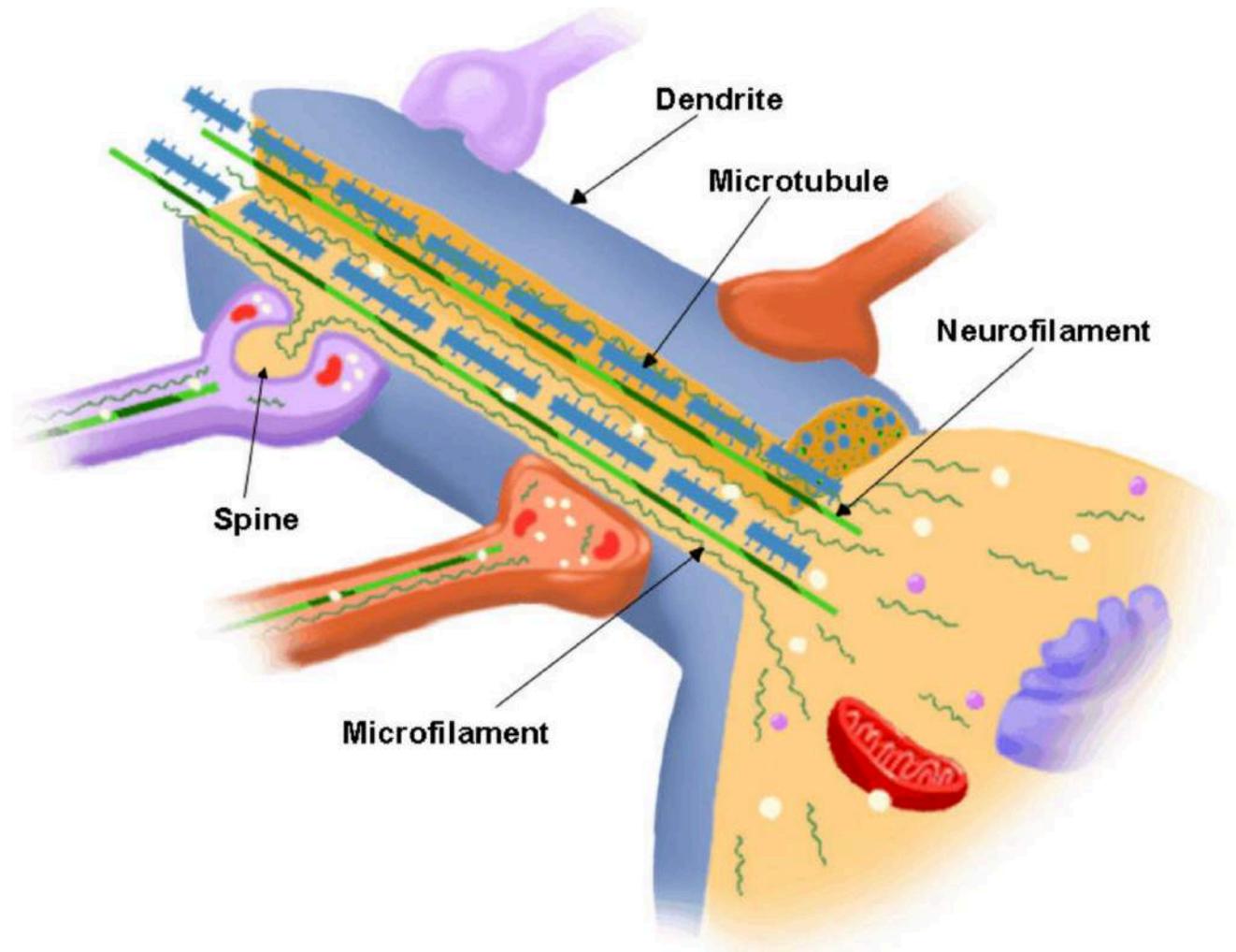
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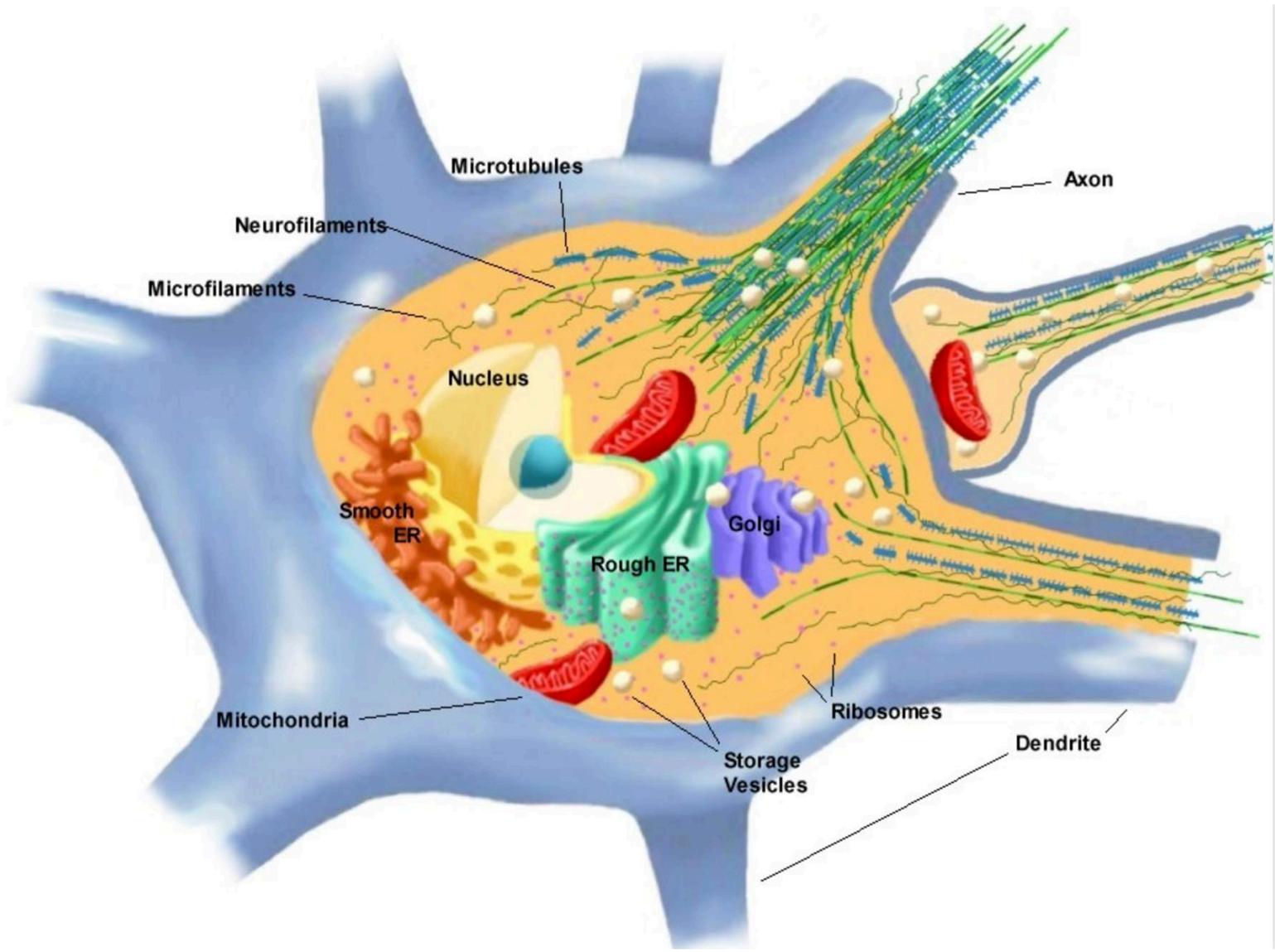


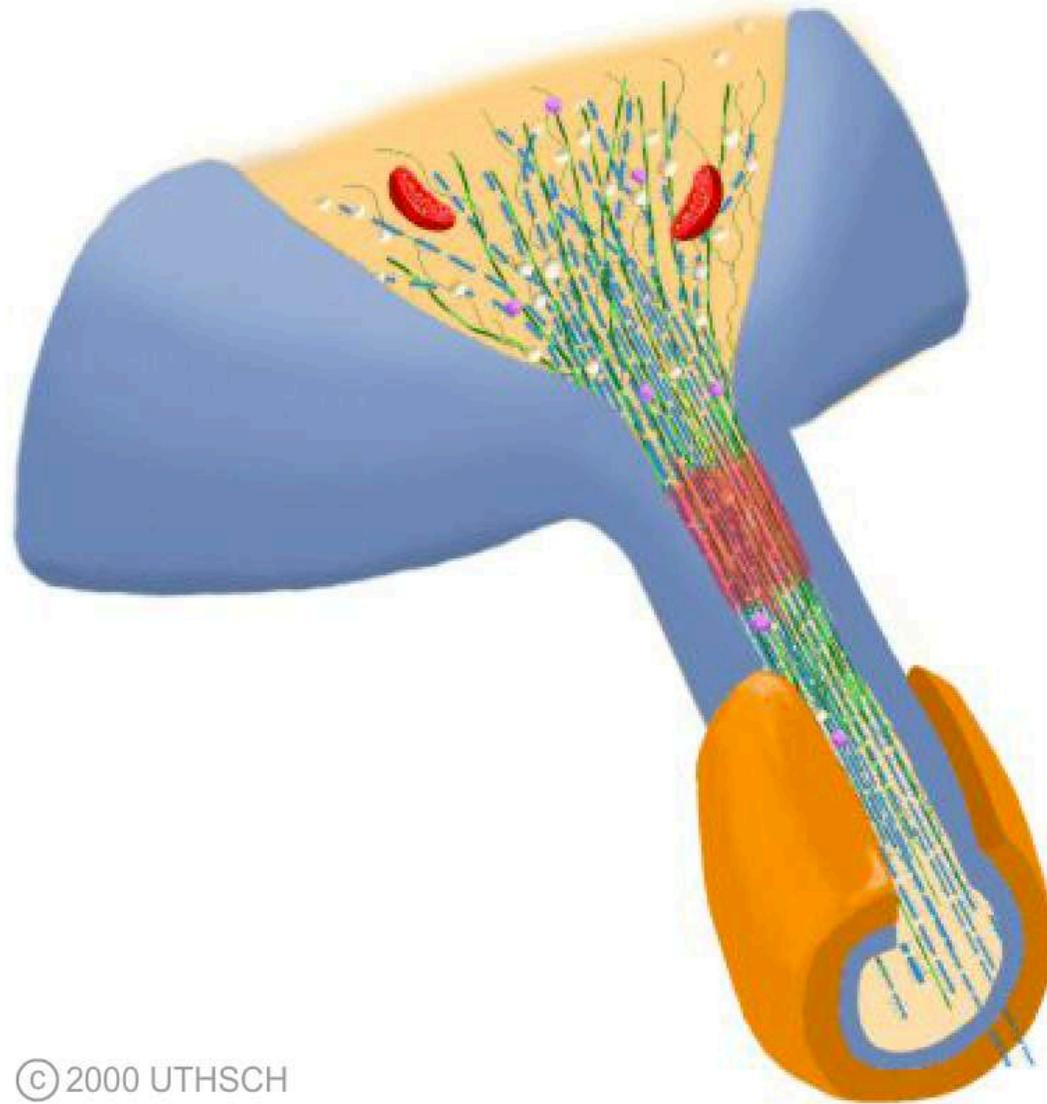




the movement of the secretory vesicles through the rough and the smooth ER. The smooth ER extends from the RER and serves as a site for lipid biosynthesis for the production of endosomes, lysosomes and plasma membrane as well as for the neurotransmitter vesicles. New membrane protein that begins its synthesis in the RER continues in the SER where pieces of the SER bud off to form transport vesicles that shuttle to the Golgi apparatus with their contents.



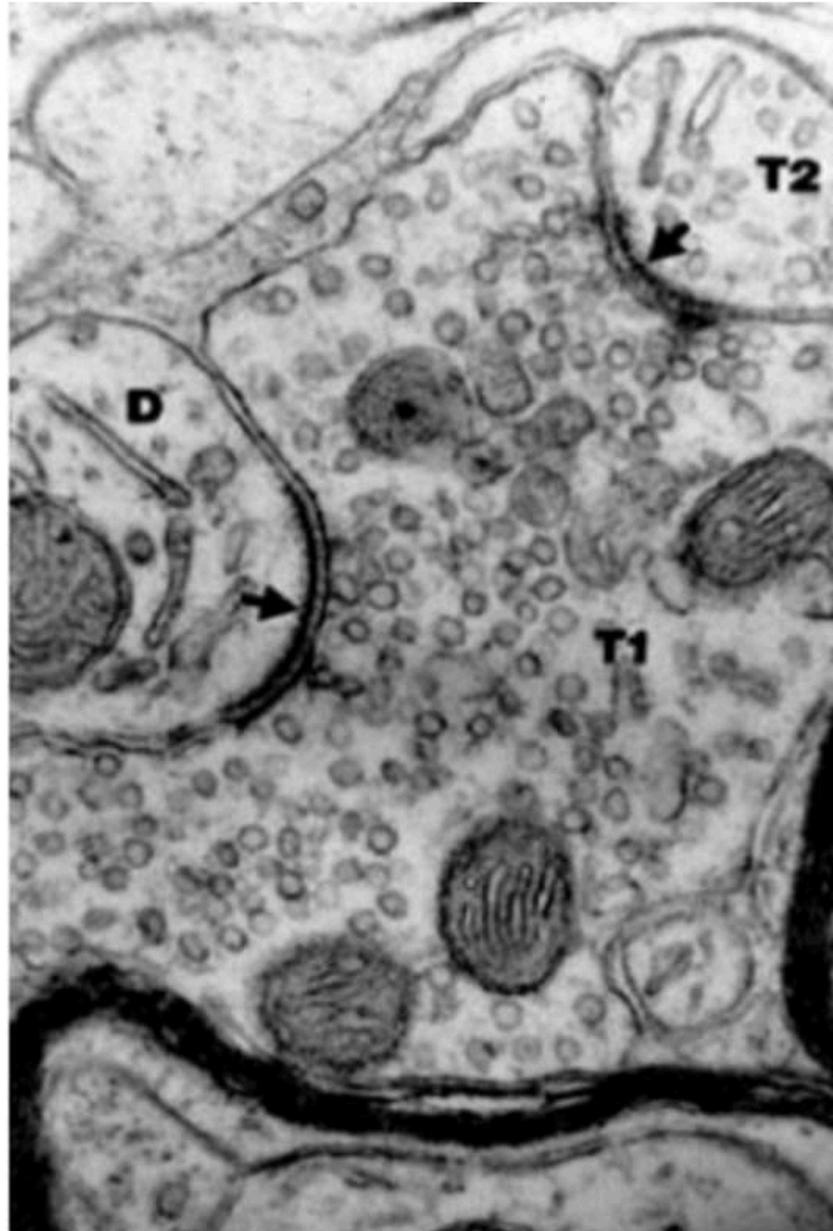




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**Figure 8.4 (See [enlarged view](#))**  
**Diagrammatic representation of the initial segment of a neuron, emphasizing the areas in which the action potential is initiated.**

# Синапс – электронная фотография

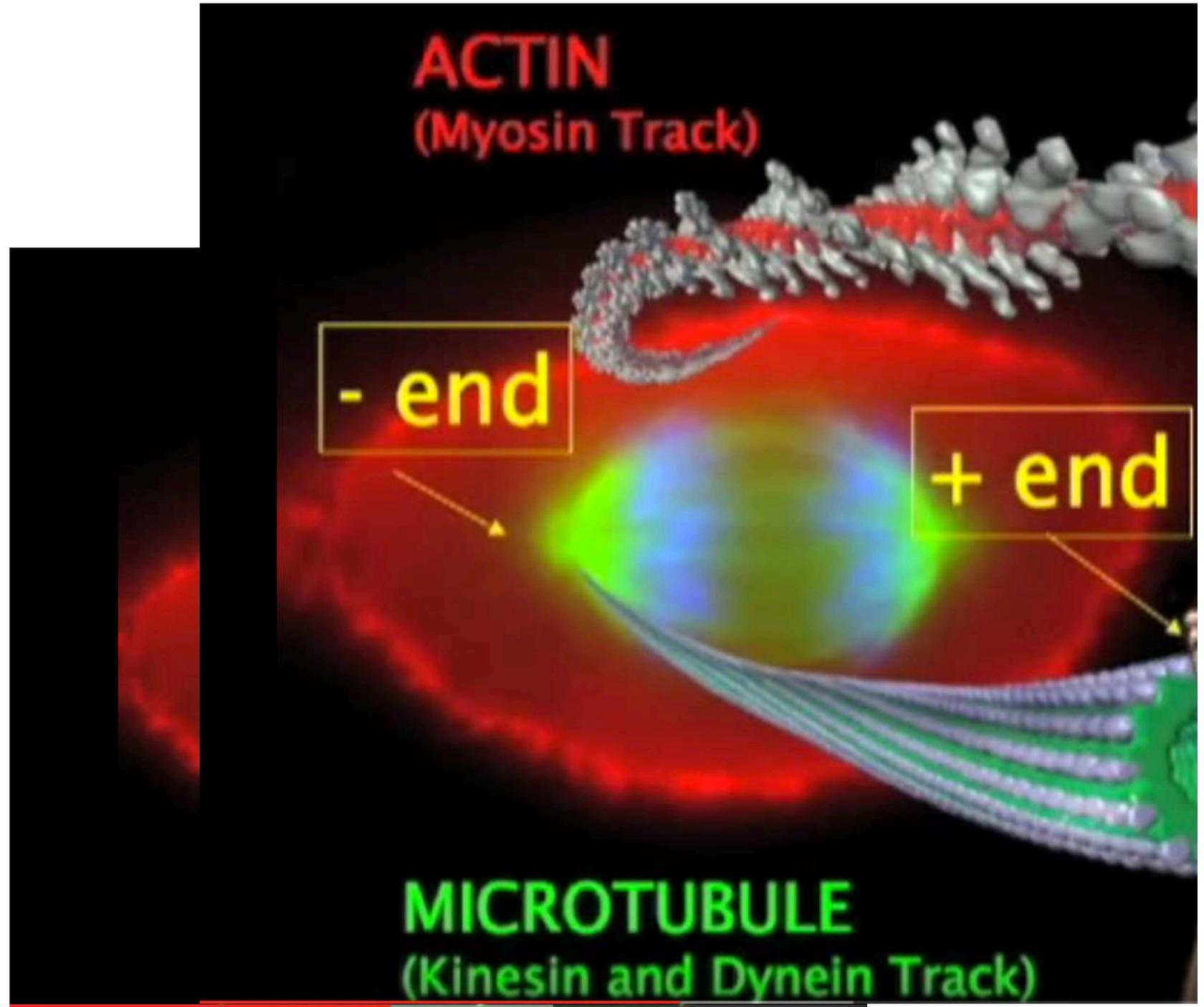


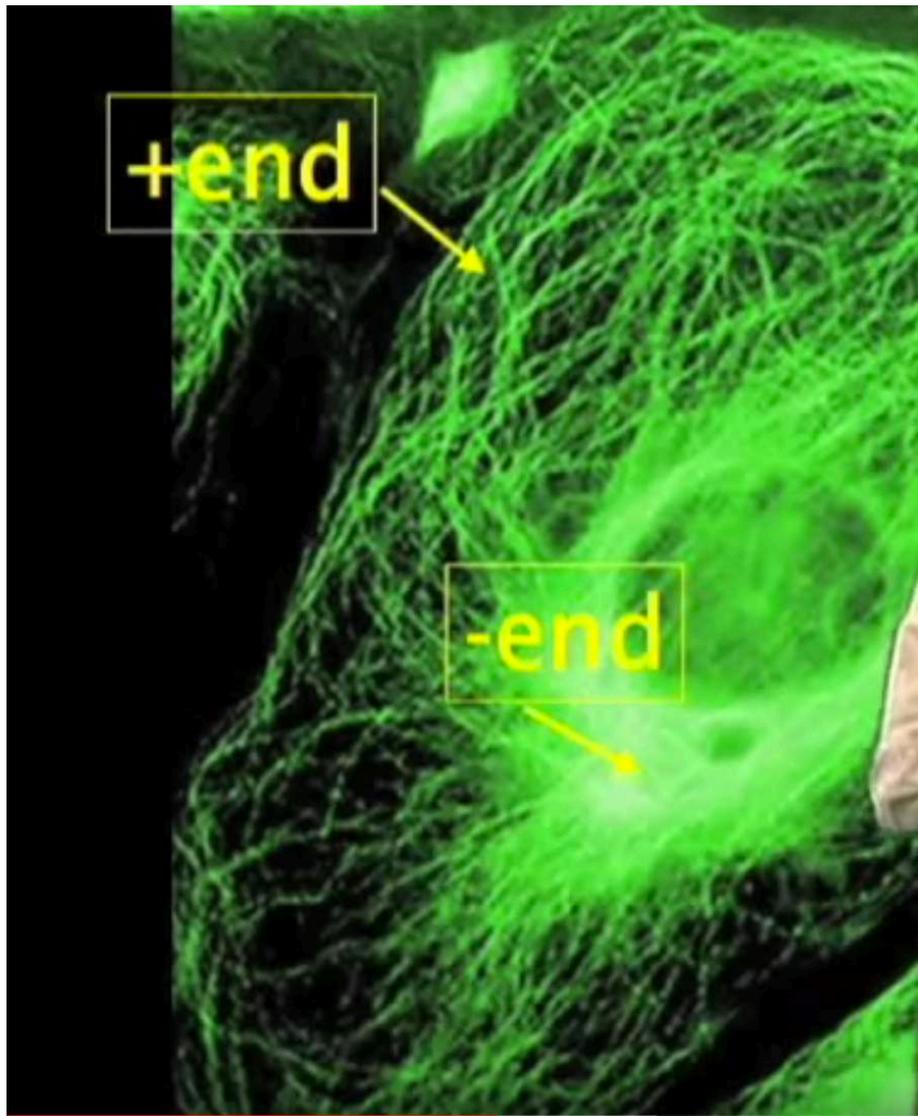
**ACTIN**  
(Myosin Track)

**- end**

**+ end**

**MICROTUBULE**  
(Kinesin and Dynein Track)

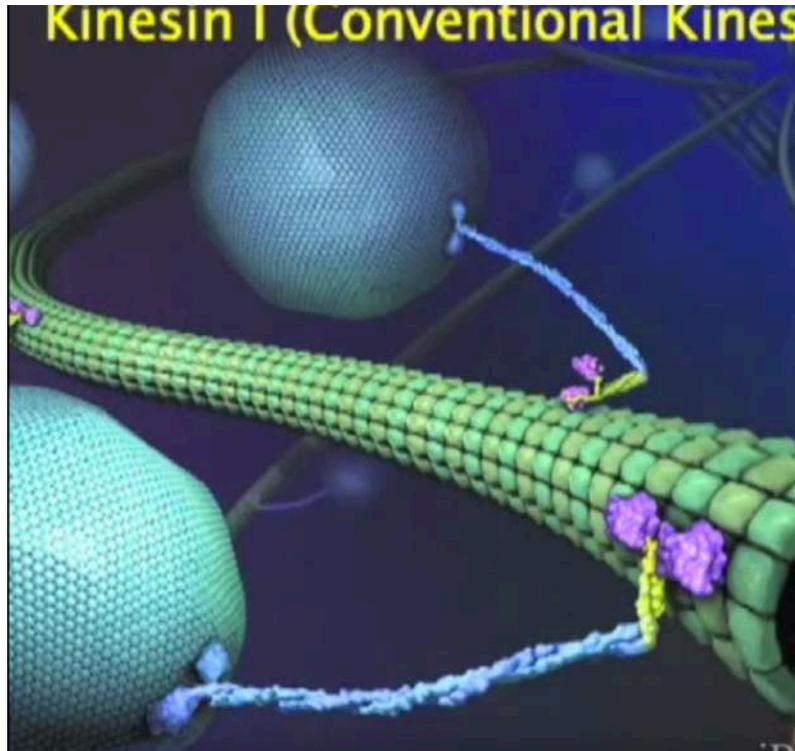




Моторные белки различают полярность и знают в каком направлении двигаться

Это – фантастически эффективная, сложная и надежно организованная система внутриклеточного транспорта

## Kinesin I (Conventional Kinesin)



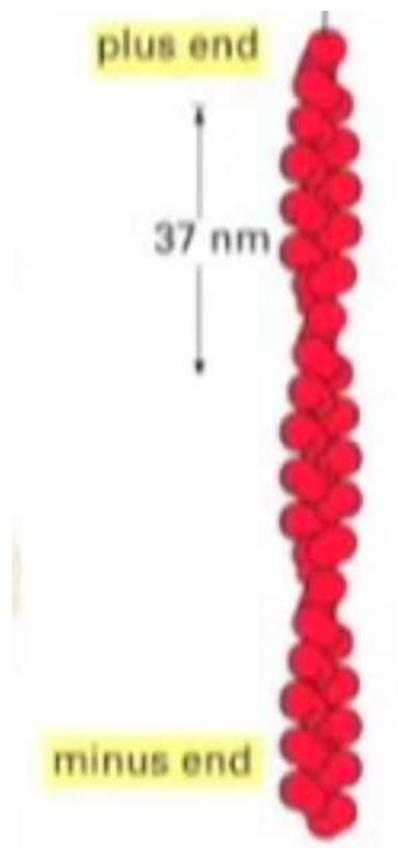
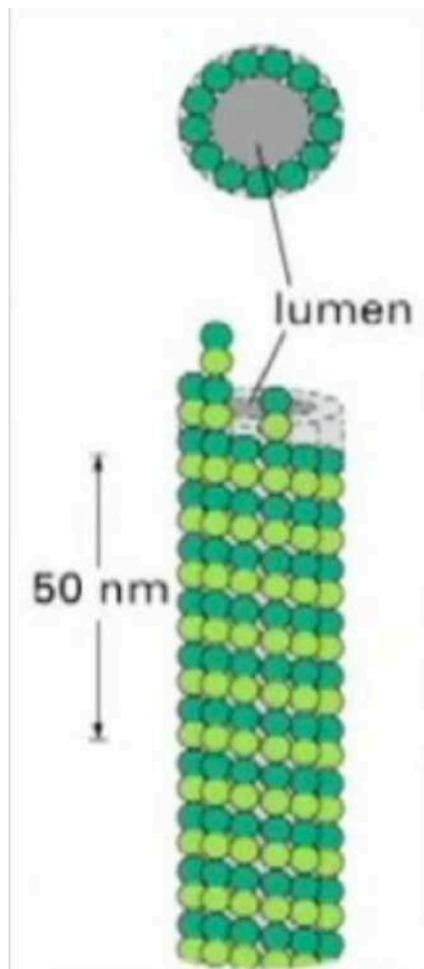
	<u>Kinesin</u>	<u>Automobile Engine</u>
<b>Size</b>	$10^{-8}$ m	1 m
<b>Fuel</b>	ATP	Hydrocarbons
<b>Speed</b>	$4 \times 10^{-3}$ m/hr $4 \times 10^5$ lengths/hr	$10^5$ m/hr $10^5$ lengths/hr
<b>Work Efficiency</b>	~60%	~10%

Relevance to medicine:

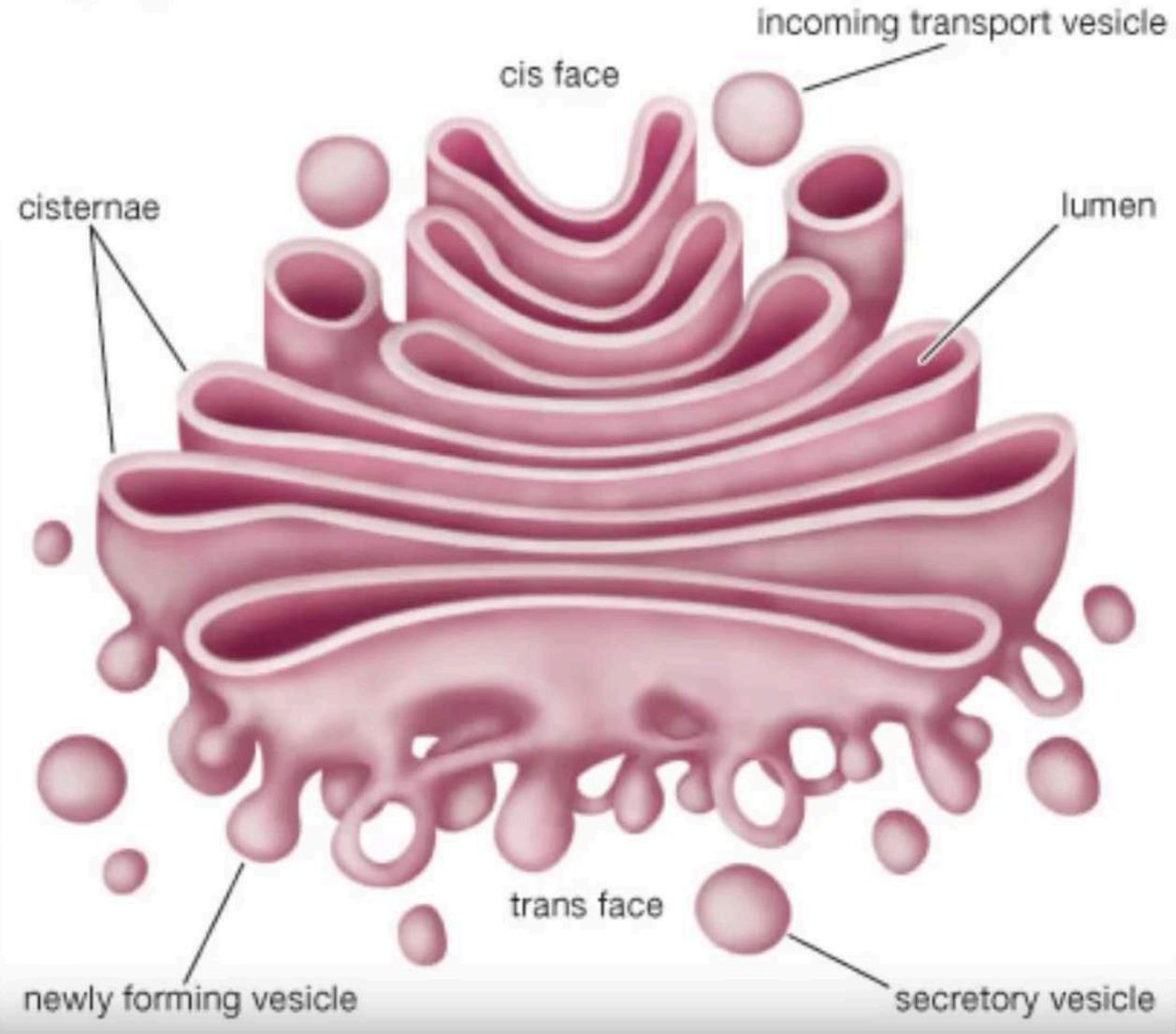
Transport defects can cause disease.

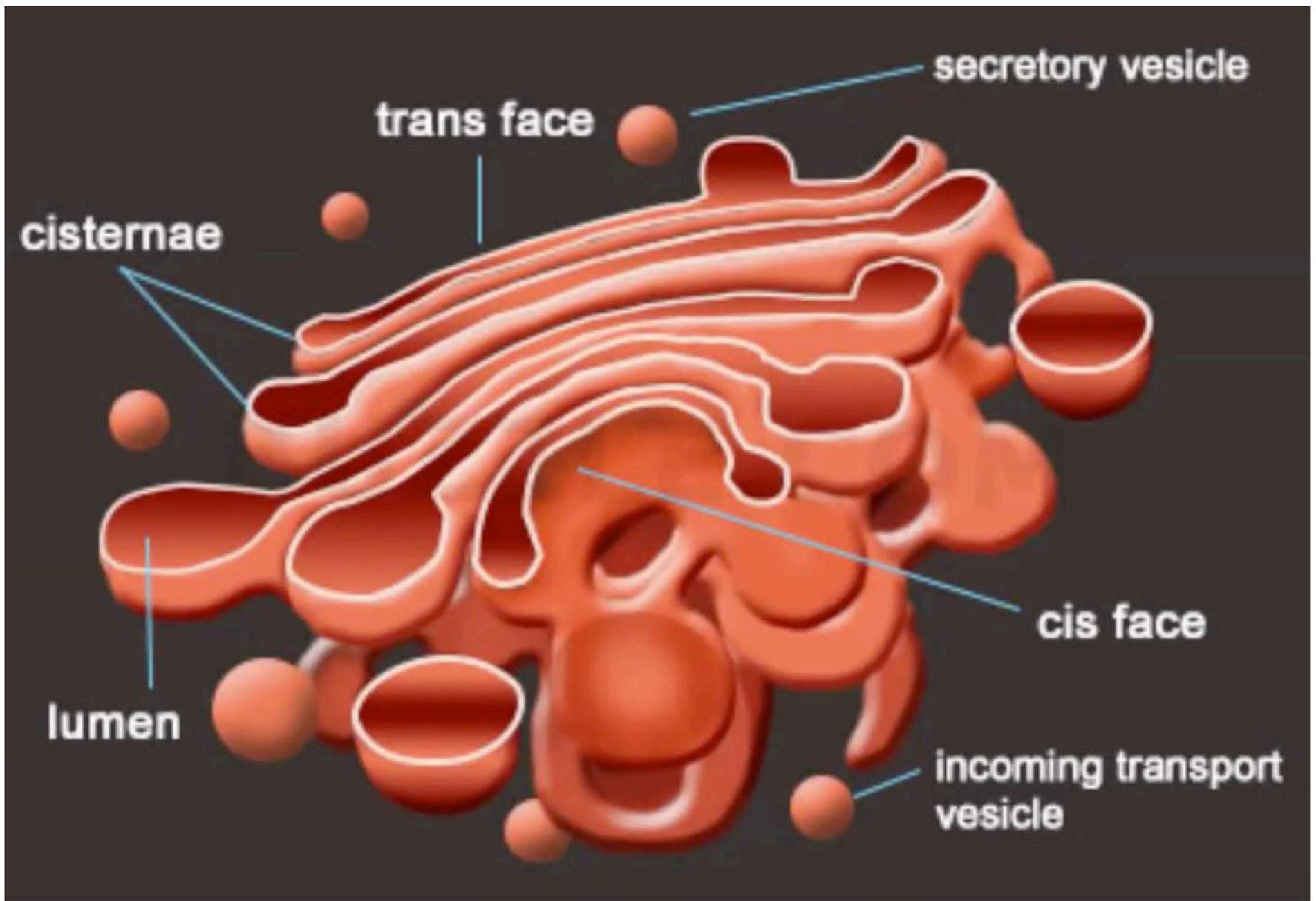
Inhibition or enhancement of motor protein activity may have therapeutic benefit.

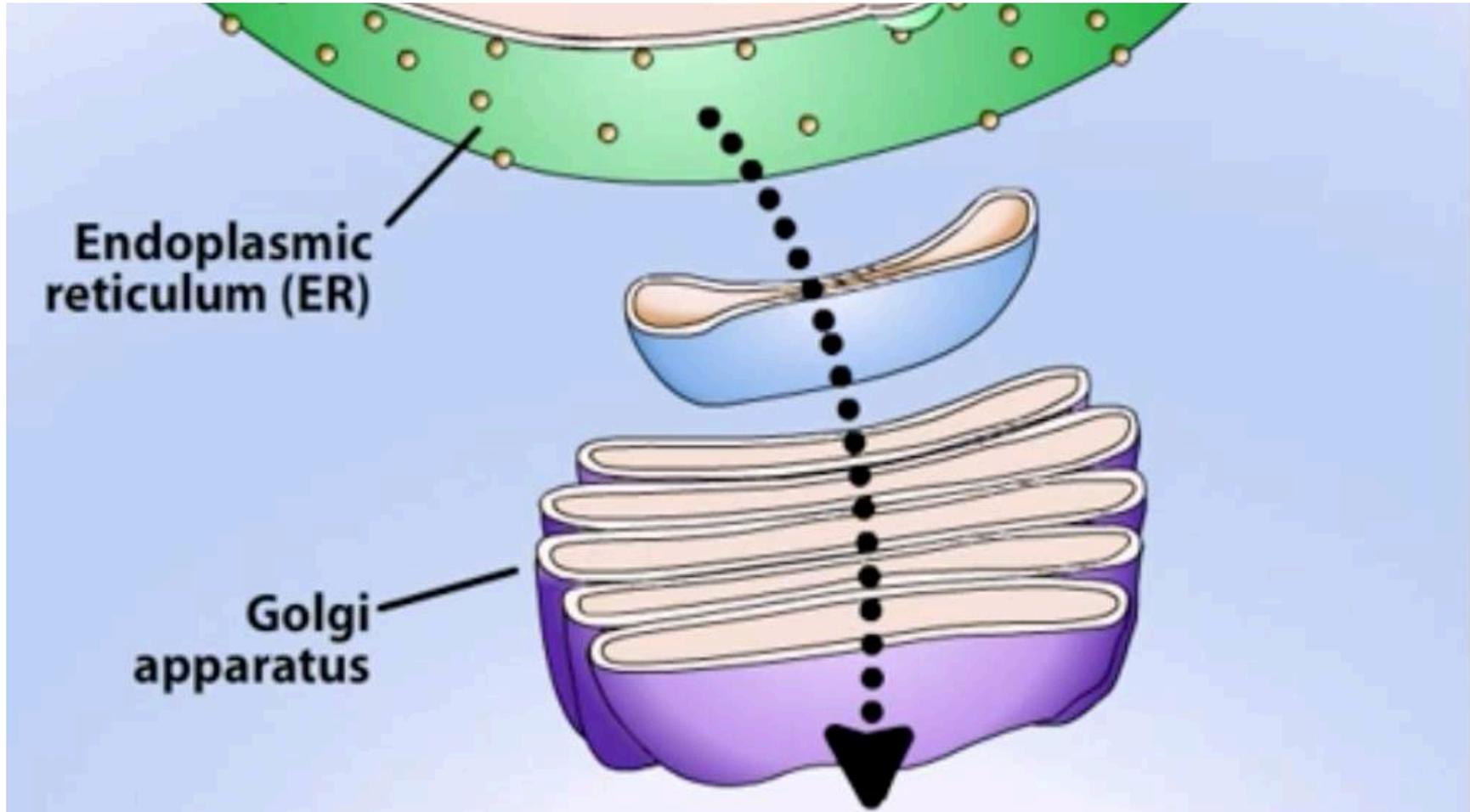
# Ключевые компоненты цитоплазматического транспорта



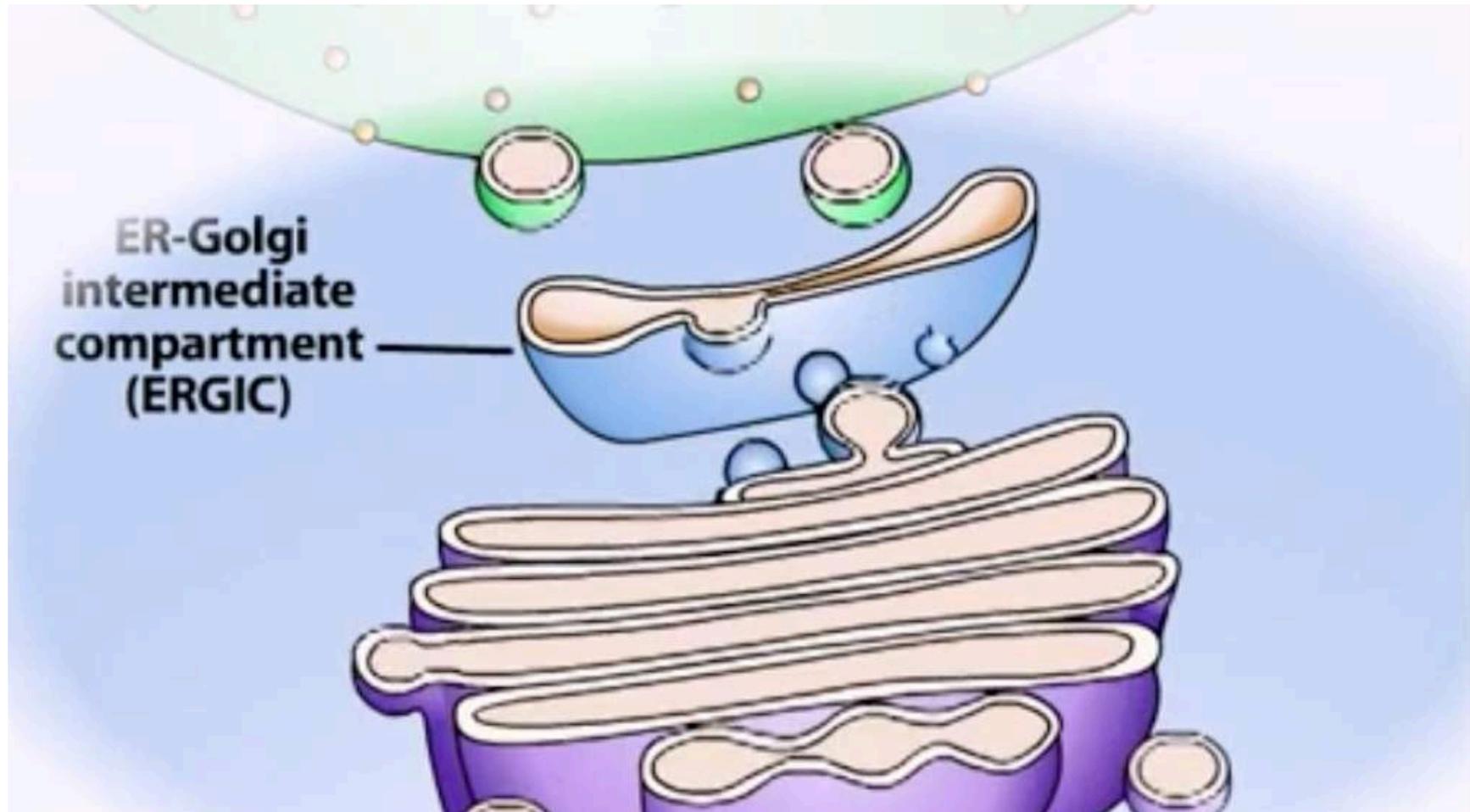
**Golgi apparatus**

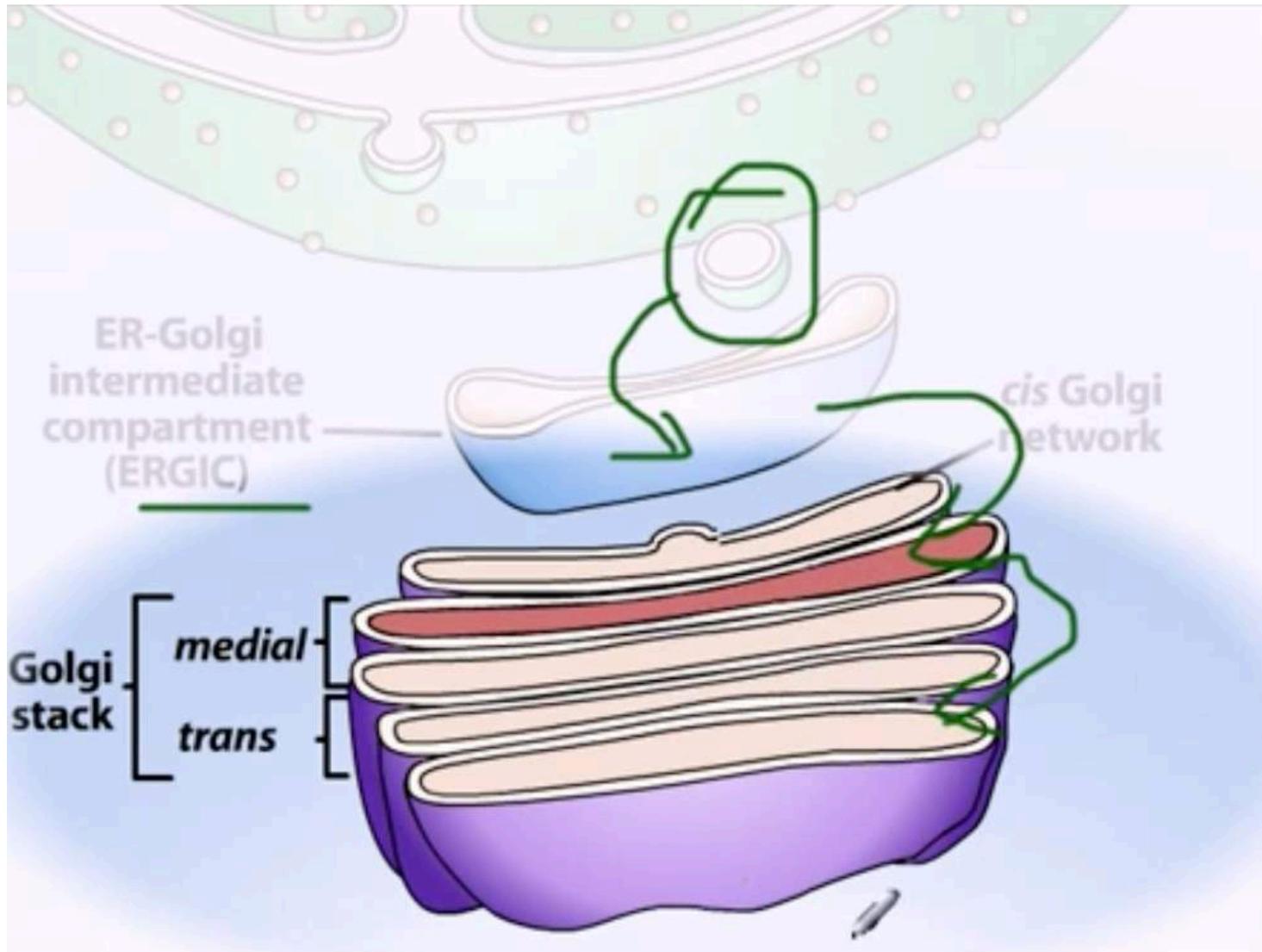






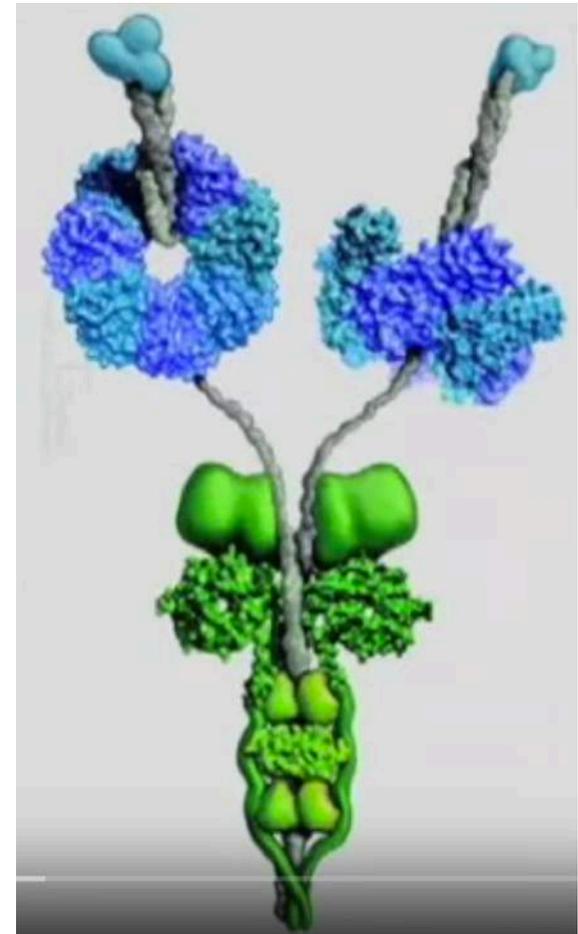
**ER-Golgi  
intermediate  
compartment  
(ERGIC)**





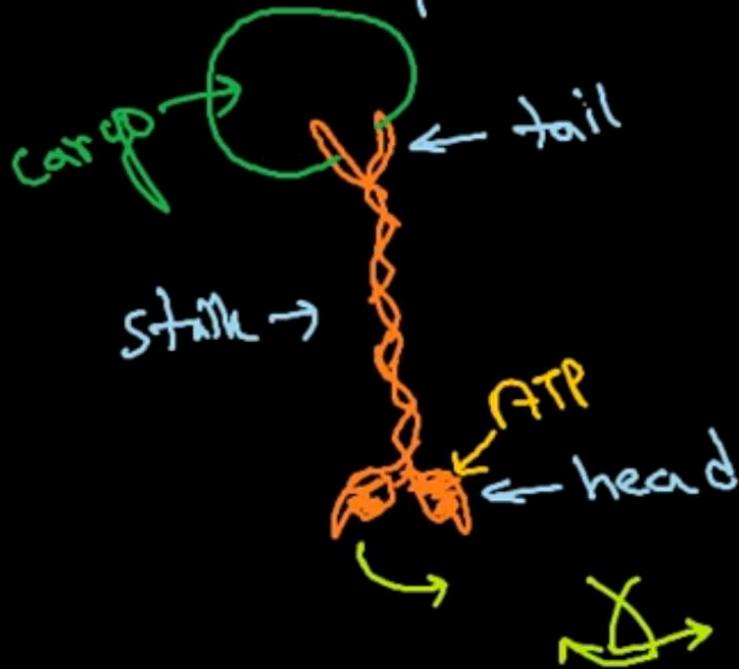


# Динеин – моторный белок ретроградного транспорта

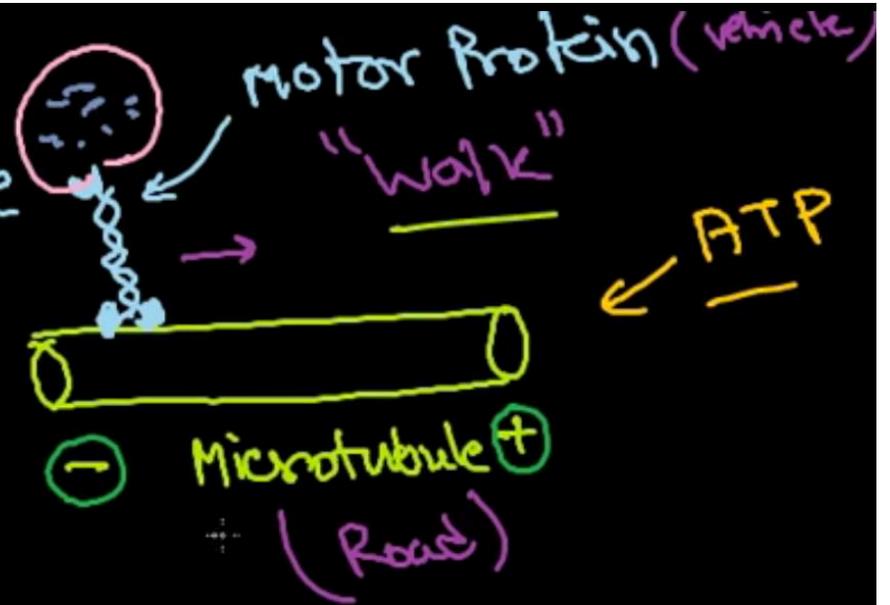


Microtubules → Road

Motor proteins → vehicle



②



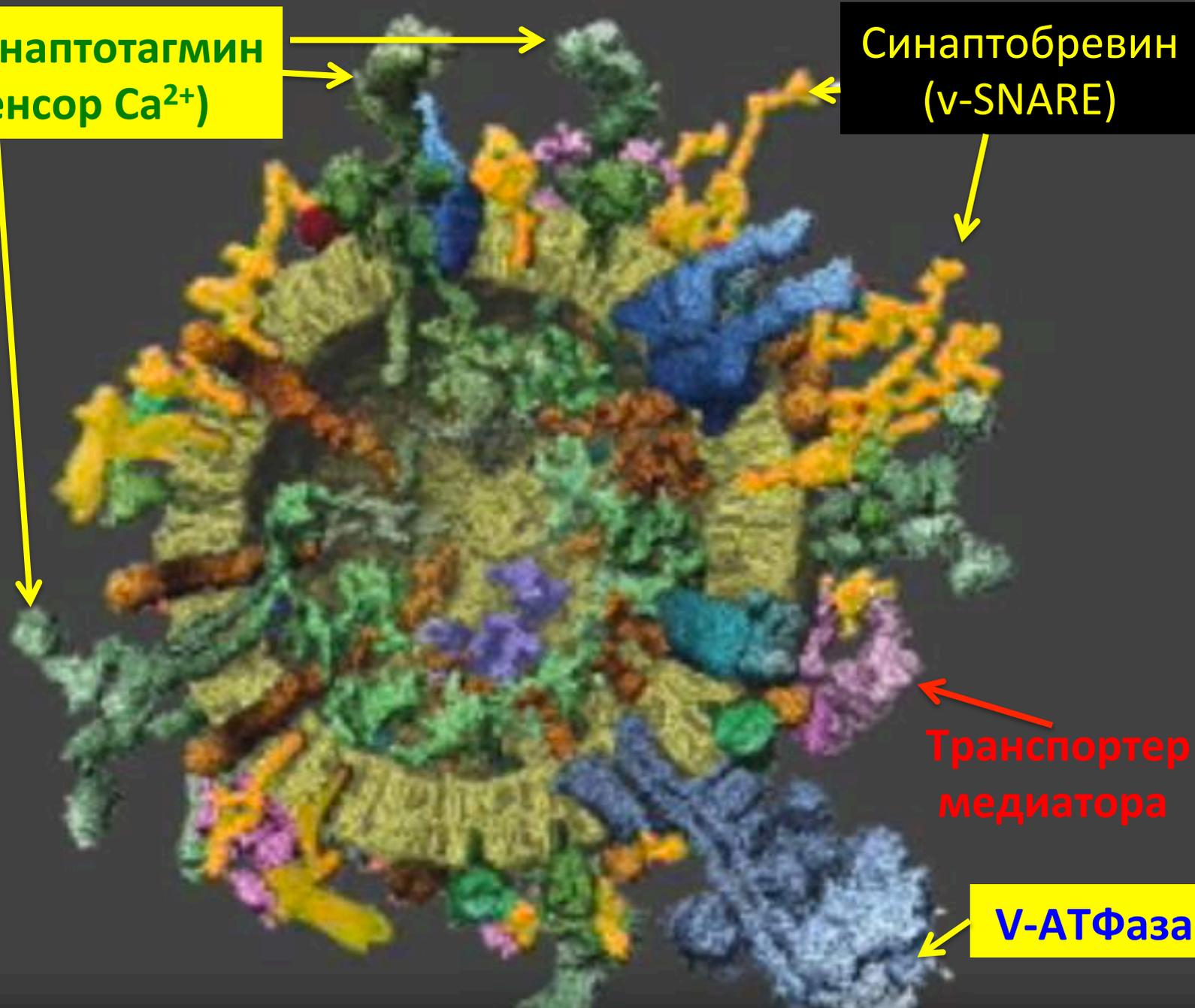
- (i) kinesin → ⊕
- (ii) Dyenin → ⊖

# Функции кинезинов

- Транспорт органелл
- Транспорт РНК и белков
- Сборка жгутиков и ресничек
- Формирование веретен деления при митозе и движение хромосом

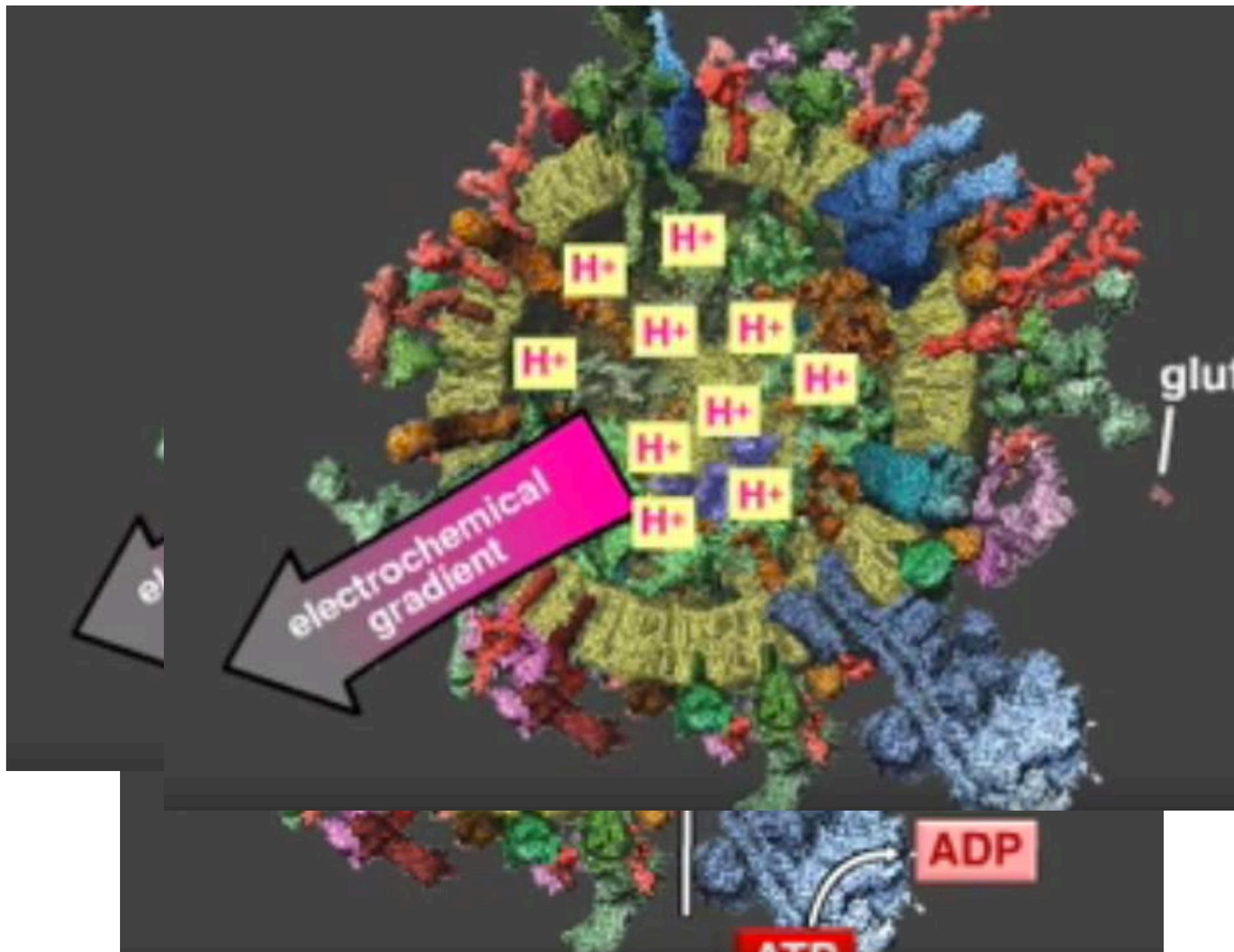
**Синаптотагмин  
(сенсор  $\text{Ca}^{2+}$ )**

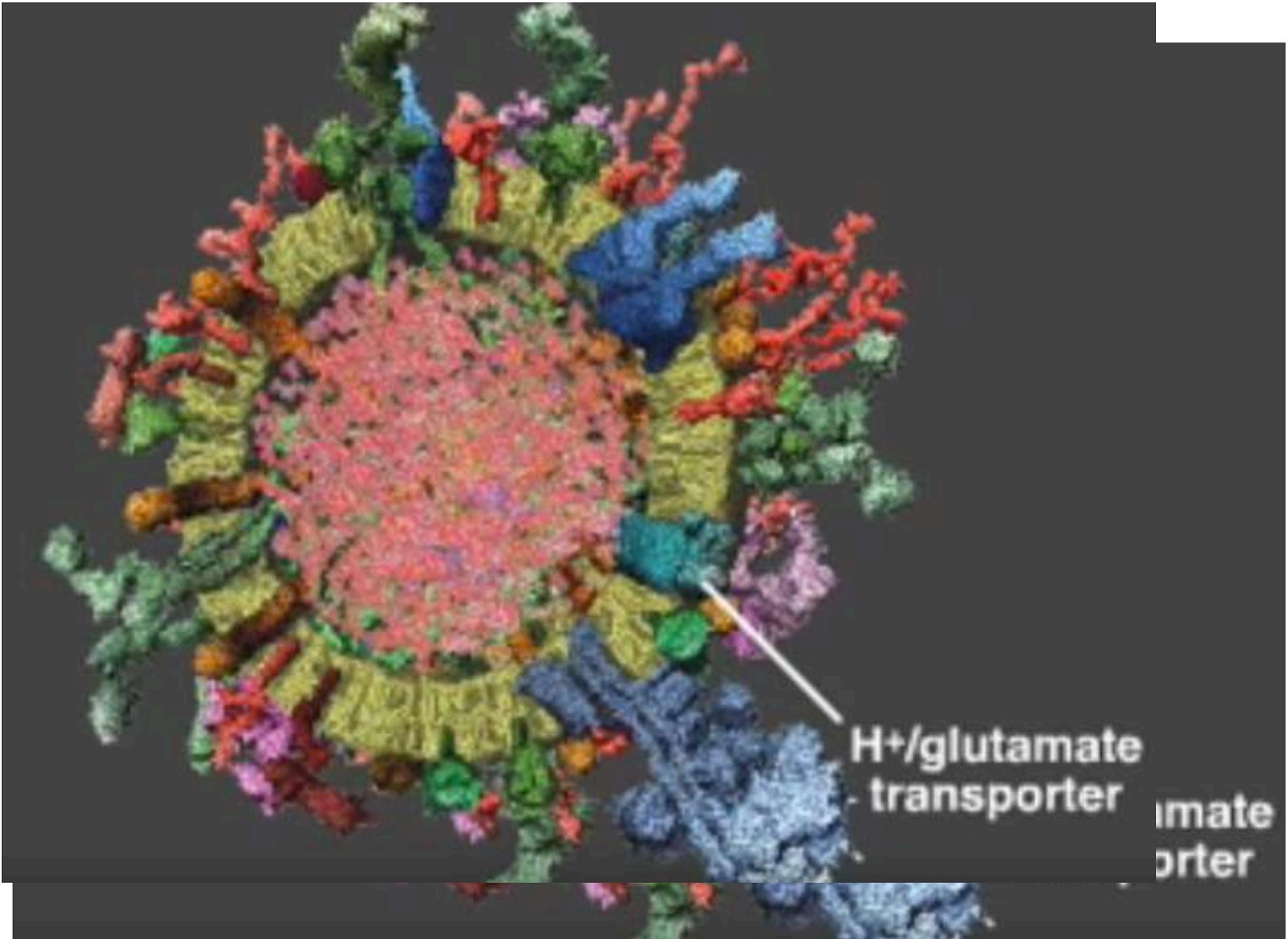
**Синаптобревин  
(v-SNARE)**

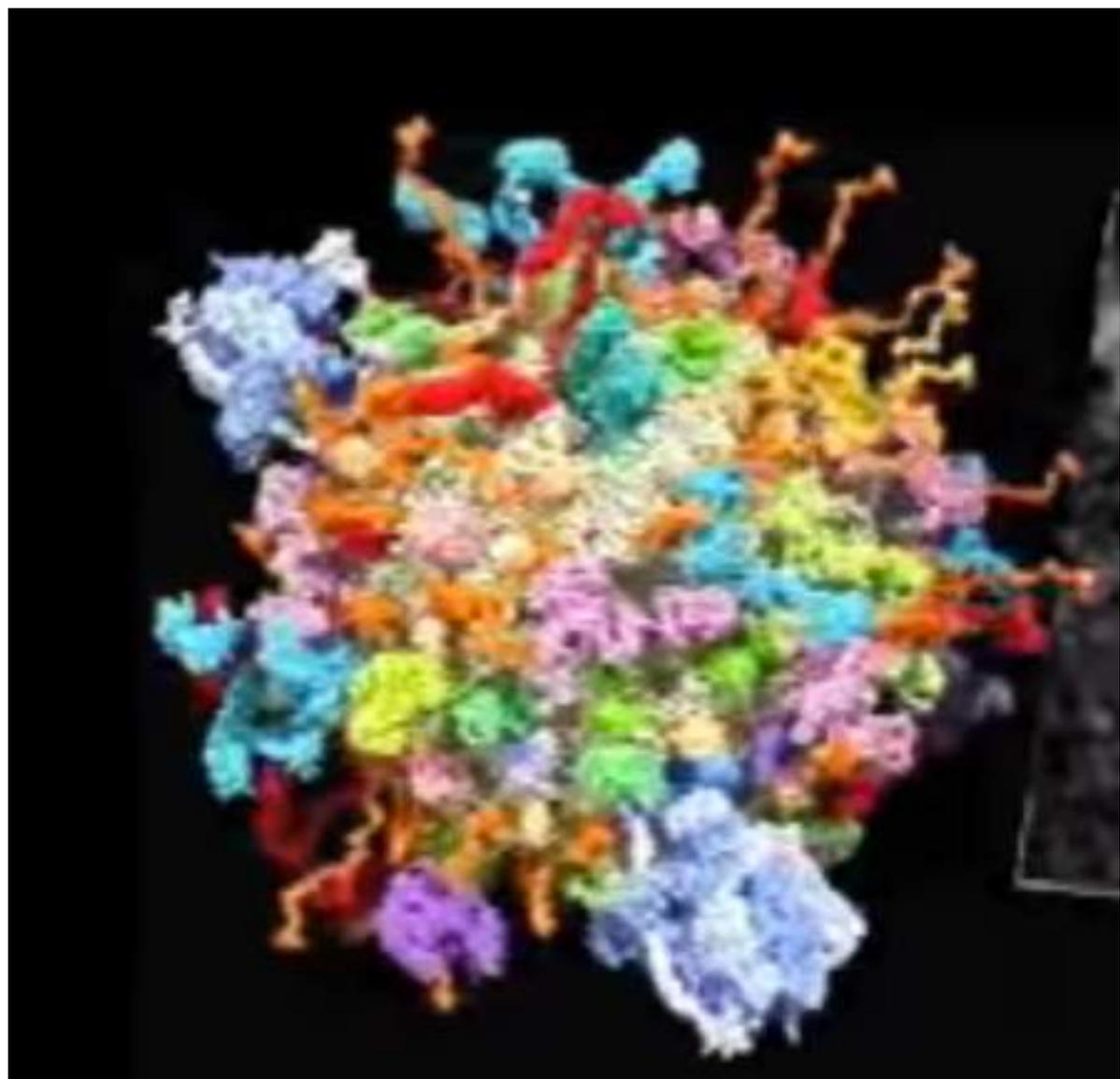


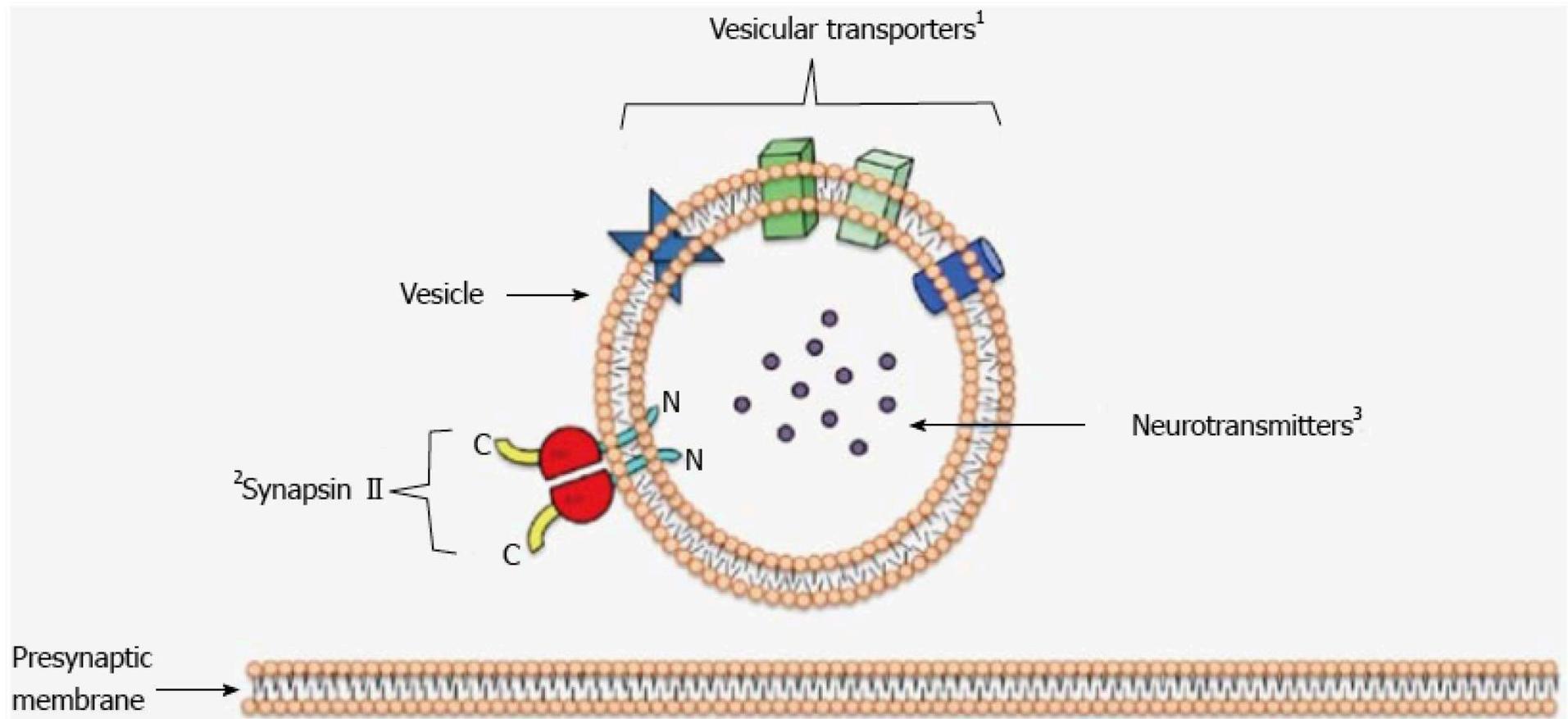
**Транспортер  
медиатора**

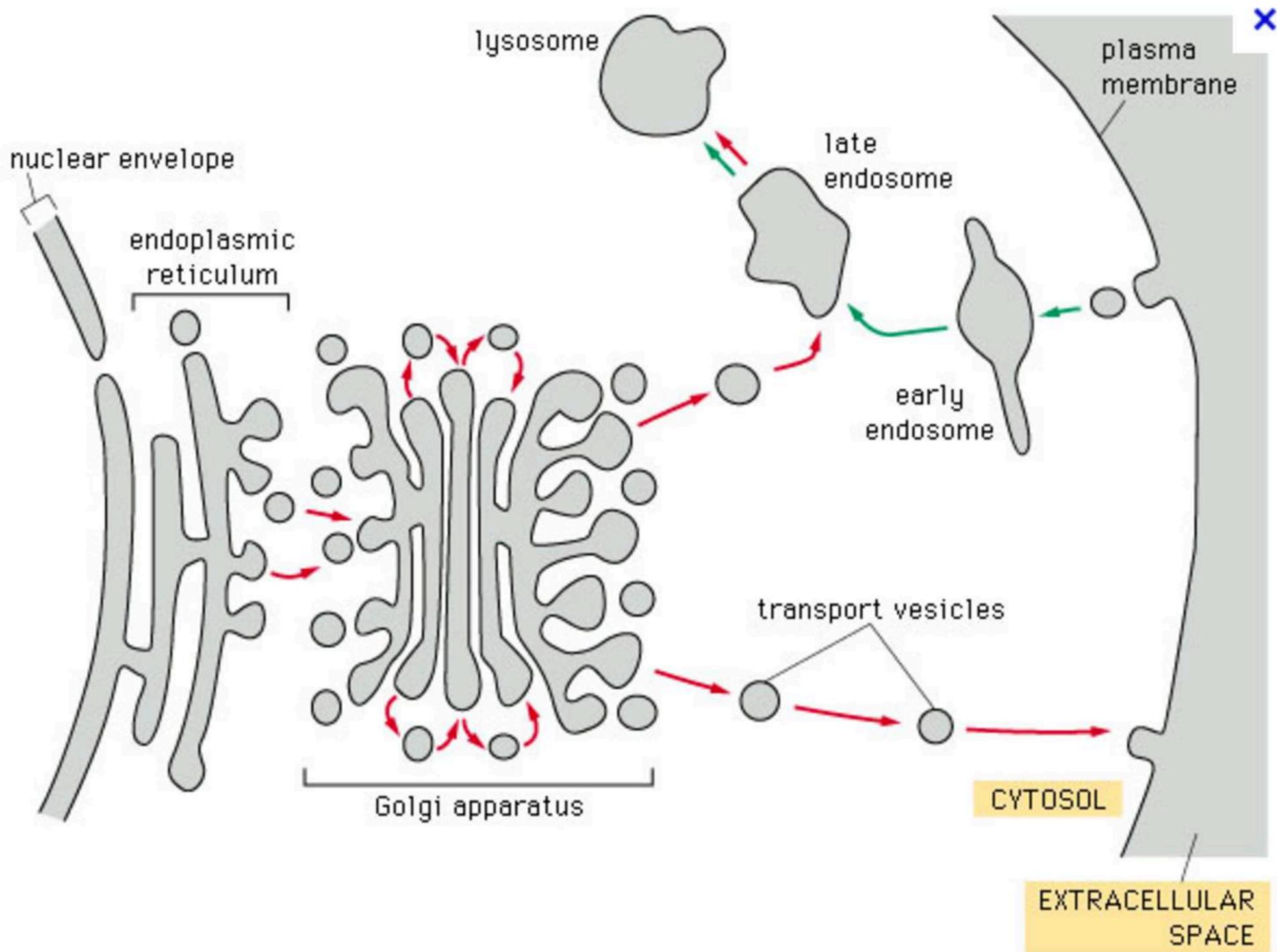
**V-АТФаза**



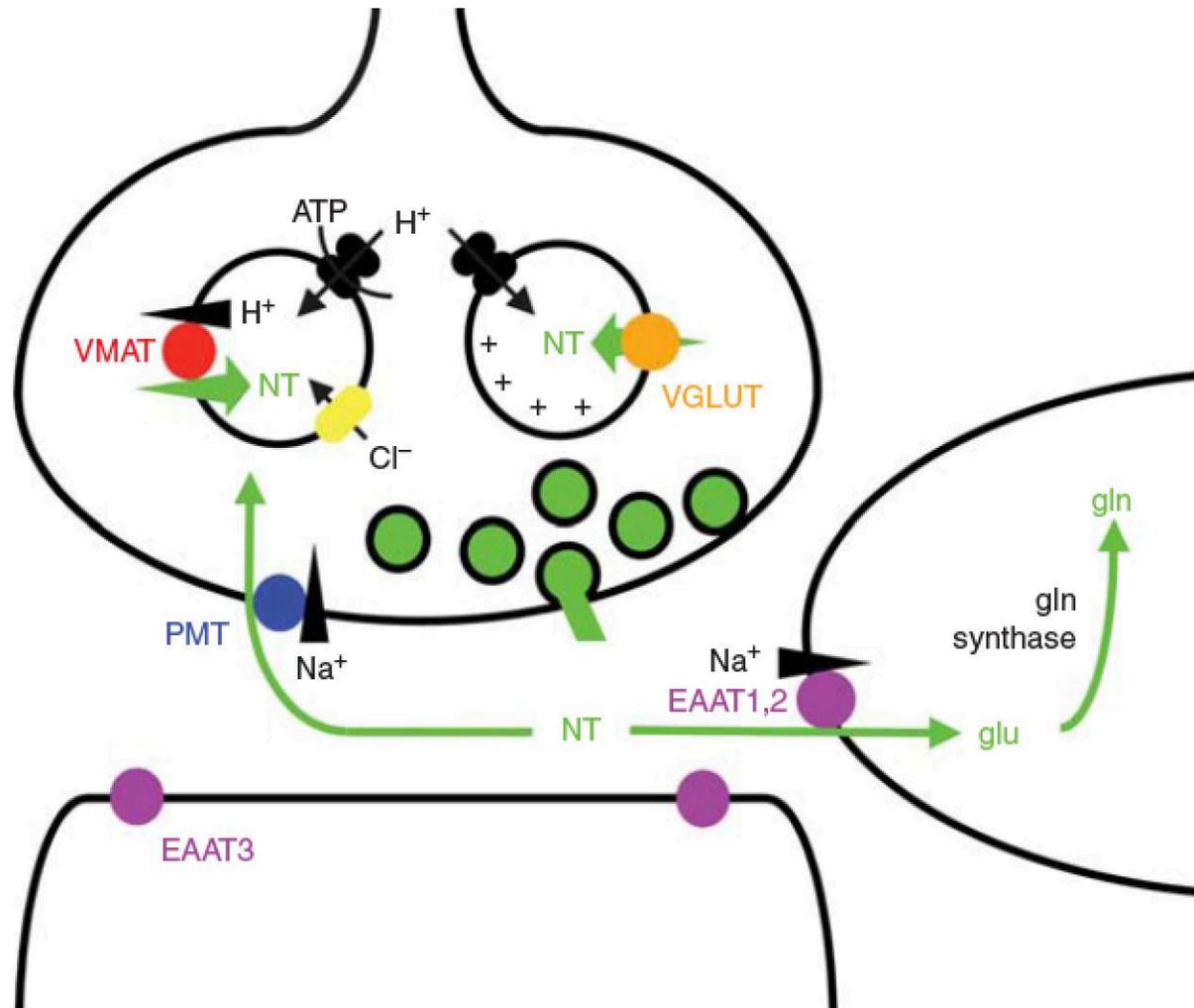






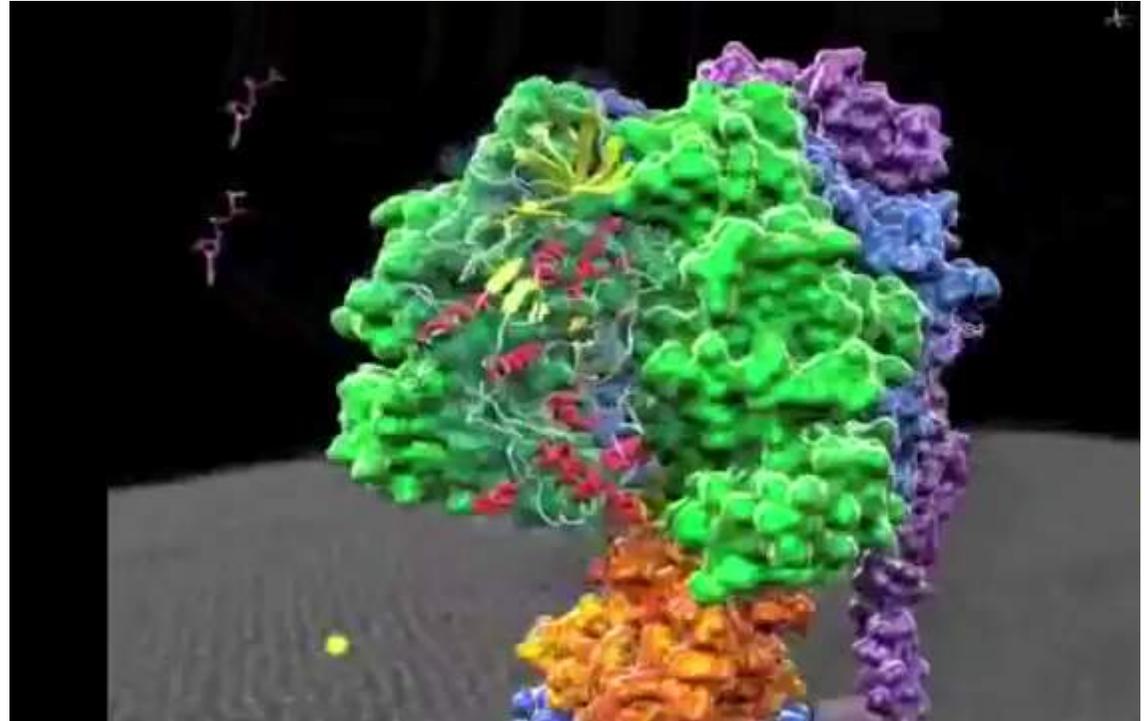
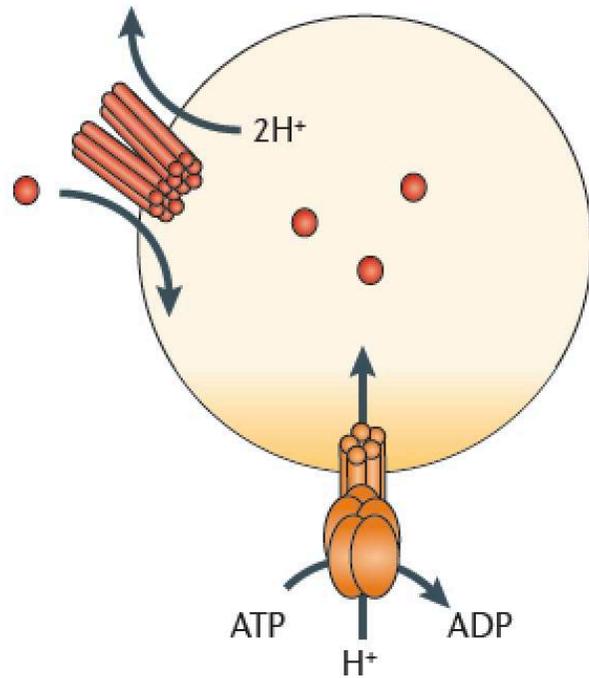


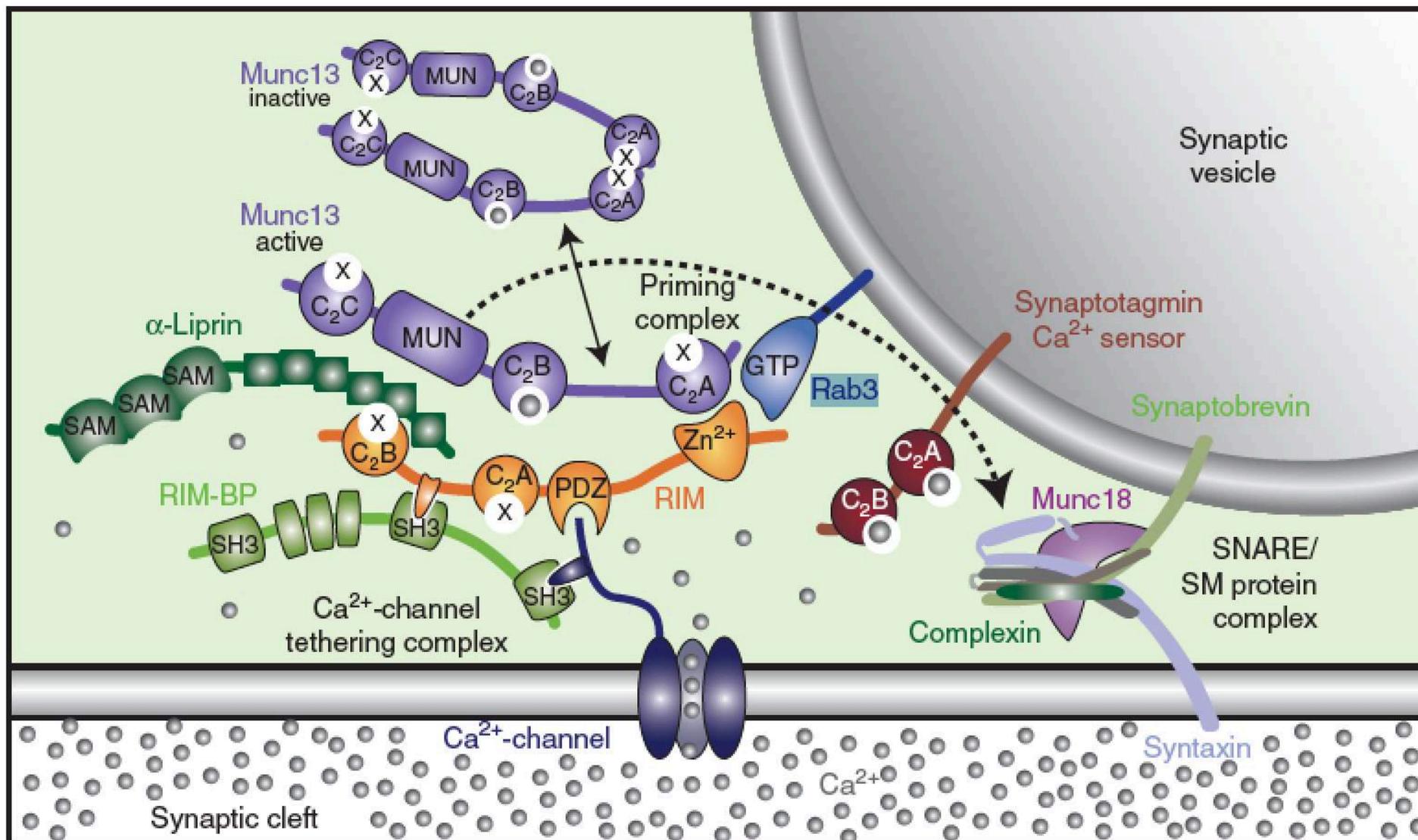
# Транспортеры нейромедиаторов

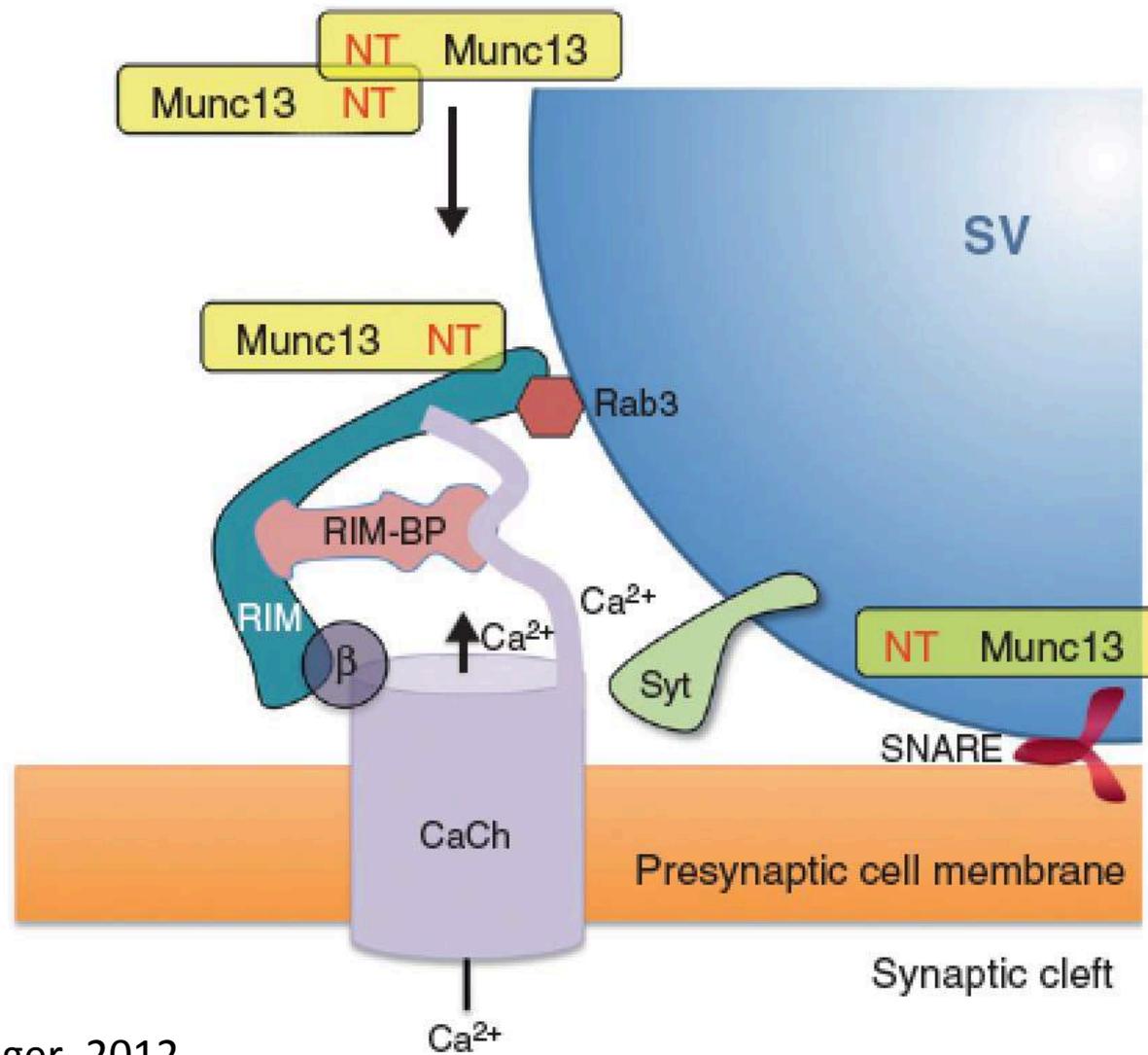


# Транспортеры нейромедиаторов

Acetylcholine vesicle







Gundelfinger, 2012

# Designer signals

- Преимущество использования Природой нейротрансмиттеров в том, что обеспечивается высокая **специфичность** работы синапсов
- Нейромедиаторы могут контролировать **возбуждение** и **торможение**
- Много лекарственных препаратов нервных расстройств действуют на синаптические белки

